

ANALYSIS OF CARBON DIOXIDE ABSORPTION IN
SOLUTIONS OF POTASSIUM SALTS OF GLYCINE AND
SARCOSINE

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by

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“L'espoir sera être réfracté pourvu que le cœur
reste transparent.”

ABSTRACT

CO₂ absorption in aqueous solutions of amino acid salts (AAS) involves simultaneous mass transfer with multiple reversible chemical reactions. The process is mathematically modeled by a set of nonlinear ordinary differential equations (ODEs) which cannot be solved analytically. However, near-exact solutions may be derived using the “Thick Film” linearization technique developed by K. A. Smith for application to the analysis of carrier-facilitated membrane transport [1, 2]. This technique is a variant of singular perturbation methods and is computationally efficient.

In this thesis, the Thick Film analysis is applied, in the context of the Film Model of mass transfer, to calculate rates of absorption of CO₂ in solutions of potassium glycinate (PG) and potassium sarcosinate (PS). Calculated enhancement factors, E , by which reaction multiplies absorption rates are in excellent agreement with values obtained via more computation-intensive numerical methods.

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1. INTRODUCTION

Anthropogenic emissions of the greenhouse gas, carbon dioxide, are responsible for some 60% global warming effects [3]. The total amount of carbon on earth, and in the atmosphere and lithosphere, was relatively constant before industrialization. However, the atmospheric CO₂ concentration increased dramatically from an estimated 282 ppmv in the year 1800 to 377 ppmv in the year 2004 [4]. Combustion of fossil fuels for electrical power generation is the primary source of CO₂ emissions.

Reducing these emissions (to stabilize atmospheric CO₂ levels) will require [5] a combination of more efficient use of energy, switching to using non-fossil fuels, and development of technologies to capture and sequester CO₂.

Conventional CO₂ capture techniques fall into essentially the three categories shown in Figure 1.1: post-combustion capture, pre-combustion capture and fuel combustion with oxygen-enriched air followed by capture of CO₂. CO₂ may be captured after combustion through a variety of separation processes including absorption, adsorption and membrane permeation [6, 7]. However, post-combustion exhaust gases typically contain low concentrations of CO₂ (4-14% v/v) [8, 9], which requires chemically reactive scrubbing solutions to capture it.

In pre-combustion capture, fuel is partially oxidized with oxygen (separated from air) and/or steam, to form a mixture of H₂ and CO₂. The CO₂ is separated from H₂, which is burned in the power plant. The higher CO₂ concentrations and pressures, compared to

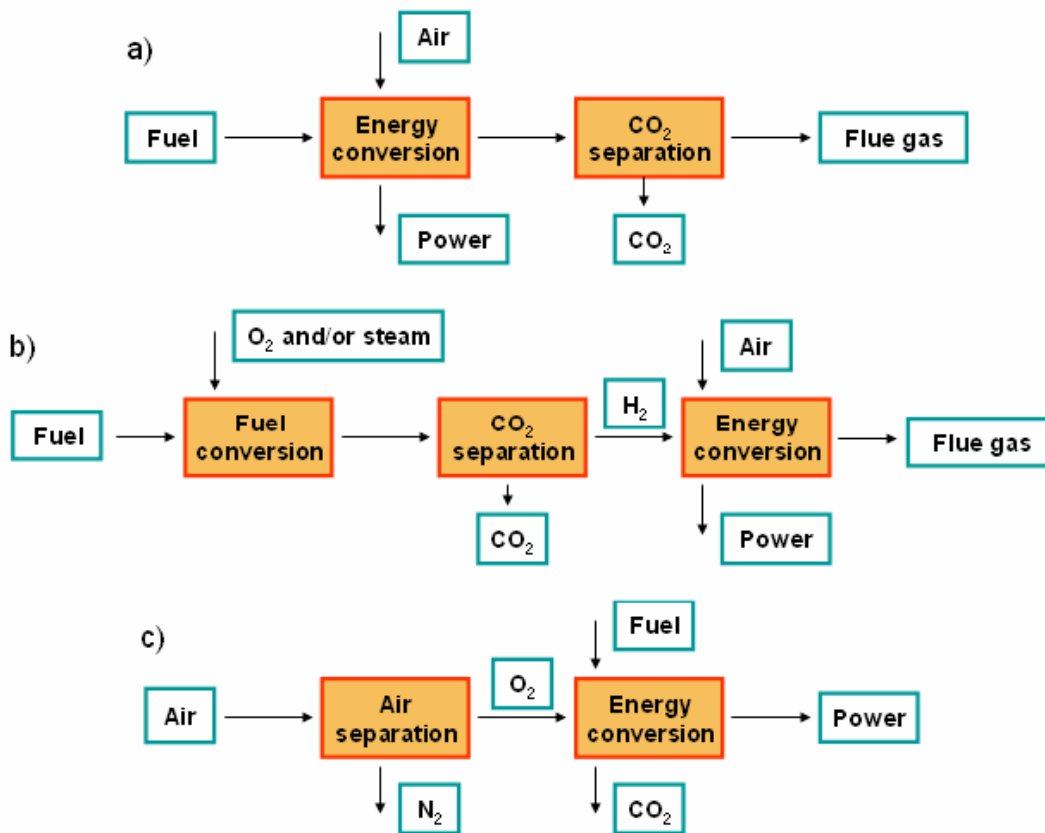


Figure 1.1 CO₂ capture schemes for fossil-fueled power plants [10]: (a) post-combustion system; (b) pre-combustion system; (c) oxy-fuel capture system.

post-combustion capture, reduce CO₂ capture operating costs, at the expenses of higher capital costs.[9]. Oxy-fuel combustion (i.e., with pure oxygen) produces CO₂ and H₂O at elevated temperatures and with negligible NO_x production at high temperature, because of the absence of N₂. The steam is separated by condensation [8, 11]. The advantages related to oxy-fuel capture are the high concentration of CO₂ in the output stream (above 80% v/v), and NO_x is not formed, while the main cost associated with oxy-fuel capture is the separation of O₂ from air.

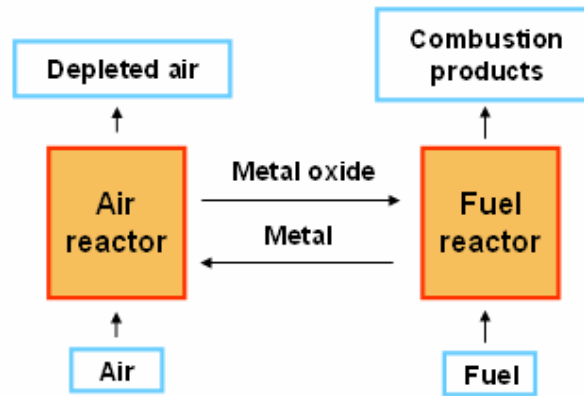


Figure 1.2 Chemical-looping combustion [12].

The novel chemical-looping combustion (CLC) process shown in Figure 1.2 includes internal CO_2 capture. Small particles of a metal oxide such as Fe_2O_3 , NiO , CuO or Mn_2O_3 carry oxygen between two separate reactors fed with air and fuel, respectively. This avoids the direct air/fuel contact. In one reactor, the oxygen-depleted carrier is oxidized by air. In the other reactor, the carrier is reduced by the fuel. The advantages of CLC are that the exhaust gas from oxidation reactor is mainly harmless N_2 ; and negligible NO_x is formed at moderate temperatures in the absence of a flame. The exhaust gas from the reduction reactor consists mainly of CO_2 and H_2O , which is separated by condensation.

Regardless of the source of CO_2 , the technologies for its capture include both physical and chemical processes, and combinations thereof. The focus here is on absorption.

2. REVIEW OF LITERATURE

2.1 CO₂ absorption technology

The flue gas CO₂ capture systems of greatest current interest involve absorption and desorption (stripping) using solvents containing alkanolamines, amino acid salts, ammonia and ionic liquids, as well as carbonation/calcination cycles [13-42].

Most scrubbing solutions contain alkanolamines – e.g., monoethanolamine (MEA), diethanolamine (DEA), di-isopropanolamine (DIPA) and methyldiethanolamine (MDEA). MEA is particularly well-suited to remove CO₂ from flue gases [13]. In the regenerative process, MEA reacts in solution with CO₂ to form MEA carbamate; the CO₂-rich MEA solution is fed to a stripper where concentrated CO₂ is released by contact with steam; the regenerated (stripped) MEA solution is sent to the absorber [14].

This process is capital and energy-intensive [15-17]. The steam supplied to the stripper, which operates at 110 ~ 130 °C, is responsible for up to 70% of the operating costs of CO₂ capture [18]. Reactions of the amine with flue gas SO₂, NO₂, HCl, HF and oxygen forms corrosive degradation products, which requires construction with stainless steel. Other drawbacks of MEA-based processes are relatively low CO₂ loading capacity and amine volatility.

Figure 2.1 shows the reaction pathways between CO₂ and primary or secondary amines. Most of the CO₂ absorbed at relatively low temperature (*ca.* 40 °C) forms carbamate and protonated alkanolamine. It can be calculated from the reaction scheme

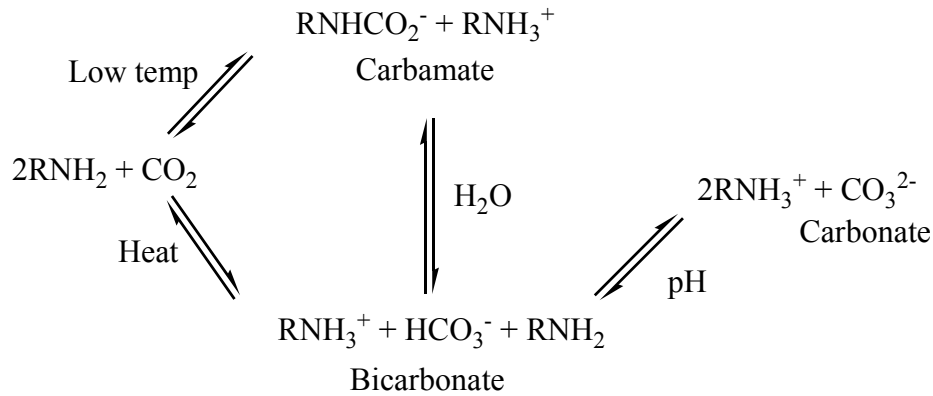
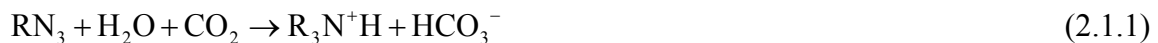


Figure 2.1 Proposed mechanism of reactions between CO_2 and primary or secondary amines [19].

that, there is a theoretical maximum CO_2 loading of 0.5 mol CO_2 /mol amine in the absence of hydrolysis of the carbamate [19]. The reaction between amine and CO_2 is very fast in comparison to the carbamate hydrolysis reaction [20]; the correspondingly high heat of carbamate formation results in high solvent regeneration costs. Depending on the residence time of the liquid in the gas-liquid contactor, some degree of carbamate hydrolysis may occur, which results in maximum CO_2 loading values larger than 0.5. Also depending on the temperature and pH value of the reaction system, product proportion of bicarbonate and carbonate vary with the amine, temperature and pH.

Unlike primary and secondary amines, tertiary amines have no hydrogen atom attached to the nitrogen atom and, therefore, form no carbamate. However, tertiary amines facilitate CO_2 hydrolysis, forming bicarbonate via a base-catalyzed hydration [21]:



The reaction heat released in bicarbonate formation is lower than that of carbamate

formation, thus reducing solvent regeneration costs. As can be seen from equation 2.1.1, tertiary amines have a high CO₂ loading capacity of 1 mol CO₂/mol amine. In addition, rates of CO₂ absorption in solutions of tertiary amines can be enhanced by the addition of small amounts of primary or secondary amines [22].

Mixed amines can combine the desirable qualities of the individual amines. Mixtures of tertiary amines with primary amines or secondary amines offer the CO₂ reactivity of primary or secondary amines *and* higher CO₂ capacities – therefore lower required solvent circulation rates and regeneration costs similar to those of tertiary amines [18].

Not long ago, a new class of amines - *sterically hindered amines* - with regeneration costs lower than those of conventional primary and secondary amines, received considerable attention. A sterically hindered amine is a primary amine whose amino group is attached to a tertiary carbon atom, or a secondary amine whose amino group is attached to a secondary or tertiary carbon atom [23]. With a large group attached to the nitrogen atom, these amines form unstable carbamates that hydrolyze to form bicarbonate, thus resulting in a CO₂ capacity of 1 mol CO₂/mol amine [24-26].

Because of the use of alkanolamines for CO₂ absorption is capital and operating expense intensive, development of more efficient solvents is essential for reducing the cost of CO₂ capture. Amino acid salts (AAS), due to their physical and chemical properties, are promising, albeit more expensive, alternatives to alkanolamines. An AAS is formed by neutralizing a weak organic nonvolatile amino acid with an equinormal

amount of strong inorganic base. For example, potassium glycinate (aminoacetic acid, denoted as PG) is the neutral product of glycine and potassium hydroxide.

AAS's are less subject to degradation (less production of toxic degradation products) when exposed to flue gases [27]. Their ionic nature results in negligible volatility and increased surface tension [28]. Additionally, many amino acids are naturally occurring so that they have less environmental or toxicity issues. Their CO₂ absorption capabilities are comparable to those of alkanolamines [29]. Under certain operating conditions, solid precipitates composed of amino acids and more complex, CO₂-containing species have been observed.

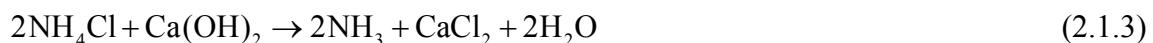
Furthermore, amino acids can be neutralized, not only by strong inorganic bases, but also by organic bases including amines. The latter mixtures are called amine amino acid salts (AAAS). Neutralization is only partial because of the weak reaction between weak acid and weak base. AAAS's show higher CO₂ loading, higher cyclic capacity and absorb larger amount of CO₂ than AAS's at the same concentration. The explanation is not entirely clear; it may be related to a lower equilibrium temperature sensitivity of AAAS's [30]. It was also observed that AAAS showed no foaming, discoloration or precipitation under atmospheric conditions up to 80 °C [30], demonstrating good signs of stability.

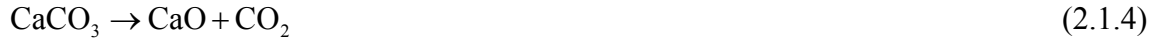
Ammonia is yet another alternative that avoid the drawbacks of MEA [16, 17]. Aqueous ammonia absorbs CO₂ to form ammonium bicarbonate and ammonium carbonate. Before reacting with aqueous ammonia in a wet scrubber at low temperature

(0 ~ 20 °C), flue gas needs to be pretreated by oxidizing SO₂ and NO to SO₃ and NO₂. In this temperature range, ammonium carbonate and bicarbonate precipitate in the absorber. The ammonia is regenerated simply by thermal decomposition of the products at high temperatures (100 ~ 200 °C) [31]. Due to the lower heat of desorption of CO₂, this process reportedly requires 60% less energy input than a process based on MEA. Major byproducts are common fertilizers.

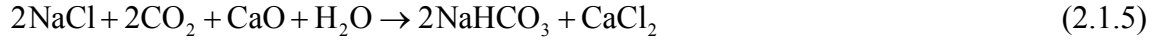
Although this process potentially allows the burning of cheap and sulfur abundant coals, it also produces SO₃ and NO₂ in oxidizing flue gas pretreatment. Another problem is that the absorber off-gas phase contains ammonia, the emission of which must be avoided.

The dual-alkali approach employs ammonia as the primary alkali to promote reaction of CO₂ with NaCl to produce sodium bicarbonate and ammonium chloride. The ammonia is recovered using the second alkali, lime. Thus, the process involves the use of a calcium base that is less expensive and offers higher CO₂ reactivity, and less scaling problems (because the sorbent is a solid). However, there are some obvious drawbacks: Limestone serves as the source of lime which renders the process ineffective, and considerable energy is consumed in calcination. In addition, as indicated below, for every two moles of CO₂ captured, one mole of CO₂ is released by calcination.

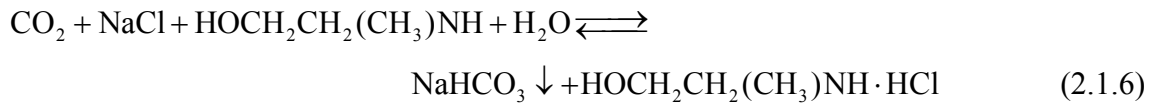




The overall reaction is:



A proposed modified dual-alkali approach is to use methylaminoethanol (MAE) as the primary alkali in the following reaction [32]:



However, a suitable secondary alkali to regenerate the primary alkali, MAE, remains to be identified.

Carbonation/calcination cycles involve two fluidized bed reactors, one an adsorber, the other a regenerator, operating at atmospheric pressure. In the adsorber, CaO is carbonated to CaCO₃ at 600-700 °C. In the regenerator, which operates above 900 °C, CaCO₃ is calcinated to release concentrated CO₂. The high percentage of CO₂ captured (greater than 80%) makes this economically competitive technology [33-36]. The major drawback is the decay of absorbent activity; however, it maintains a higher CO₂ uptake rate than many other sorbents, even when fully degraded.

Ionic liquids are known to exhibit unique physiochemical properties that make them attractive for gas separations [37-42]: i.e., extremely low vapor pressure, wide liquid temperature range, high thermal and chemical stability, and ability to dissolve a variety of chemical compounds. In combination with amines, ionic liquids have been shown to offer such advantages as lower heat capacities, lower water contents and energy

savings [37]. Uptake of CO₂ per mole of ionic liquid can approach 0.5, using an ionic liquid consisting of an imidazolium cation with a primary amine moiety and tetrafluoroborate anion, which makes it effective for CO₂ absorption [38]. However, engineering scale-up of CO₂ capture via ionic liquid-based technology has been limited due to high costs and relatively high viscosities [37, 39]. Furthermore, the toxicology of most ionic liquids remains to be evaluated [40-42].

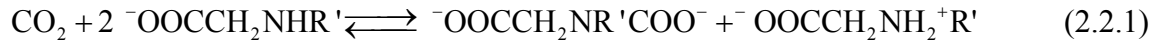
2.2 Kinetics study of CO₂ reactions with amino acid salts

Because the reactive functional groups of amino acids are similar to those of alkanolamines, the reactions mechanism can be expected to be similar. The salts of amino acids have the additional advantage of being an ionic solution, which minimizes volatility, which is especially important at stripper conditions (lowered pressure and elevated temperature). The salt functionality of AAS's increases their surface tension, which makes them promising for gas-liquid membrane applications. In contrast with MEA, which can only be used in combination with expensive membranes due to wetting problems, AAS can be employed in conjunction with cheaper and commercially available membranes such as polypropylene, economically improving the process [43]. Therefore, a review of reaction kinetics of CO₂ with AAS is important for design of gas-liquid contactors.

Among various kinds of AAS's, two potassium salts stand out: the potassium salt of glycine (PG) presents several interesting properties such as very good thermal stability and fast reaction rate towards CO₂. Additionally it is commercially available and relatively cheap. For these merits, it is commonly used for characterization as a CO₂ absorbent. Another potassium amino acid salt, potassium sarcosinate (PS) is also a promising candidate sorbent because of its relatively high CO₂ loading capacity and reactivity, and its relative large window of operation without precipitation during CO₂ absorption [28].

During the absorption of CO₂ in aqueous potassium salt of amino acid, the

following reactions can occur ($R' = -H$ for PG, and $R' = -CH_3$ for PS):

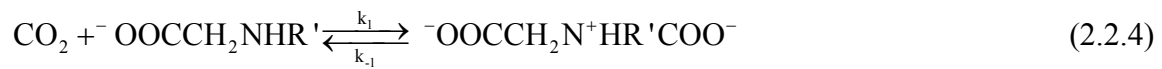


Reaction 2.2.1 is the key reaction in the absorption system, and its forward reaction rate needs to be determined accurately. The forward rate constants and the equilibrium constants of the reactions 2.2.1 and 2.2.2 are available in the literature [44-48]. The reaction between CO_2 and OH^{-} (equation 2.2.2) is fast; however, due to the trace amounts of OH^{-} , this reaction is often negligible [47]. The total rate of CO_2 absorption in an aqueous AAS solution is given by the sum of the reaction rates given by equations 2.2.1-3.

A detailed knowledge of the CO_2 reaction mechanism for alkanolamines is essential for a fundamental understanding of the reaction systems. Generally, the reaction kinetics can be described by either the two-step zwitterion mechanism (proposed originally by Caplow [49] and later reintroduced by Danckwerts [24]) or by the single-step termolecular mechanism (proposed originally by Crooks and Donellan [50] and recently revisited by da Silva and Svendsen [51]). Both mechanisms are extendable to aqueous AAS solution.

The principle difference in applying the zwitterion mechanism to aqueous alkanolamines and AAS is the additional ionic charge associated with the reactant,

product and intermediate species. This may significantly influence the stability and deprotonation rate of the zwitterion, and thereby the overall order of the reaction in aqueous AAS solutions. In this mechanism, the initial step in the reaction of CO₂ with amino acid anion is the formation of the protonated di-carbamate (the key short-lived intermediate) which can then be deprotonated by any base, B_j, in a competitive way, to form the di-carbamate anion according to the following reactions:



The base can be the amino acid anion, water, or a hydroxyl ion [52]. Note that if deprotonation is carried out by the amino acid anion, it forms a zwitterion (protonated carbamate, which can exist at relatively large amount in the solution [53]).

Assuming a quasi-steady-state condition for the protonated di-carbamate concentration, and first order behavior of CO₂ (this is achieved by considering the second proton transfer step to be irreversible, as is the case in the kinetic constant measurement experiments carried out in the pseudo-first-order reaction regime [28]), the overall forward rate of the reaction is given by

$$r_{\text{CO}_2} = \frac{k_1 C_{\text{AAS}} C_{\text{CO}_2}}{1 + \frac{k_{-1}}{\sum k_{B_j} C_{B_j}}} \quad (2.2.6)$$

where $\sum k_{B_j} C_{B_j}$ is the contribution of all the bases present in the solution for the removal of proton. Because the hydroxyl concentration is typically low, its contribution to the

deprotonation of the protonated di-carbamate ion is generally assumed to be negligible [52, 54, 55]. However, in some concentrated AAS solutions, the pH can be quite high, in which case the hydroxyl ion contribution to the deprotonation may not be negligible.

Under certain limiting conditions, reaction 2.2.6 may be simplified as follows.

1. $k_{-1}/(\sum k_{Bj}C_{Bj}) \ll 1$: this results in a simple second order kinetics, which implies that the protonated di-carbamate ion is deprotonated relatively fast in comparison to the reversion rate to CO₂ and the amino acid anion. The overall reaction order is two:

$$r_{\text{CO}_2} = k_1 C_{\text{CO}_2} C_{\text{AAS}} \quad (2.2.7)$$

2. $k_{-1}/(\sum k_{Bj}C_{Bj}) \gg 1$: This results in a more complex kinetic rate expression.

$$r_{\text{CO}_2} = k_1 C_{\text{CO}_2} C_{\text{AAS}} \left(\sum \frac{k_{Bj}}{k_{-1}} C_{Bj} \right) \quad (2.2.8)$$

Depending on the relative contribution of various bases present in the aqueous solution to the deprotonation of the protonated di-carbamate ion, the above expression can explain any reaction order. It can also describe the shifting reaction orders with respect to AAS concentration. For very low AAS concentration, equation 2.2.8 will reduce to the following form and the partial reaction order in AAS is one.

$$r_{\text{CO}_2} = k_1 C_{\text{CO}_2} C_{\text{AAS}} \left(\frac{k_{\text{H}_2\text{O}}}{k_{-1}} C_{\text{H}_2\text{O}} \right) \quad (2.2.9)$$

At moderately high AAS concentrations, the contribution of amino acid anion and water to the protonated di-carbamate anion deprotonation are equally significant, hence equation 2.2.8 must be used in its complete form to describe the experimental

kinetic data.

3. When CO₂ absorbs in a non-aqueous (e.g., alcohol) solution of an AAS, deprotonation of the protonated di-carbamate ion is solely due to amino acid anion. In this case, reaction 2.2.6 reduces to

$$r_{\text{CO}_2} = \frac{k_1 C_{\text{AAS}} C_{\text{CO}_2}}{1 + \frac{k_{-1}}{k_{\text{AAS}} C_{\text{AAS}}}} \quad (2.2.10)$$

At low concentrations of AAS, the second term in the denominator becomes significant and the partial order in AAS is higher than one and this reduces to one at very high AAS concentrations.

For accurate determination of zwitterion rate constants, experimental measurements need to be undertaken with very low to very high concentrations of AAS. From equation 2.2.9, when AAS concentration is very low, $k_{\text{H}_2\text{O}}/k_{-1}$ can be estimated accurately, whereas when AAS is at a very high concentration (equation 2.2.7), k_1 can be obtained independently of other kinetic constants.

For aqueous AAS solutions, there seems to be a significant difference in the kinetic behavior as well as in the magnitude of the rate constants for the formation of di-carbamate anion, compared with the behavior with aqueous alkanolamines. Firstly, contrary to primary aqueous alkanolamines, the partial reaction order in AAS containing the primary amino group changes with the molar salt concentration. This is apparently

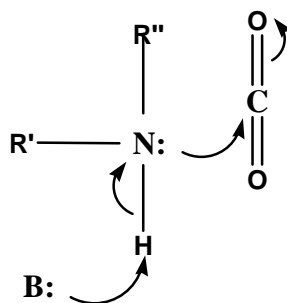
because the deprotonation step in the zwitterion mechanism is not much faster than the protonated di-carbamate ion formation step, a behavior similar to that of secondary aqueous alkanolamines. In addition, it indicates that the protonated di-carbamate ion of AAS is inherently less stable compared to the zwitterion of primary alkanolamine, due to the multiple charges associated with the protonated di-carbamate ion. Other charged species may also have a negative influence on its stability.

Secondly, the ratio of the AAS deprotonation rate constant to the reverse rate constant of protonated di-carbamate formation, k_{AAS}/k_{-1} , is lower than that for aqueous secondary alkanolamines; while that of $k_{\text{H}_2\text{O}}/k_{-1}$ is higher by almost an order of magnitude. (Note that the steric hindrance of secondary alkanolamine has a negative influence on the deprotonation of the zwitterion by a base as compared to primary alkanolamines.) It is apparently relatively easy for the uncharged water molecule to form a hydrogen bond with the protonated di-carbamate ion (which is charged at both ends) as compared to the charged amino acid anion. This indicates that water contributes significantly to the deprotonation even at moderately high AAS concentrations.

Thirdly, precipitation is a problem uniquely associated with the absorption of CO_2 in aqueous AAS solutions that may influence kinetic measurements. Precipitation occurs at high salt concentrations ($> 2 \text{ mol/L}$), depending on temperature and the nature of the AAS [27, 56]. Notably, absorption of CO_2 in aqueous PG and PS solutions is not accompanied by, precipitation, even at high CO_2 loading [27, 47].

Crooks and Donnellan [50] questioned the validity of the zwitterion mechanism,

arguing that the number of fitting parameters required to describe the experimental data is too high and the numerical values of the parameters (especially the deprotonation rate constants) in some cases seem to be physically unrealistic. They proposed a single-step, termolecular mechanism (see scheme 2.2) to describe their experimental kinetic data. This mechanism was used by Kumar et al. [57] to describe CO₂ reaction kinetics with aqueous solutions of potassium taurinate (PT) and PG, assuming that the AAS's react simultaneously with one molecule of CO₂ and one molecule of a base. The reaction is said to proceed in a single step with a loosely bound encounter complex as the intermediate, rather than a zwitterion. The complex dissociates primarily to form CO₂ and salt (i.e., the reactant molecules); a small fraction reacts with a base molecule to give carbamate.



Scheme 2.2 Termolecular mechanism (B is any base, i.e. H₂O, OH⁻ or amino acid anion in the solution) [50]



Assuming a quasi-steady-state for the encounter complex, the reaction rate for this mechanism (shown by scheme 2.2 and equation 2.2.11) is given by equation 2.2.12,

which, in fact, is analogous to one of the limiting cases ($k_{-1}/(\sum k_{Bj}C_{Bj}) \gg 1$) of the zwitterion mechanism.

$$r = (k_{R'R''NH}C_{R'R''NH} + k_{OH^-}C_{OH^-} + k_{H_2O}C_{H_2O})C_{R'R''NH}C_{CO_2} \quad (2.2.12)$$

$$\text{(Not that } k_{R'R''NH} = \frac{k_e}{k_{-1} + k_e}k_{1,R'R''NH}, k_{OH^-} = \frac{k_e}{k_{-1} + k_e}k_{1,OH^-}, \text{ and } k_{H_2O} = \frac{k_e}{k_{-1} + k_e}k_{1,H_2O}\text{)}$$

Equation 2.2.12 suggests that all bases in solution influence the reaction in parallel. Because hydroxyl ion concentrations are typically low, their contribution is generally assumed to be negligible [52, 54, 55]. When water is the dominant base and the contributions of amino acid anion and OH^- to carbamate formation are negligible, the reaction is first order with respect to the salt, i.e.:

$$r = k_{H_2O}C_{H_2O}C_{R'R''NH}C_{CO_2} \quad (2.2.13)$$

When amino acid anion is the dominant base, the reaction is second order with respect to the salt, and the rate is given by

$$r = k_{R'R''NH}C_{R'R''NH}^2C_{CO_2} \quad (2.2.14)$$

It is clear from equation 2.2.12 that the number of fitting parameters in the termolecular mechanism is fewer by one than that in the zwitterion mechanism. Although the termolecular mechanism can describe fractional and higher order kinetics for aqueous alkanolamine solutions [58], it fails to explain the changing reaction orders with respect to amine concentration in nonaqueous solutions of alkanolamines [59, 60].

For most practical purposes (which involve aqueous solutions), equation 2.2.12 and its limiting cases formulations provide a workable engineering model to a wide range

of amine kinetic behavior.

Both the zwitterion mechanism and the termolecular mechanism are commonly used in literature to fit the experimental kinetic data.

2.3 Linearization techniques

From the preceding review of the kinetic study of CO₂ with amines/AAS, it can be seen a set of reversible reactions mediate mass transfer of CO₂. The reaction rate expressions enter into differential species mass balances, which form a set of nonlinear differential equations (see chapter 3). Consequently, mathematical modeling of CO₂ absorption in AAS solutions in packed towers is a computation-intensive task with multiple dimensions of trial-and-error. The most robust algorithms developed for the latter purpose implement numerical methods of analysis [61, 62].

Closed-form approximations based on linearization techniques provide less computation-intensive alternatives and are very useful for preliminary calculations [63-66]. However, they are valid over limited ranges of operating conditions or, with the exception of Onda et al.'s work [65, 66], are applicable only to systems involving uncoupled reactions,

Two other linearization techniques, namely “Thin Film Theory” and “Thick Film Theory”, were developed by K. A. Smith for application to the analysis of carrier-facilitated membrane transport [1, 2]. The former deploys regular perturbation methods to account for reaction-induced departures from purely mass diffusion, while the latter uses singular perturbation methods to calculate diffusion-induced departures from local reaction equilibrium.

“Thick Film Theory” has been applied - within the framework of the steady-state Two-Film Theory of interphase mass transfer [67, 68] - to the analysis of absorption with

a single reversible reaction, whose mathematical description differs from that of carrier-facilitated transport only in the conditions at one boundary [69]. A simple trial-and-error search solves the residual algebra, resulting in enhancement factors, E (by which reaction multiplies absorption rates), that essentially overlap with those obtained via either numerical methods or the venerable linearization scheme developed by Van Krevelen and Hoftijzer [70]. Recently, “Thick Film Theory” was applied to the analysis of CO_2 absorption in and desorption from solutions of monoethanolamine (MEA), a more challenging mathematical problem involving multiple reversible reactions with widely disparate time scales [71].

More-realistic non-steady-state models, such as “Penetration Theory” [72] and “Surface Renewal Theory” [73], involve partial differential equations accounting for the dependence of local concentrations on position as well as time. It is notable that the steady-state and non-steady-state models give identical absorption rates when diffusivities are equal. Moreover, absorption rates based on the “Film Theory” approach those based on “Surface Renewal Theory” when diffusivity ratios are replaced in the analysis by their square roots [74, 75].

2.4 Summary

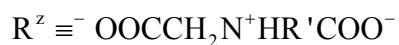
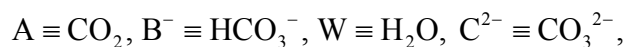
The mathematical analysis adopted here originated with Smith's singular perturbation method [1, 2], and is a direct derivative of the recent work by Meldon [69, 71]. Mathematical analysis of the CO₂-AAS system is very similar to that of the CO₂-MEA system developed by Meldon and Morales-Cabrera [71]. A Film Model is set up with appropriate boundary conditions; differential mass balances are solved via linearizing approximations; yielding a set of algebraic relations that is solved by trial-and-error.

Unlike the reaction kinetics of CO₂ and MEA, where carbamate zwitterion formation is rate-determining (i.e. deprotonation of the zwitterion is fast), the reaction kinetics is more complex. Accordingly, the analysis of CO₂ absorption in solutions of potassium salts of glycine and sarcosine is more complex than that developed for MEA, and is more generally applicable. Deprotonation of the protonated di-carbamate by both the amino acid anion and water are taken into account in the CO₂-AAS system, whereas the contribution of water is neglected in the CO₂-MEA system. Accordingly, the denominator of the di-carbamate consumption rate expression (equation 3.1.9) is algebraically more complex in the CO₂-AAS analysis, whereas in the case of MEA, the algebraic term denominator is negligible. Another major difference is the intrinsically ionic nature of potassium salts, which influences physiochemical properties as well as reaction kinetics.

3. DEVELOPMENT OF THE MATHEMATICAL MODEL

3.1 Reactions

The following shorthand notation for species is adopted here:



Note that for PG, $R' = -\text{H}$; while for PS, $R' = -\text{CH}_3$.

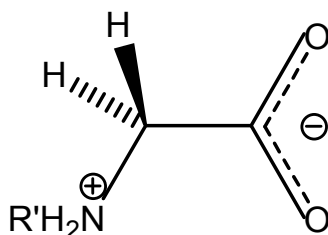


Figure 3.1.1 Zwitterionic structure of glycine ($R' = -\text{H}$) or sarcosine ($R' = -\text{CH}_3$)

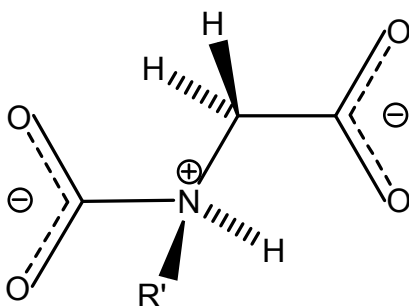


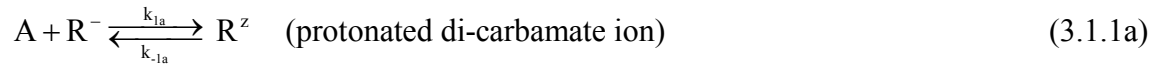
Figure 3.1.2 Structure of protonated di-carbamate ion (R^z) of glycine ($R' = -\text{H}$) or sarcosine ($R' = -\text{CH}_3$)

Potassium amino acid salts are completely ionized in water as potassium cations, and amino acid anions (R^-) with a deprotonated amino group that reacts with CO_2 to form a di-carbamate (R^{2-}). The zwitterionic structure of amino acids in the solid state has been

confirmed by neutron diffraction measurements [76]. As such, the zwitterion (R) formed by protonation of the amino acid anion, may assume the structure shown in Figure 3.1.1. The protonated di-carbamate ion (R^z) is a short-lived intermediate with charge at three locations; its possible structure is shown in Figure 3.1.2.

It is assumed that di-carbamate formation follows the two-step “zwitterion mechanism” [24, 49, 57]:

i) formation of the protonated di-carbamate ion, R^z :



$$r_{1a} = k_{1a}[A][R^-] - k_{-1a}[R^z]$$

$$K_{1a} = \frac{k_{1a}}{k_{-1a}} = \left(\frac{[R^z]}{[A][R^-]} \right)^{\text{eq}}$$

Here, “eq” denotes a condition of reaction equilibrium.

ii) deprotonation of protonated di-carbamate ion:

The protonated di-carbamate ion undergoes deprotonation by any base in solution, B_j , in a competitive way, resulting in a di-carbamate.



$B_j = W, R^-$ (the OH^- is usually at negligible concentration)

$$r_{1b_j} = k_{B_j}[R^z][B_j] - k_{-B_j}[R^{2-}][B_jH^+]$$

$$K_{1b_j} = \frac{k_{B_j}}{k_{-B_j}} = \left(\frac{[R^{2-}][B_jH^+]}{[R^z][B_j]} \right)^{\text{eq}} = \left(\frac{[R^{2-}][H^+]}{[R^z]} \cdot \frac{[B_jH^+]}{[B_j][H^+]} \right)^{\text{eq}}$$

$$= K_{\text{Acid}_j} \left(\frac{[\text{R}^{2-}][\text{H}^+]}{[\text{R}^z]} \right)^{\text{eq}} = K_{\text{Acid}_j} \left(\frac{[\text{A}][\text{R}^-]}{[\text{R}^z]} \right)^{\text{eq}} \left(\frac{[\text{R}^{2-}][\text{H}^+]}{[\text{A}][\text{R}^-]} \right)^{\text{eq}} = \frac{K_{\text{Acid}_j} K_{\text{Carb}}}{K_{1a}}$$

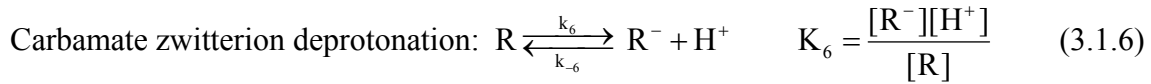
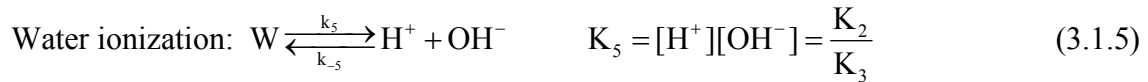
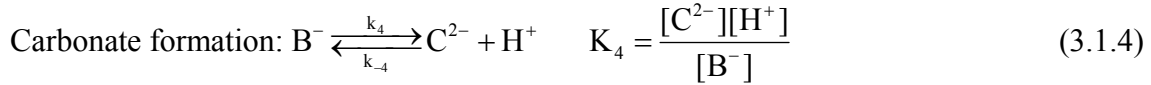
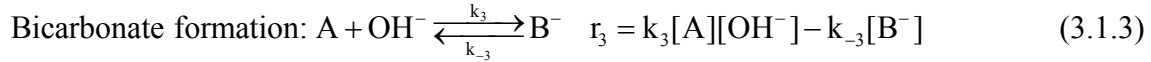
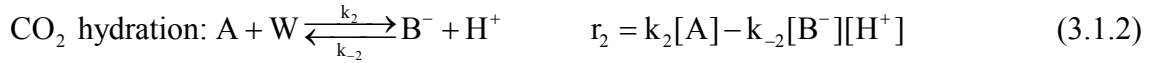
$$\text{where } K_{\text{Acid}_j} \equiv \left(\frac{[\text{B}_j\text{H}^+]}{[\text{B}_j][\text{H}^+]} \right)^{\text{eq}}, \quad K_{\text{Carb}} \equiv \left(\frac{[\text{R}^{2-}][\text{H}^+]}{[\text{A}][\text{R}^-]} \right)^{\text{eq}}$$

Overall reaction j (the sum of 3.1.1a and 3.1.1b):



$$K_{1j} = K_{1a} K_{1bj} = \left(\frac{[\text{R}^{2-}]}{[\text{A}][\text{R}^-]} \right)^{\text{eq}} \left(\frac{[\text{B}_j\text{H}^+]}{[\text{B}_j]} \right)^{\text{eq}} = \left(\frac{[\text{R}^{2-}][\text{H}^+]}{[\text{A}][\text{R}^-]} \right)^{\text{eq}} \left(\frac{[\text{B}_j\text{H}^+]}{[\text{B}_j][\text{H}^+]} \right)^{\text{eq}} \equiv K_{\text{Carb}} K_{\text{Acid}_j}$$

Other reactions in solution are:



Reaction 3.1.4-6 are effectively instantaneous. Reaction 3.1.2 combines the slow formation and rapid dissociation of carbonic acid. The concentration of water is implicitly incorporated into k_2 and K_5 .

The quasi-steady-state assumption is applied to the protonated di-carbamate ion:

$$\frac{d[\text{R}^z]}{dt} = r_{1a} - \Sigma r_{1b} = 0 = k_{1a}[\text{A}][\text{R}^-] - k_{-1a}[\text{R}^z] - \Sigma k_{B_j}[\text{B}_j][\text{R}^z] + \Sigma k_{-B_j}[\text{R}^{2-}][\text{B}_j\text{H}^+] \quad (3.1.7)$$

The protonated di-carbamate ion concentration then becomes:

$$[R^z] = \frac{k_{1a}[A][R^-] + [R^{2-}]\Sigma k_{-B_j}[B_j H^+]}{k_{-1a} + \Sigma k_{B_j}[B_j]} \quad (3.1.8)$$

Therefore,

$$\begin{aligned} r_{1a} &= \frac{k_{1a}[A][R^-]\Sigma k_{B_j}[B_j] - k_{-1a}[R^{2-}]\Sigma k_{-B_j}[B_j H^+]}{k_{-1a} + \Sigma k_{B_j}[B_j]} \\ &= \frac{k_{1a}[A][R^-]\Sigma \left(k_{B_j}[B_j] \left(1 - \frac{[R^{2-}][B_j H^+]}{K_{1a} K_{1b_j}[B_j][A][R^-]} \right) \right)}{k_{-1a} + \Sigma k_{B_j}[B_j]} \\ &= \frac{\Sigma \left[\left(k_{1a} k_{B_j}[B_j][A][R^-] \right) \left(1 - \frac{[R^{2-}][B_j H^+]}{K_{Carb} K_{Acid_j}[B_j][A][R^-]} \right) \right]}{k_{-1a} + \Sigma k_{B_j}[B_j]} \end{aligned} \quad (3.1.9)$$

3.2 Mass transfer model

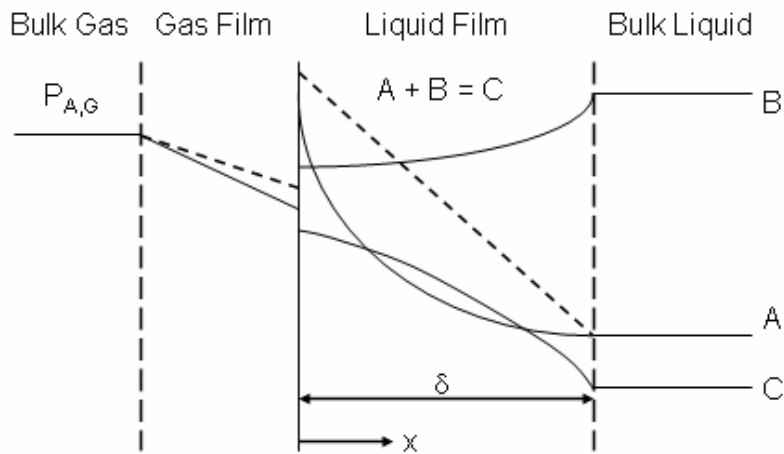


Figure 3.2 Concentration profiles in the steady-state “Film Theory” model (for $A + B = C$ reaction); (dotted lines: physical absorption; solid lines: absorption with reversible reaction)

The analysis is based on simple steady-state Film Theory [67, 68] (Figure 3.2) in which interphase mass transfer is controlled by diffusion and reaction in gas and liquid films that separate well-mixed bulk fluids. Given the compositions of the latter, calculation of purely physical mass transfer rates is straightforward; while calculation of reaction-enhanced mass transfer rates requires the solution of nonlinear ordinary differential equations.

3.3 Differential mass balance

The key equations are species mass balances:

$$D_i \frac{d^2 C_i}{dx^2} = \rho_i \quad (0 \leq x \leq \delta) \quad (3.3)$$

ρ_i is the local rate of consumption and D_i the diffusion coefficient of component i ; δ is the liquid film thickness, calculated as D_{CO_2}/k_L^o , where k_L^o is CO_2 's liquid phase mass transfer coefficient. Gas phase mass transfer resistance is neglected, and electrical effects due to differences in diffusivities of ionic species are also neglected. For simplicity, ionic species are assumed to have the same diffusion coefficient, D^* . Analysis of electrical effects indicates that the above assumption introduces little error when applied to the system considered here [2, 64].

3.4 Boundary conditions

$$\text{At } x = 0: [A] = \alpha p_{A,G}, \quad \frac{d[i]}{dx} = 0 \quad (i \neq A) \quad (3.4.1)$$

$$\text{At } x = \delta: [i] = [i]^{eq} \quad (3.4.2)$$

The assumption of overall reaction equilibrium at $x = \delta$ implies sufficiently long residence times in bulk liquid, i.e. sufficient liquid hold-up.

3.5 Preliminary analysis

Electroneutrality, which effectively prevails locally within the liquid film, is expressed as follows (neglecting the contributions of short-lived protonated di-carbamate ions, H^+ and OH^-):

$$[K^+] = [B^-] + [R^-] + 2[R^{2-}] + 2[C^{2-}] \quad ([K^+] \text{ is a constant}) \quad (3.5.1)$$

“CO₂ loading”, L , is defined as follows (again neglecting the contributions of the short-lived protonated di-carbamate ions):

$$L \cdot [R]_{\text{total}} = [A] + [R^{2-}] + [B^-] + [C^{2-}] = \alpha p_{A,G} + [R^{2-}] + [B^-] + [C^{2-}] \quad (3.5.2)$$

i.e. L is total moles of reversibly absorbed CO₂ per mole of amine or amino acid.

The reaction terms in equation 3.3 may be expanded as follows:

$$\begin{aligned} \rho_A &= r_{1a} + r_2 + r_3, & \rho_{B^-} &= -r_2 - r_3 + r_4, & \rho_{R^-} &= r_{1a} + r_{1b,R^-} - r_6, & \rho_{R^{2-}} &= -\sum r_{1b} = -r_{1a}, \\ \rho_R &= -r_{1b,R^-} + r_6, & \rho_{C^{2-}} &= -r_4 \end{aligned}$$

We neglect the contribution of OH^- to protonated di-carbamate deprotonation due to its negligible concentration in the solution, i.e. the significant bases are amino acid anion and water (k_W and k_{R^-} denote rate constants for protonated di-carbamate deprotonation by water and amino acid anion, respectively). Equations 3.1.5 and 3.1.6 are used to eliminate H^+ and OH^- . Thus, equation 3.3, applied to CO₂, becomes:

$$\begin{aligned} D_A \frac{d^2[A]}{dx^2} &= \frac{\sum \left[\left(k_{1a} k_{B_j} [B_j] [A] [R^-] \right) \left(1 - \frac{[R^{2-}] [B_j H^+]}{K_{\text{Carb}} K_{\text{Acid}_j} [B_j] [A] [R^-]} \right) \right]}{k_{-1a} + \sum k_{B_j} [B_j]} \\ &+ k_2 [A] - k_{-2} [B^-] [H^+] + k_3 [A] [OH^-] - k_{-3} [B^-] \end{aligned}$$

$$= \frac{\left[\begin{array}{l} k_{1a}k_{R^-}[A][R^-]^2 \left(1 - \frac{[R^{2-}][R]}{K_{\text{Carb}}K_{\text{Acid},R^-}[A][R^-]^2} \right) + \\ k_{1a}k_w[W][A][R^-] \left(1 - \frac{[R^{2-}][H^+]}{K_{\text{Carb}}K_{\text{Acid},W}[W][A][R^-]} \right) \end{array} \right]}{k_{-1a} + k_{R^-}[R^-] + k_w[W]} \\ + k_2[A] - \frac{k_{-2}K_6[B^-][R]}{[R^-]} + \frac{k_3K_2[A][R^-]}{K_3K_6[R]} - k_{-3}[B^-]$$

Note that $K_{\text{Acid},R^-} = \frac{1}{K_6}$ and $K_{\text{Acid},W} = \frac{1}{[W]}$, and the above equation becomes:

$$= \frac{k_{1a}k_{R^-}[A][R^-]^2 \left(1 - \frac{K_6[R^{2-}][R]}{K_{\text{Carb}}[A][R^-]^2} \right) + k_{1a}k_w[W][A][R^-] \left(1 - \frac{[W][R^{2-}][H^+]}{K_{\text{Carb}}[W][A][R^-]} \right)}{k_{-1a} + k_{R^-}[R^-] + k_w[W]} \\ + k_2[A] - \frac{k_{-2}K_6[B^-][R]}{[R^-]} + \frac{k_3K_2[A][R^-]}{K_3K_6[R]} - k_{-3}[B^-] \\ = \frac{k_{1a}k_{R^-}[A][R^-]^2 \left(1 - \frac{K_6[R^{2-}][R]}{K_{\text{Carb}}[A][R^-]^2} \right) + k_{1a}k_w[W][A][R^-] \left(1 - \frac{K_6[R^{2-}][R]}{K_{\text{Carb}}[A][R^-]^2} \right)}{k_{-1a} + k_{R^-}[R^-] + k_w[W]} \\ + k_2[A] - \frac{k_{-2}K_6[B^-][R]}{[R^-]} + \frac{k_3K_2[A][R^-]}{K_3K_6[R]} - k_{-3}[B^-] \\ = \frac{k_{1a}[A][R^-] \left(k_{R^-}[R^-] + k_w[W] \right) \left(1 - \frac{K_6[R^{2-}][R]}{K_{\text{Carb}}[A][R^-]^2} \right)}{k_{-1a} + k_{R^-}[R^-] + k_w[W]} \\ + k_2[A] - \frac{k_{-2}K_6[B^-][R]}{[R^-]} + \frac{k_3K_2[A][R^-]}{K_3K_6[R]} - k_{-3}[B^-] \quad (3.5.3)$$

Application of equation 3.3 to the three amino acid species and summation of the results gives:

$$D^* \frac{d^2}{dx^2} ([R^-] + [R^{2-}] + [R]) = 0 \quad (3.5.4)$$

Successive integrations of equation 3.5.4, and application of zero-flux boundary

conditions and the specified total amine concentration in bulk liquid, yields the following relationship, which is applicable throughout the liquid film:

$$[R^-] + [R^{2-}] + [R] = [R]_{\text{total}} \quad (3.5.5)$$

Likewise, application of equation 3.3 to CO₂, bicarbonate, carbonate and di-carbamate, followed by summation, yields:

$$D_A \frac{d^2[A]}{dx^2} + D^* \frac{d^2}{dx^2} ([B^-] + [R^{2-}] + [C^{2-}]) = 0 \quad (3.5.6)$$

Integration gives:

$$D_A \frac{d[A]}{dx} + D^* \frac{d}{dx} ([B^-] + [R^{2-}] + [C^{2-}]) = -\Phi_1 \quad (3.5.7)$$

Φ_1 may be identified as the CO₂ absorption rate expressed as a flux.

A second integration of equation 3.5.7, gives:

$$D_A [A] + D^* ([B^-] + [R^{2-}] + [C^{2-}]) = -\Phi_1 x + \Phi_2 \quad (3.5.8)$$

Application of equation 3.5.8 at $x = 0$, gives:

$$\Phi_2 = D_A [A]_{x=0} + D^* ([B^-]_{x=0} + [R^{2-}]_{x=0} + [C^{2-}]_{x=0}) \quad (3.5.9a)$$

Application of equation 3.5.8 at $x = \delta$, gives:

$$\Phi_2 = \Phi_1 \delta + D_A [A]_{x=\delta} + D^* ([B^-]_{x=\delta} + [R^{2-}]_{x=\delta} + [C^{2-}]_{x=\delta}) \quad (3.5.9b)$$

Subtraction of equation 3.5.9a from 3.5.9b and rearrangement yields the following expression for the absorption rate:

$$\Phi_1 = \frac{D_A ([A]_{x=0} - [A]_{x=\delta}) + D^* ([B^-]_{x=0} - [B^-]_{x=\delta} + [R^{2-}]_{x=0} - [R^{2-}]_{x=\delta} + [C^{2-}]_{x=0} - [C^{2-}]_{x=\delta})}{\delta} \quad (3.5.10)$$

Note that $[B^-]_{x=0}$, $[R^{2-}]_{x=0}$ and $[C^{2-}]_{x=0}$ are unknowns.

Reaction effects are typically expressed in terms of the “enhancement factor”, E ,

defined as the ratio of absorption rates with and without reaction:

$$E = 1 + \beta \left(\frac{[B^-]_{x=0} - [B^-]_{x=\delta} + [R^{2-}]_{x=0} - [R^{2-}]_{x=\delta} + [C^{2-}]_{x=0} - [C^{2-}]_{x=\delta}}{[A]_{x=0} - [A]_{x=\delta}} \right) \left(\beta \equiv \sqrt{\frac{D^*}{D_A}} \right) \quad (3.5.11)$$

Replacement of the ratio of diffusivities, β , with the ratio of their square roots, brings mass transfer rates calculated on the basis of the Film Model closer to those based on non-steady-state Surface Renewal Theory. The latter, more physically realistic mass transfer model, accounts for periodic replacement of surface liquid with liquid from the bulk. The two models yield identical absorption rates when diffusivities are all equal [74, 75].

3.6 Limiting cases

When δ is small in the sense that reaction times far exceed diffusion times, the extents of reaction 3.1.1-4 are negligible except in bulk liquid. Concentrations of species in the liquid film other than CO_2 are then essentially uniform. The enhancement factor approaches 1, and the absorption rate simplifies to:

$$\Phi_1^0 = k_L^0 ([A]_{x=0} - [A]_{x=\delta}) \quad (3.6.1)$$

where k_L^0 is CO_2 's liquid phase mass transfer coefficient.

Correspondingly, when δ is large enough, all reactions approach local equilibrium effectively instantaneously, and the absorption rate and enhancement factor become:

$$\Phi_1^{\text{eq}} = \frac{D_A ([A]_{x=0} - [A]_{x=\delta}) + D^* ([B^-]_{x=0}^{\text{eq}} - [B^-]_{x=\delta} + [R^{2-}]_{x=0}^{\text{eq}} - [R^{2-}]_{x=\delta} + [C^{2-}]_{x=0}^{\text{eq}} - [C^{2-}]_{x=\delta})}{\delta} \quad (3.6.2)$$

$$E^{\text{eq}} = 1 + \beta \left(\frac{[B^-]_{x=0}^{\text{eq}} - [B^-]_{x=\delta} + [R^{2-}]_{x=0}^{\text{eq}} - [R^{2-}]_{x=\delta} + [C^{2-}]_{x=0}^{\text{eq}} - [C^{2-}]_{x=\delta}}{[A]_{x=0} - [A]_{x=\delta}} \right) \quad (3.6.3)$$

where $[B^-]_{x=0}^{\text{eq}}$, $[C^{2-}]_{x=0}^{\text{eq}}$ and $[R^{2-}]_{x=0}^{\text{eq}}$ follow from equations 3.1.4, 3.1.6, 3.5.1, 3.5.5 and:

$$\left(\frac{[R^{2-}][H^+]}{[A][R^-]} \right)^{\text{eq}} = K_{\text{Carb}} \quad (3.6.4)$$

$$\left(\frac{[B^-][H^+]}{[A]} \right)^{\text{eq}} = K_2 \quad (3.6.5)$$

3.7 General case

Equation 3.3, when applied to R^{2-} , gives:

$$D^* \frac{d^2[R^{2-}]}{dx^2} = - \frac{k_{la}[A][R^-] (k_{R^-}[R^-] + k_w[W]) \left(1 - \frac{K_6[R^{2-}][R]}{K_{Carb}[A][R^-]^2} \right)}{k_{-la} + k_{R^-}[R^-] + k_w[W]} \quad (3.7.1)$$

The ‘‘Thick Film’’ linearization technique [1, 2] is suggested by the asymptotic approach to local reaction equilibrium in thick films. Reaction rates may then be safely approximated by retaining only first order terms in their Taylor series expansions (see equation 3.7.2).

$$r \approx \sum_i \Delta[i] \left(\frac{\partial r}{\partial [i]} \right)_{j \neq i} \quad (3.7.2)$$

$$\Delta[i] \equiv [i] - \underline{[i]} \quad (3.7.3)$$

$\Delta[i]$ is the departure from reaction equilibrium; local pseudo-equilibrium concentrations,

$\underline{[i]}$, are defined to satisfy equations 3.5.1, 3.5.5, 3.5.8 and all reaction equilibria, i.e.:

$$\underline{[R^-]} + \underline{[R]} + \underline{[R^{2-}]} = [R]_{total} \quad (3.7.4)$$

$$\underline{[K^+]} = \underline{[B^-]} + \underline{[R^-]} + 2\underline{[R^{2-}]} + 2\underline{[C^{2-}]} \quad (3.7.5)$$

$$D_A \underline{[A]} + D^* (\underline{[B^-]} + \underline{[R^{2-}]} + \underline{[C^{2-}]}) = -\Phi_1 x + \Phi_2 \quad (3.7.6)$$

$$\frac{\underline{[R^{2-}][R]}}{\underline{[A][R^-]^2}} = \frac{K_{Carb}}{K_6} \quad (3.7.7)$$

$$\frac{\underline{[B^-][R]}}{\underline{[A][R^-]}} = \frac{K_2}{K_6} \quad (3.7.8)$$

Together with equation 3.7.3, paired equations 3.5.1 and 3.7.4, 3.5.5 and 3.7.5, and 3.5.8 and 3.7.6, imply that:

$$\Delta[R^-] + \Delta[R] + \Delta[R^{2-}] = 0 \quad (3.7.9)$$

$$\Delta[B^-] + \Delta[R^-] + 2\Delta[R^{2-}] + 2\Delta[C^{2-}] = 0 \quad (3.7.10)$$

$$D_A \Delta[A] + D^* (\Delta[B^-] + \Delta[R^{2-}] + \Delta[C^{2-}]) = 0 \quad (3.7.11)$$

From the reaction equilibrium $\frac{[H^+][R^-]}{[R]} = K_6$, i.e. $[H^+] = \frac{K_6[R]}{[R^-]}$, substituting

this into $\frac{[C^{2-}][H^+]}{[B^-]} = K_4$ gives:

$$\frac{[C^{2-}][R]}{[R^-][B^-]} = \frac{[C^{2-}][R]}{[R^-][B^-]} = \frac{K_4}{K_6} \quad (3.7.12)$$

$$\Rightarrow \left\{ \begin{array}{l} [C^{2-}] = \frac{K_4 [R^-][B^-]}{K_6 [R]} \end{array} \right. \quad (3.7.13)$$

$$\Rightarrow \left\{ \begin{array}{l} [C^{2-}] = \frac{K_4 [R^-][B^-]}{K_6 [R]} \end{array} \right. \quad (3.7.14)$$

Subtraction of equation 3.7.14 from 3.7.13 gives:

$$\begin{aligned} \Delta[C^{2-}] &= \frac{K_4}{K_6} \left(\frac{([R^-] + \Delta[R^-])([B^-] + \Delta[B^-])}{[R] + \Delta[R]} - \frac{[R^-][B^-]}{[R]} \right) \\ &= \frac{K_4}{K_6} \left(\frac{([R^-] + \Delta[R^-])([B^-] + \Delta[B^-])([R] - \Delta[R])}{[R]^2} - \frac{[R^-][B^-]}{[R]} \right) \\ &= \frac{K_4}{K_6} \left(\frac{[R][R^-][B^-] + [R][B^-]\Delta[R^-] + [R][R^-]\Delta[B^-] - [R^-][B^-]\Delta[R]}{[R]^2} - \frac{[R^-][B^-]}{[R]} \right) \\ &= \frac{K_4}{K_6} \left(\frac{[R][B^-]\Delta[R^-] + [R][R^-]\Delta[B^-] - [R^-][B^-]\Delta[R]}{[R]^2} \right) \\ &= \frac{K_4 [B^-]}{K_6 [R]} \Delta[R^-] + \frac{K_4 [R^-]}{K_6 [R]} \Delta[B^-] - \frac{K_4 [R^-][B^-]}{K_6 [R]^2} \Delta[R] \end{aligned} \quad (3.7.15)$$

Using equations 3.7.9-11 and 3.7.15, $\Delta[B^-]$, $\Delta[R]$, and $\Delta[R^-]$ may be expressed in terms of $\Delta[A]$ and $\Delta[R^{2-}]$:

$$\begin{pmatrix} 0 & 1 & 1 \\ 1 + \frac{2K_4[R^-]}{K_6[R]} & -\frac{2K_4[R^-][B^-]}{K_6[R]^2} & 1 + \frac{2K_4[B^-]}{K_6[R]} \\ 1 + \frac{K_4[R^-]}{K_6[R]} & -\frac{K_4[R^-][B^-]}{K_6[R]^2} & \frac{K_4[B^-]}{K_6[R]} \end{pmatrix} \cdot \begin{pmatrix} \Delta[B^-] \\ \Delta[R] \\ \Delta[R^-] \end{pmatrix} = \begin{pmatrix} -1 & 0 \\ -2 & 0 \\ -1 & -\frac{D_A}{D^*} \end{pmatrix} \cdot \begin{pmatrix} \Delta[R^{2-}] \\ \Delta[A] \end{pmatrix}$$

$$\begin{pmatrix} \Delta[B^-] \\ \Delta[R] \\ \Delta[R^-] \end{pmatrix} = \begin{pmatrix} 0 & 1 & 1 \\ 1 + \frac{2K_4[R^-]}{K_6[R]} & -\frac{2K_4[R^-][B^-]}{K_6[R]^2} & 1 + \frac{2K_4[B^-]}{K_6[R]} \\ 1 + \frac{K_4[R^-]}{K_6[R]} & -\frac{K_4[R^-][B^-]}{K_6[R]^2} & \frac{K_4[B^-]}{K_6[R]} \end{pmatrix}^{-1} \cdot \begin{pmatrix} -1 & 0 \\ -2 & 0 \\ -1 & -\frac{D_A}{D^*} \end{pmatrix} \cdot \begin{pmatrix} \Delta[R^{2-}] \\ \Delta[A] \end{pmatrix}$$

$$\Rightarrow \begin{pmatrix} \Delta[B^-] \\ \Delta[R] \\ \Delta[R^-] \end{pmatrix} \equiv \text{DeltaMatrix}_{3 \times 2} \cdot \begin{pmatrix} \Delta[R^{2-}] \\ \Delta[A] \end{pmatrix}$$

Thus,

$$\Delta[B^-] = \omega_1 \Delta[R^{2-}] + \omega_2 \Delta[A] \quad (3.7.16)$$

$$\Delta[R] = \omega_3 \Delta[R^{2-}] + \omega_4 \Delta[A] \quad (3.7.17)$$

$$\Delta[R^-] = \omega_5 \Delta[R^{2-}] + \omega_6 \Delta[A] \quad (3.7.18)$$

$$\begin{pmatrix} \omega_1 = \text{DeltaMatrix}(1,1) & \omega_2 = \text{DeltaMatrix}(1,2) \\ \omega_3 = \text{DeltaMatrix}(2,1) & \omega_4 = \text{DeltaMatrix}(2,2) \\ \omega_5 = \text{DeltaMatrix}(3,1) & \omega_6 = \text{DeltaMatrix}(3,2) \end{pmatrix}$$

Assuming, further, that $\frac{d^2[i]}{dx^2} \ll \frac{d^2\Delta[i]}{dx^2}$, and inserting the linearizing approximations,

equation 3.7.3, for species concentrations, the righthand side of equation 3.7.1 becomes:

$$D^* \frac{d^2\Delta[R^{2-}]}{dx^2} = - \frac{k_{-1a} ([A] + \Delta[A])([R^-] + \Delta[R^-]) [k_{R^-} ([R^-] + \Delta[R^-]) + k_w [W]]}{k_{-1a} + k_{R^-} ([R^-] + \Delta[R^-]) + k_w [W]} \cdot \left(1 - \frac{K_6 ([R^{2-}] + \Delta[R^{2-}])([R] + \Delta[R])}{K_{Carb} ([A] + \Delta[A])([R^-] + \Delta[R^-])^2} \right)$$

What follows makes use of the approximations:

$$\begin{aligned} & \frac{1}{k_{-1a} + k_{R^-} ([R^-] + \Delta[R^-]) + k_w [W]} \\ &= \frac{k_{-1a} + k_{R^-} [R^-] + k_w [W] - k_{R^-} \Delta[R^-]}{(k_{-1a} + k_{R^-} [R^-] + k_w [W] + k_{R^-} \Delta[R^-]) (k_{-1a} + k_{R^-} [R^-] + k_w [W] - k_{R^-} \Delta[R^-])} \\ &= \frac{k_{-1a} + k_{R^-} [R^-] + k_w [W] - k_{R^-} \Delta[R^-]}{\left[(k_{-1a} + k_{R^-} [R^-] + k_w [W])^2 + k_{R^-} \Delta[R^-] (k_{-1a} + k_{R^-} [R^-] + k_w [W]) \right.} \\ & \quad \left. - k_{R^-} \Delta[R^-] (k_{-1a} + k_{R^-} [R^-] + k_w [W]) - (k_{R^-} \Delta[R^-])^2 \right]} \\ &= \frac{k_{-1a} + k_{R^-} [R^-] + k_w [W] - k_{R^-} \Delta[R^-]}{(k_{-1a} + k_{R^-} [R^-] + k_w [W])^2 - (k_{R^-} \Delta[R^-])^2} \\ &\approx \frac{k_{-1a} + k_{R^-} [R^-] + k_w [W] - k_{R^-} \Delta[R^-]}{(k_{-1a} + k_{R^-} [R^-] + k_w [W])^2} \quad (\text{by retaining delta quantities to the first power}) \\ &= \frac{1}{k_{-1a} + k_{R^-} [R^-] + k_w [W]} - \frac{k_{R^-} \Delta[R^-]}{(k_{-1a} + k_{R^-} [R^-] + k_w [W])^2} \end{aligned}$$

By the same token, $\frac{1}{[A] + \Delta[A]} \approx \frac{[A] - \Delta[A]}{[A]^2}$ and $\frac{1}{([R^-] + \Delta[R^-])^2} \approx \frac{([R^-] - \Delta[R^-])^2}{[R^-]^4}$;

the righthand side of equation 3.7.1 then becomes:

$$\begin{aligned}
&= -k_{1a} \left(\underline{[A][R^-]} + \underline{[R^-]\Delta[A]} + \underline{[A]\Delta[R^-]} \right) \left(k_{R^-} \underline{[R^-]} + k_{R^-} \Delta[R^-] + k_w [W] \right) \\
&\quad \cdot \left(\frac{1}{k_{-1a} + k_{R^-} \underline{[R^-]} + k_w [W]} - \frac{k_{R^-} \Delta[R^-]}{\left(k_{-1a} + k_{R^-} \underline{[R^-]} + k_w [W] \right)^2} \right) \\
&\quad \cdot \left(1 - \frac{K_6 (\underline{[R^{2-}][R]} + \underline{[R]\Delta[R^{2-}]} + \underline{[R^{2-}]\Delta[R]}) (\underline{[A]} - \Delta[A]) (\underline{[R^-]} - \Delta[R^-])^2}{K_{\text{Carb}} \underline{[A]}^2 \underline{[R^-]}^4} \right)
\end{aligned}$$

The equilibrium expression, equation 3.7.7, may be used to reduce the following expression:

$$\begin{aligned}
&\left(1 - \frac{K_6 (\underline{[R^{2-}][R]} + \underline{[R]\Delta[R^{2-}]} + \underline{[R^{2-}]\Delta[R]}) (\underline{[A]} - \Delta[A]) (\underline{[R^-]} - \Delta[R^-])^2}{K_{\text{Carb}} \underline{[A]}^2 \underline{[R^-]}^4} \right) \text{ to:} \\
&-\left(\frac{\Delta[R^{2-}]}{\underline{[R^{2-}]} + \frac{\Delta[R]}{[R]} - \frac{\Delta[A]}{[A]} - \frac{2\Delta[R^-]}{[R^-]}} \right).
\end{aligned}$$

This simplifies the righthand side of equation 3.7.1 to:

$$\begin{aligned}
&= k_{1a} \left(k_{R^-} \underline{[A][R^-]}^2 + k_{R^-} \underline{[R^-]}^2 \Delta[A] + 2k_{R^-} \underline{[R^-][A]\Delta[R^-]} \right) \\
&\quad + k_w [W] \underline{[A][R^-]} + k_w [W] \underline{[R^-]\Delta[A]} + k_w [W] \underline{[A]\Delta[R^-]} \\
&\quad \cdot \left(\frac{1}{k_{-1a} + k_{R^-} \underline{[R^-]} + k_w [W]} - \frac{k_{R^-} \Delta[R^-]}{\left(k_{-1a} + k_{R^-} \underline{[R^-]} + k_w [W] \right)^2} \right) \\
&\quad \cdot \left(\frac{\Delta[R^{2-}]}{\underline{[R^{2-}]} + \frac{\Delta[R]}{[R]} - \frac{\Delta[A]}{[A]} - \frac{2\Delta[R^-]}{[R^-]}} \right)
\end{aligned}$$

Finally, by retaining only the first order delta terms, the righthand side of equation 3.7.1 simplifies to:

$$= k_{1a} \left(\frac{k_{R^-} [A][R^-]^2 + k_w [W][A][R^-]}{k_{-1a} + k_{R^-} [R^-] + k_w [W]} \right) \cdot \left(\frac{\Delta[R^{2-}]}{[R^{2-}]} + \frac{\Delta[R]}{[R]} - \frac{\Delta[A]}{[A]} - \frac{2\Delta[R^-]}{[R^-]} \right)$$

If we then insert equations 3.7.16-18, the above equation becomes:

$$= k_{1a} \left(\frac{k_{R^-} [A][R^-]^2 + k_w [W][A][R^-]}{k_{-1a} + k_{R^-} [R^-] + k_w [W]} \right) \cdot \left(\frac{\Delta[R^{2-}]}{[R^{2-}]} + \frac{\omega_3 \Delta[R^{2-}] + \omega_4 \Delta[A]}{[R]} - \frac{\Delta[A]}{[A]} - \frac{2(\omega_5 \Delta[R^{2-}] + \omega_6 \Delta[A])}{[R^-]} \right)$$

$$= k_{1a} \left(\frac{k_{R^-} [A][R^-]^2 + k_w [W][A][R^-]}{k_{-1a} + k_{R^-} [R^-] + k_w [W]} \right) \cdot \left(\left(\frac{1}{[R^{2-}]} + \frac{\omega_3}{[R]} - \frac{2\omega_5}{[R^-]} \right) \Delta[R^{2-}] + \left(\frac{\omega_4}{[R]} - \frac{1}{[A]} - \frac{2\omega_6}{[R^-]} \right) \Delta[A] \right)$$

or :

$$\frac{d^2 \Delta[R^{2-}]}{dx^2} = \eta_1 \Delta[R^{2-}] + \eta_2 \Delta[A] \quad (3.7.19)$$

$$\left[\begin{array}{l} \eta_1 \equiv \frac{k_{1a}}{D^*} \left(\frac{k_{R^-} [A][R^-]^2 + k_w [W][A][R^-]}{k_{-1a} + k_{R^-} [R^-] + k_w [W]} \right) \cdot \left(\frac{1}{[R^{2-}]} + \frac{\omega_3}{[R]} - \frac{2\omega_5}{[R^-]} \right) \\ \eta_2 \equiv \frac{k_{1a}}{D^*} \left(\frac{k_{R^-} [A][R^-]^2 + k_w [W][A][R^-]}{k_{-1a} + k_{R^-} [R^-] + k_w [W]} \right) \cdot \left(\frac{\omega_4}{[R]} - \frac{1}{[A]} - \frac{2\omega_6}{[R^-]} \right) \end{array} \right]$$

In the same fashion, when the above linearization methods are applied to the sum of reaction rates r_2 and r_3 , the result is:

$$\begin{aligned}
r_2 + r_3 &= k_2[A] - k_{-2}[B^-][H^+] + k_3[A][OH^-] - k_{-3}[B^-] \\
&= k_2[A] - \frac{k_{-2}K_6[B^-][R]}{[R^-]} + \frac{k_3K_5[A][R^-]}{K_6[R]} - k_{-3}[B^-] \\
&= k_2(\underline{[A]} + \Delta[A]) - \frac{k_2K_6(\underline{[B^-]} + \Delta[B^-])(\underline{[R]} + \Delta[R])(\underline{[R^-]} - \Delta[R^-])}{K_2\underline{[R^-]}^2} \\
&\quad + \frac{k_3K_5(\underline{[A]} + \Delta[A])(\underline{[R^-]} + \Delta[R^-])(\underline{[R]} - \Delta[R])}{K_6\underline{[R]}^2} - \frac{k_3(\underline{[B^-]} + \Delta[B^-])}{K_3} \\
&= k_2\underline{[A]} + k_2\Delta[A] - \frac{k_3\underline{[B^-]} + k_3\Delta[B^-]}{K_3} \\
&\quad - \frac{k_2K_6(\underline{[B^-]}[R][R^-] + [R][R^-]\Delta[B^-] + [B^-][R^-]\Delta[R] - [B^-][R]\Delta[R^-])}{K_2\underline{[R^-]}^2} \\
&\quad + \frac{k_3K_5(\underline{[A]}[R^-][R] + [R^-][R]\Delta[A] + [A][R]\Delta[R^-] - [A][R^-]\Delta[R])}{K_6\underline{[R]}^2} \\
&= k_2\Delta[A] - \frac{k_3\Delta[B^-]}{K_3} - \frac{k_2[A]\Delta[B^-]}{\underline{[B^-]}} - \frac{k_2[A]\Delta[R]}{\underline{[R]}} + \frac{k_2[A]\Delta[R^-]}{\underline{[R^-]}} \\
&\quad + \frac{k_3K_5\underline{[R^-]}\Delta[A]}{K_6\underline{[R]}} + \frac{k_3K_5[A]\Delta[R^-]}{K_6\underline{[R]}} - \frac{k_3\underline{[B^-]}\Delta[R]}{K_3\underline{[R]}} \\
&= k_2\Delta[A] - \frac{k_3(\omega_1\Delta[R^{2-}] + \omega_2\Delta[A])}{K_3} - \frac{k_2[A](\omega_1\Delta[R^{2-}] + \omega_2\Delta[A])}{\underline{[B^-]}} \\
&\quad - \frac{k_2[A](\omega_3\Delta[R^{2-}] + \omega_4\Delta[A])}{\underline{[R]}} + \frac{k_2[A](\omega_5\Delta[R^{2-}] + \omega_6\Delta[A])}{\underline{[R^-]}} + \frac{k_3K_5\underline{[R^-]}\Delta[A]}{K_6\underline{[R]}} \\
&\quad + \frac{k_3K_5[A](\omega_5\Delta[R^{2-}] + \omega_6\Delta[A])}{K_6\underline{[R]}} - \frac{k_3\underline{[B^-]}(\omega_3\Delta[R^{2-}] + \omega_4\Delta[A])}{K_3\underline{[R]}}
\end{aligned}$$

$$\begin{aligned}
&= k_2 \Delta[A] - \frac{k_3 \omega_1 \Delta[R^{2-}]}{K_3} - \frac{k_3 \omega_2 \Delta[A]}{K_3} - \frac{k_2 [A] \omega_1 \Delta[R^{2-}]}{[B^-]} - \frac{k_2 [A] \omega_2 \Delta[A]}{[B^-]} \\
&\quad - \frac{k_2 [A] \omega_3 \Delta[R^{2-}]}{[R]} - \frac{k_2 [A] \omega_4 \Delta[A]}{[R]} + \frac{k_2 [A] \omega_5 \Delta[R^{2-}]}{[R^-]} + \frac{k_2 [A] \omega_6 \Delta[A]}{[R^-]} + \frac{k_3 K_5 [R^-] \Delta[A]}{K_6 [R]} \\
&\quad + \frac{k_3 K_5 [A] \omega_5 \Delta[R^{2-}]}{K_6 [R]} + \frac{k_3 K_5 [A] \omega_6 \Delta[A]}{K_6 [R]} - \frac{k_3 [B^-] \omega_3 \Delta[R^{2-}]}{K_3 [R]} - \frac{k_3 [B^-] \omega_4 \Delta[A]}{K_3 [R]} \\
&= \left(k_2 - \frac{k_3 \omega_2}{K_3} - \frac{k_2 \omega_2 [A]}{[B^-]} - \frac{k_2 \omega_4 [A]}{[R]} + \frac{k_2 \omega_6 [A]}{[R^-]} + \frac{k_3 K_5 [R^-]}{K_6 [R]} + \frac{k_3 K_5 \omega_6 [A]}{K_6 [R]} - \frac{k_3 \omega_4 [B^-]}{K_3 [R]} \right) \Delta[A] \\
&\quad + \left(\frac{k_2 \omega_5 [A]}{[R^-]} - \frac{k_2 \omega_3 [A]}{[R]} + \frac{k_3 K_5 \omega_5 [A]}{K_6 [R]} - \frac{k_3 \omega_3 [B^-]}{K_3 [R]} - \frac{k_3 \omega_1}{K_3} - \frac{k_2 \omega_1 [A]}{[B^-]} \right) \Delta[R^{2-}] \\
&= \lambda_1 \Delta[R^{2-}] + \lambda_2 \Delta[A] \tag{3.7.20}
\end{aligned}$$

$$\left[\begin{array}{l} \lambda_1 \equiv \left(\frac{k_2 \omega_5 [A]}{[R^-]} - \frac{k_2 \omega_3 [A]}{[R]} + \frac{k_3 K_5 \omega_5 [A]}{K_6 [R]} - \frac{k_3 \omega_3 [B^-]}{K_3 [R]} - \frac{k_3 \omega_1}{K_3} - \frac{k_2 \omega_1 [A]}{[B^-]} \right) \\ \lambda_2 \equiv \left(k_2 - \frac{k_3 \omega_2}{K_3} - \frac{k_2 \omega_2 [A]}{[B^-]} - \frac{k_2 \omega_4 [A]}{[R]} + \frac{k_2 \omega_6 [A]}{[R^-]} + \frac{k_3 K_5 [R^-]}{K_6 [R]} + \frac{k_3 K_5 \omega_6 [A]}{K_6 [R]} - \frac{k_3 \omega_4 [B^-]}{K_3 [R]} \right) \end{array} \right]$$

By analogy to the derivation of equations 3.7.19 and 3.7.20, equation 3.5.3 becomes:

$$\begin{aligned}
\Rightarrow \frac{d^2 \Delta[A]}{dx^2} &= -\frac{D^*}{D_A} \eta_1 \Delta[R^{2-}] - \frac{D^*}{D_A} \eta_2 \Delta[A] + \frac{\lambda_1}{D_A} \Delta[R^{2-}] + \frac{\lambda_2}{D_A} \Delta[A] \\
&= \eta_3 \Delta[R^{2-}] + \eta_4 \Delta[A] \quad \left(\eta_3 \equiv \frac{\lambda_1}{D_A} - \frac{D^*}{D_A} \eta_1 \quad \eta_4 \equiv \frac{\lambda_2}{D_A} - \frac{D^*}{D_A} \eta_2 \right) \tag{3.7.21}
\end{aligned}$$

Equations 3.7.19 and 3.7.21 are, finally, linearized by setting each of the η parameters to their values at $x = 0$.

The solution to the set of two coupled linear differential equations is then assumed

to be as follows:

$$\Delta[R^{2-}] = ae^{mx} \quad (3.7.22a), \quad \Delta[A] = be^{mx} \quad (3.7.23a)$$

Thus :

$$\frac{d\Delta[R^{2-}]}{dx} = mae^{mx} \quad (3.7.22b), \quad \frac{d\Delta[A]}{dx} = mbe^{mx} \quad (3.7.23b)$$

$$\begin{cases} \frac{d^2\Delta[R^{2-}]}{dx^2} = m^2ae^{mx} = \eta_1ae^{mx} + \eta_2be^{mx} \\ \frac{d^2\Delta[A]}{dx^2} = m^2be^{mx} = \eta_3ae^{mx} + \eta_4be^{mx} \end{cases} \quad \text{So : } \begin{cases} (m^2 - \eta_1)a - \eta_2b = 0 \\ \eta_3a + (\eta_4 - m^2)b = 0 \end{cases}$$

The two linear equations in a and b are satisfied only when:

$$\begin{cases} a = \frac{\eta_2}{m^2 - \eta_1} b = \frac{m^2 - \eta_4}{\eta_3} b \equiv \varepsilon b \quad \text{and:} \\ \begin{vmatrix} (m^2 - \eta_1) & -\eta_2 \\ \eta_3 & (\eta_4 - m^2) \end{vmatrix} = 0 \end{cases}$$

It follows that $\eta_2\eta_3 - (u - \eta_4)(u - \eta_1) = 0$ where $u \equiv m^2$

$$\text{or: } u^2 - (\eta_1 + \eta_4)u + (\eta_1\eta_4 - \eta_2\eta_3) = 0$$

$$\text{There are two roots, } u_1, u_2 = \frac{(\eta_1 + \eta_4) \pm \sqrt{(\eta_1 + \eta_4)^2 - 4(\eta_1\eta_4 - \eta_2\eta_3)}}{2}$$

Accordingly, there are 4 m values: $m_1 = \sqrt{u_1}, m_2 = -m_1, m_3 = \sqrt{u_2}, m_4 = -m_3$

$$\text{and 4 } \varepsilon \text{ values: } \varepsilon_1 = \varepsilon_2 \equiv E_1 = \frac{\eta_2}{u_1 - \eta_1}; \quad \varepsilon_3 = \varepsilon_4 \equiv E_2 = \frac{\eta_2}{u_2 - \eta_1}$$

And it follows that:

$$\left\{ \begin{array}{l} \Delta[R^{2-}] = \sum_{j=1}^4 a_j e^{m_j x} = \sum_{j=1}^4 \varepsilon_j b_j e^{m_j x} \quad (3.7.22c) \\ \Delta[A] = \sum_{j=1}^4 b_j e^{m_j x} \quad (3.7.23c) \end{array} \right.$$

The two first derivatives become:

$$\left\{ \begin{array}{l} \frac{d\Delta[R^{2-}]}{dx} = \sum_{j=1}^4 \varepsilon_j m_j b_j e^{m_j x} \quad (3.7.22d) \\ \frac{d\Delta[A]}{dx} = \sum_{j=1}^4 m_j b_j e^{m_j x} \quad (3.7.23d) \end{array} \right.$$

The b's are constants of integration, which are determined by the boundary conditions:

$$\begin{aligned} \text{i) } [A]_{x=0} &= \underline{[A]}_{x=0} + \Delta[A]_{x=0} = \underline{[A]}_{x=0} + \sum_{j=1}^4 b_j \\ &\Rightarrow b_1 + b_2 + b_3 + b_4 = [A]_{x=0} - \underline{[A]}_{x=0} \end{aligned} \quad (3.7.24)$$

$$\begin{aligned} \text{ii) } \left(\frac{d[R^{2-}]}{dx} \right)_{x=0} &= 0 \\ &\Rightarrow \left(\frac{d\Delta[R^{2-}]}{dx} \right)_{x=0} = E_1 m_1 (b_1 - b_2) + E_2 m_3 (b_3 - b_4) = - \left(\frac{d[\underline{R}^{2-}]}{dx} \right)_{x=0} \end{aligned} \quad (3.7.25)$$

$$\begin{aligned} \text{iii) } \Delta[A]_{x=\delta} &= b_1 e^{m_1 \delta} + b_2 e^{-m_1 \delta} + b_3 e^{m_3 \delta} + b_4 e^{-m_3 \delta} = 0 \\ \text{iv) } \Delta[R^{2-}]_{x=\delta} &= E_1 (b_1 e^{m_1 \delta} + b_2 e^{-m_1 \delta}) + E_2 (b_3 e^{m_3 \delta} + b_4 e^{-m_3 \delta}) = 0 \end{aligned} \quad \left. \vphantom{\begin{array}{l} \text{iii) } \\ \text{iv) } \end{array}} \right\}$$

$$\Rightarrow b_1 = -b_2 e^{-2m_1 \delta}, \quad b_3 = -b_4 e^{-2m_3 \delta} \quad (3.7.26)$$

$$\Rightarrow \Delta[R^{2-}]_{x=0} = E_1 (b_1 + b_2) + E_2 (b_3 + b_4) \quad (3.7.27)$$

Equation 3.7.24 becomes:

$$b_2(1 - e^{-2m_1\delta}) + b_4(1 - e^{-2m_3\delta}) = [A]_{x=0} - \underline{[A]}_{x=0} \equiv c_1 \quad (3.7.28)$$

Equation 3.7.25 becomes:

$$E_1 m_1 b_2(1 + e^{-2m_1\delta}) + E_2 m_3 b_4(1 + e^{-2m_3\delta}) = \left(\frac{d[R^{2-}]}{dx} \right)_{x=0} \equiv c_2 \quad (3.7.29)$$

The 2 c's are constants, which are determined by the underbarred concentrations at the interface. From equations 3.7.28 and 3.7.29 it follows that:

$$\begin{aligned} b_2 &= \frac{\begin{vmatrix} c_1 & 1 - e^{-2m_3\delta} \\ c_2 & E_2 m_3 (1 + e^{-2m_3\delta}) \end{vmatrix}}{\begin{vmatrix} 1 - e^{-2m_1\delta} & 1 - e^{-2m_3\delta} \\ E_1 m_1 (1 + e^{-2m_1\delta}) & E_2 m_3 (1 + e^{-2m_3\delta}) \end{vmatrix}} \\ &= \frac{E_2 m_3 c_1 (1 + e^{-2m_3\delta}) - (1 - e^{-2m_3\delta}) c_2}{E_2 m_3 (1 + e^{-2m_3\delta}) (1 - e^{-2m_1\delta}) - E_1 m_1 (1 + e^{-2m_1\delta}) (1 - e^{-2m_3\delta})} \end{aligned} \quad (3.7.30)$$

$$\begin{aligned} b_4 &= \frac{\begin{vmatrix} 1 - e^{-2m_1\delta} & c_1 \\ E_1 m_1 (1 + e^{-2m_1\delta}) & c_2 \end{vmatrix}}{\begin{vmatrix} 1 - e^{-2m_1\delta} & 1 - e^{-2m_3\delta} \\ E_1 m_1 (1 + e^{-2m_1\delta}) & E_2 m_3 (1 + e^{-2m_3\delta}) \end{vmatrix}} \\ &= \frac{(1 - e^{-2m_1\delta}) c_2 - E_1 m_1 c_1 (1 + e^{-2m_1\delta})}{E_2 m_3 (1 + e^{-2m_3\delta}) (1 - e^{-2m_1\delta}) - E_1 m_1 (1 + e^{-2m_1\delta}) (1 - e^{-2m_3\delta})} \end{aligned} \quad (3.7.31)$$

Therefore:

$$\begin{aligned}
\Delta[A] &= \frac{\left[E_2 m_3 c_1 (1 + e^{-2m_3 \delta}) - (1 - e^{-2m_3 \delta}) c_2 \right] (e^{-m_1 x} - e^{-2m_1 \delta} e^{m_1 x})}{E_2 m_3 (1 + e^{-2m_3 \delta}) (1 - e^{-2m_1 \delta}) - E_1 m_1 (1 + e^{-2m_1 \delta}) (1 - e^{-2m_3 \delta})} \\
&\quad + \frac{\left[(1 - e^{-2m_1 \delta}) c_2 - E_1 m_1 c_1 (1 + e^{-2m_1 \delta}) \right] (e^{-m_3 x} - e^{-2m_3 \delta} e^{m_3 x})}{E_2 m_3 (1 + e^{-2m_3 \delta}) (1 - e^{-2m_1 \delta}) - E_1 m_1 (1 + e^{-2m_1 \delta}) (1 - e^{-2m_3 \delta})} \\
&= \frac{\left[E_2 m_3 c_1 (e^{m_3 \delta} + e^{-m_3 \delta}) - c_2 (e^{m_3 \delta} - e^{-m_3 \delta}) \right] \left[e^{m_1 (\delta - x)} - e^{-m_1 (\delta - x)} \right]}{E_2 m_3 (e^{m_3 \delta} + e^{-m_3 \delta}) (e^{m_1 \delta} - e^{-m_1 \delta}) - E_1 m_1 (e^{m_1 \delta} + e^{-m_1 \delta}) (e^{m_3 \delta} - e^{-m_3 \delta})} \\
&\quad + \frac{\left[(e^{m_1 \delta} - e^{-m_1 \delta}) c_2 - E_1 m_1 c_1 (e^{m_1 \delta} + e^{-m_1 \delta}) \right] \left[e^{m_3 (\delta - x)} - e^{-m_3 (\delta - x)} \right]}{E_2 m_3 (e^{m_3 \delta} + e^{-m_3 \delta}) (e^{m_1 \delta} - e^{-m_1 \delta}) - E_1 m_1 (e^{m_1 \delta} + e^{-m_1 \delta}) (e^{m_3 \delta} - e^{-m_3 \delta})} \\
&= \frac{\left[E_2 m_3 c_1 \cosh(m_3 \delta) - c_2 \sinh(m_3 \delta) \right] \sinh[m_1 (\delta - x)]}{E_2 m_3 \sinh(m_1 \delta) \cosh(m_3 \delta) - E_1 m_1 \sinh(m_3 \delta) \cosh(m_1 \delta)} \\
&\quad + \frac{\left[c_2 \sinh(m_1 \delta) - E_1 m_1 c_1 \cosh(m_1 \delta) \right] \sinh[m_3 (\delta - x)]}{E_2 m_3 \sinh(m_1 \delta) \cosh(m_3 \delta) - E_1 m_1 \sinh(m_3 \delta) \cosh(m_1 \delta)} \\
&= \frac{\left[E_2 m_3 c_1 - c_2 \tanh(m_3 \delta) \right] \sinh[m_1 (\delta - x)]}{E_2 m_3 \sinh(m_1 \delta) - E_1 m_1 \tanh(m_3 \delta) \cosh(m_1 \delta)} \\
&\quad + \frac{\left[c_2 \tanh(m_1 \delta) - E_1 m_1 c_1 \right] \sinh[m_3 (\delta - x)]}{E_2 m_3 \tanh(m_1 \delta) \cosh(m_3 \delta) - E_1 m_1 \sinh(m_3 \delta)} \tag{3.7.32}
\end{aligned}$$

Accordingly:

$$\begin{aligned}
\Delta[A]_{x=0} &= \frac{\left[E_2 m_3 c_1 - c_2 \tanh(m_3 \delta) \right] \sinh(m_1 \delta)}{E_2 m_3 \sinh(m_1 \delta) - E_1 m_1 \tanh(m_3 \delta) \cosh(m_1 \delta)} \\
&\quad + \frac{\left[c_2 \tanh(m_1 \delta) - E_1 m_1 c_1 \right] \sinh(m_3 \delta)}{E_2 m_3 \tanh(m_1 \delta) \cosh(m_3 \delta) - E_1 m_1 \sinh(m_3 \delta)}
\end{aligned}$$

$$\begin{aligned}
&= \frac{[E_2 m_3 c_1 - c_2 \tanh(m_3 \delta)] \tanh(m_1 \delta)}{E_2 m_3 \tanh(m_1 \delta) - E_1 m_1 \tanh(m_3 \delta)} + \frac{[c_2 \tanh(m_1 \delta) - E_1 m_1 c_1] \tanh(m_3 \delta)}{E_2 m_3 \tanh(m_1 \delta) - E_1 m_1 \tanh(m_3 \delta)} \\
&= \frac{[E_2 m_3 c_1 - c_2 \tanh(m_3 \delta)] \tanh(m_1 \delta) + [c_2 \tanh(m_1 \delta) - E_1 m_1 c_1] \tanh(m_3 \delta)}{E_2 m_3 \tanh(m_1 \delta) - E_1 m_1 \tanh(m_3 \delta)} \quad (3.7.33)
\end{aligned}$$

$$\begin{aligned}
\frac{d\Delta[A]}{dx} &= -\frac{m_1 [E_2 m_3 c_1 - c_2 \tanh(m_3 \delta)] \cosh[m_1 (\delta - x)]}{E_2 m_3 \sinh(m_1 \delta) - E_1 m_1 \tanh(m_3 \delta) \cosh(m_1 \delta)} \\
&\quad - \frac{m_3 [c_2 \tanh(m_1 \delta) - E_1 m_1 c_1] \cosh[m_3 (\delta - x)]}{E_2 m_3 \tanh(m_1 \delta) \cosh(m_3 \delta) - E_1 m_1 \sinh(m_3 \delta)} \quad (3.7.34)
\end{aligned}$$

$$\begin{aligned}
\left(\frac{d\Delta[A]}{dx} \right)_{x=0} &= -\frac{m_1 [E_2 m_3 c_1 - c_2 \tanh(m_3 \delta)] \cosh(m_1 \delta)}{E_2 m_3 \sinh(m_1 \delta) - E_1 m_1 \tanh(m_3 \delta) \cosh(m_1 \delta)} \\
&\quad - \frac{m_3 [c_2 \tanh(m_1 \delta) - E_1 m_1 c_1] \cosh(m_3 \delta)}{E_2 m_3 \tanh(m_1 \delta) \cosh(m_3 \delta) - E_1 m_1 \sinh(m_3 \delta)} \\
&= -\frac{m_1 [E_2 m_3 c_1 - c_2 \tanh(m_3 \delta)]}{E_2 m_3 \tanh(m_1 \delta) - E_1 m_1 \tanh(m_3 \delta)} - \frac{m_3 [c_2 \tanh(m_1 \delta) - E_1 m_1 c_1]}{E_2 m_3 \tanh(m_1 \delta) - E_1 m_1 \tanh(m_3 \delta)} \\
&= -\frac{m_1 [E_2 m_3 c_1 - c_2 \tanh(m_3 \delta)] + m_3 [c_2 \tanh(m_1 \delta) - E_1 m_1 c_1]}{E_2 m_3 \tanh(m_1 \delta) - E_1 m_1 \tanh(m_3 \delta)} \quad (3.7.35)
\end{aligned}$$

$$\begin{aligned}
\frac{d^2 \Delta[A]}{dx^2} &= \frac{m_1^2 [E_2 m_3 c_1 - c_2 \tanh(m_3 \delta)] \sinh[m_1 (\delta - x)]}{E_2 m_3 \sinh(m_1 \delta) - E_1 m_1 \tanh(m_3 \delta) \cosh(m_1 \delta)} \\
&\quad + \frac{m_3^2 [c_2 \tanh(m_1 \delta) - E_1 m_1 c_1] \sinh[m_3 (\delta - x)]}{E_2 m_3 \tanh(m_1 \delta) \cosh(m_3 \delta) - E_1 m_1 \sinh(m_3 \delta)} \quad (3.7.36)
\end{aligned}$$

Next we proceed to derive the derivatives of the underbarred concentrations:

Differentiation of equations 3.7.4-5, 3.7.7-8 and 3.7.14, and cancel out $d[\underline{C}^{2-}]$,

gives:

$$d[\underline{R}^-] + d[\underline{R}] + d[\underline{R}^{2-}] = 0 \Rightarrow d[\underline{R}] = -d[\underline{R}^-] - d[\underline{R}^{2-}] \quad (3.7.37)$$

$$d[\underline{B}^-] + d[\underline{R}^-] + 2d[\underline{R}^{2-}] + 2d[\underline{C}^{2-}] = 0 \quad (3.7.38)$$

$$[\underline{R}^{2-}]d[\underline{R}] + [\underline{R}]d[\underline{R}^{2-}] = \frac{2K_{\text{Carb}}[\underline{A}][\underline{R}^-]}{K_6}d[\underline{R}^-] + \frac{K_{\text{Carb}}[\underline{R}^-]^2}{K_6}d[\underline{A}] \quad (3.7.39)$$

$$[\underline{B}^-]d[\underline{R}] + [\underline{R}]d[\underline{B}^-] = \frac{K_2[\underline{A}]}{K_6}d[\underline{R}^-] + \frac{K_2[\underline{R}^-]}{K_6}d[\underline{A}] \quad (3.7.40)$$

$$d[\underline{C}^{2-}] = \frac{K_4[\underline{B}^-]}{K_6[\underline{R}]}d[\underline{R}^-] + \frac{K_4[\underline{R}^-]}{K_6[\underline{R}]}d[\underline{B}^-] - \frac{K_4[\underline{R}^-][\underline{B}^-]}{K_6[\underline{R}]^2}d[\underline{R}] \quad (3.7.41)$$

$$\Rightarrow \left\{ \begin{array}{l} \left(1 + \frac{2K_4[\underline{R}^-]}{K_6[\underline{R}]}\right)d[\underline{B}^-] + \left(1 + \frac{2K_4[\underline{B}^-]}{K_6[\underline{R}]} + \frac{2K_4[\underline{R}^-][\underline{B}^-]}{K_6[\underline{R}]^2}\right)d[\underline{R}^-] \\ \quad + \left(2 + \frac{2K_4[\underline{R}^-][\underline{B}^-]}{K_6[\underline{R}]^2}\right)d[\underline{R}^{2-}] = 0 \\ \left([\underline{R}^{2-}] + \frac{2K_{\text{Carb}}[\underline{A}][\underline{R}^-]}{K_6}\right)d[\underline{R}^-] + ([\underline{R}^{2-}] - [\underline{R}])d[\underline{R}^{2-}] = -\frac{K_{\text{Carb}}[\underline{R}^-]^2}{K_6}d[\underline{A}] \\ -[\underline{R}]d[\underline{B}^-] + \left([\underline{B}^-] + \frac{K_2[\underline{A}]}{K_6}\right)d[\underline{R}^-] + [\underline{B}^-]d[\underline{R}^{2-}] = -\frac{K_2[\underline{R}^-]}{K_6}d[\underline{A}] \end{array} \right.$$

$$d[\underline{B}^-] = \frac{\theta_2}{\theta_1} d[\underline{A}] \quad (3.7.42)$$

$$d[\underline{R}^-] = \frac{\theta_3}{\theta_1} d[\underline{A}] \quad (3.7.43)$$

$$\Rightarrow d[\underline{R}^{2-}] = \frac{\theta_4}{\theta_1} d[\underline{A}] \quad (3.7.44)$$

$$d[\underline{R}] = -\frac{\theta_3 + \theta_4}{\theta_1} d[\underline{A}] \quad (3.7.45)$$

$$d[\underline{C}^{2-}] = \frac{\theta_5}{\theta_1} d[\underline{A}] \quad (3.7.46)$$

$$\text{where } \theta_1 \equiv \begin{vmatrix} \left(1 + \frac{2K_4[\underline{R}^-]}{K_6[\underline{R}]}\right) & \left(1 + \frac{2K_4[\underline{B}^-]}{K_6[\underline{R}]} + \frac{2K_4[\underline{R}^-][\underline{B}^-]}{K_6[\underline{R}]^2}\right) & \left(2 + \frac{2K_4[\underline{R}^-][\underline{B}^-]}{K_6[\underline{R}]^2}\right) \\ 0 & \left([\underline{R}^{2-}] + \frac{2K_{\text{Carb}}[\underline{A}][\underline{R}^-]}{K_6}\right) & ([\underline{R}^{2-}] - [\underline{R}]) \\ -[\underline{R}] & \left([\underline{B}^-] + \frac{K_2[\underline{A}]}{K_6}\right) & [\underline{B}^-] \end{vmatrix}$$

$$\theta_2 \equiv \begin{vmatrix} 0 & \left(1 + \frac{2K_4[\underline{B}^-]}{K_6[\underline{R}]} + \frac{2K_4[\underline{R}^-][\underline{B}^-]}{K_6[\underline{R}]^2}\right) & \left(2 + \frac{2K_4[\underline{R}^-][\underline{B}^-]}{K_6[\underline{R}]^2}\right) \\ \left(-\frac{K_{\text{carb}}[\underline{R}^-]^2}{K_6}\right) & \left([\underline{R}^{2-}] + \frac{2K_{\text{Carb}}[\underline{A}][\underline{R}^-]}{K_6}\right) & ([\underline{R}^{2-}] - [\underline{R}]) \\ \left(-\frac{K_2[\underline{R}^-]}{K_6}\right) & \left([\underline{B}^-] + \frac{K_2[\underline{A}]}{K_6}\right) & [\underline{B}^-] \end{vmatrix}$$

$$\theta_3 \equiv \begin{vmatrix} \left(1 + \frac{2K_4[\underline{R}^-]}{K_6[\underline{R}]}\right) & 0 & \left(2 + \frac{2K_4[\underline{R}^-][\underline{B}^-]}{K_6[\underline{R}]^2}\right) \\ 0 & \left(-\frac{K_{\text{carb}}[\underline{R}^-]^2}{K_6}\right) & ([\underline{R}^{2-}] - [\underline{R}]) \\ -[\underline{R}] & \left(-\frac{K_2[\underline{R}^-]}{K_6}\right) & [\underline{B}^-] \end{vmatrix}$$

$$\theta_4 \equiv \begin{vmatrix} \left(1 + \frac{2K_4[\underline{R}^-]}{K_6[\underline{R}]}\right) & \left(1 + \frac{2K_4[\underline{B}^-]}{K_6[\underline{R}]} + \frac{2K_4[\underline{R}^-][\underline{B}^-]}{K_6[\underline{R}]^2}\right) & 0 \\ 0 & \left([\underline{R}^{2-}] + \frac{2K_{\text{carb}}[\underline{A}][\underline{R}^-]}{K_6}\right) & \left(-\frac{K_{\text{carb}}[\underline{R}^-]^2}{K_6}\right) \\ -[\underline{R}] & \left([\underline{B}^-] + \frac{K_2[\underline{A}]}{K_6}\right) & \left(-\frac{K_2[\underline{R}^-]}{K_6}\right) \end{vmatrix}$$

$$\theta_5 \equiv \frac{K_4[\underline{B}^-]}{K_6[\underline{R}]} \theta_3 + \frac{K_4[\underline{R}^-]}{K_6[\underline{R}]} \theta_2 + \frac{K_4[\underline{R}^-][\underline{B}^-]}{K_6[\underline{R}]^2} (\theta_3 + \theta_4)$$

Differentiation of equation 3.7.6 and insertion of equations 3.7.42, 3.7.44 and 3.7.46 gives:

$$D_A d[\underline{A}] + D^* \left(\frac{\theta_2 + \theta_4 + \theta_5}{\theta_1} \right) d[\underline{A}] = -\Phi_1 dx$$

or :

$$\frac{d[\underline{A}]}{dx} = -\frac{\theta_1 \Phi_1}{\theta_1 D_A + D^* (\theta_2 + \theta_4 + \theta_5)} \quad (3.7.47)$$

It follows that:

$$\frac{d[\underline{R}^{2-}]}{dx} = -\frac{\theta_4 \Phi_1}{\theta_1 D_A + D^* (\theta_2 + \theta_4 + \theta_5)} \quad (3.7.48)$$

The solution is implemented as a search for $\underline{[A]}_{x=0}$:

- 1) a guess of $\underline{[A]}_{x=0}$ is made;
- 2) all other underbar quantities at $x = 0$ are calculated from equations 3.7.4-5, 3.7.7-8, and 3.7.14;
- 3) Φ_1 follows from equation 3.7.6 and the boundary conditions:

$$\Phi_1 = \frac{D_A \left(\underline{[A]}_{x=0} - \underline{[A]}_{x=\delta} \right) + D^* \left(\underline{[B^-]}_{x=0} - \underline{[B^-]}_{x=\delta} + \underline{[R^{2-}]}_{x=0} - \underline{[R^{2-}]}_{x=\delta} + \underline{[C^{2-}]}_{x=0} - \underline{[C^{2-}]}_{x=\delta} \right)}{\delta} \quad (3.7.49);$$

- 4) values of $\theta_1 - \theta_5$, $\omega_1 - \omega_6$, λ_1 , λ_2 , $\eta_1 - \eta_4$, u_1 , u_2 , $m_1 - m_4$, E_1 , and E_2 at $x = 0$ are calculated;
- 5) $\Delta[A]_{x=0} = \alpha p_{A,G} - \underline{[A]}_{x=0}$, from which c_1 follows;

- 6) $\left(\frac{d[A]}{dx} \right)_{x=0}$ and $\left(\frac{d[R^{2-}]}{dx} \right)_{x=0}$ are calculated from equations 3.7.47- 48, from which c_2 follows;

- 7) $\left(\frac{d\Delta[A]}{dx} \right)_{x=0}$ is calculated from equation 3.7.35;

- 8) $\Phi_1 = -D_A \left[\left(\frac{d[A]}{dx} \right)_{x=0} + \left(\frac{d\Delta[A]}{dx} \right)_{x=0} \right]$ is compared with the value calculated in step 3).

3.8 Numerical validation

An exact numerical solution to the same differential equations provides a test of the above analysis. The two second order differential equations will be transformed into a set of first order differential equations and solved as an initial value problem.

As a first step, $[B^-]$, $[R]$, $[R^-]$ and $[C^{2-}]$ are expressed as functions of $[A]$, $[R^{2-}]$, y and Φ_1 :

$$D_A[A] + D^*([B^-] + [R^{2-}] + [C^{2-}]) = \Phi_1 \delta(1-y) + \xi \quad (x \equiv \delta y, y = [0,1])$$

$$(\xi \equiv D_A[A]_{x=\delta} + D^*([B^-]_{x=\delta} + [R^{2-}]_{x=\delta} + [C^{2-}]_{x=\delta}))$$

Thus :

$$[B^-] + [R^{2-}] + [C^{2-}] = \frac{\Phi_1 \delta(1-y) + \xi - D_A[A]}{D^*} \equiv Q \quad (3.8.1)$$

$$\begin{cases} [R] + [R^-] + [R^{2-}] = [K^+] \\ [B^-] + [R^-] + 2[R^{2-}] + 2[C^{2-}] = [K^+] \\ [B^-] + [R^{2-}] + [C^{2-}] = Q \end{cases} \quad \text{Thus :} \quad \begin{cases} [C^{2-}] = Q - [R^{2-}] - [B^-] \\ [R^-] = [K^+] - 2Q + [B^-] \\ [R] = 2Q - [B^-] - [R^{2-}] \end{cases} \quad (3.8.2)$$

$$(3.8.3)$$

$$(3.8.4)$$

$$\frac{[C^{2-}][R]}{[B^-][R^-]} = \frac{K_4}{K_6}$$

It follows from the last three equations that:

$$(Q - [R^{2-}] - [B^-])(2Q - [B^-] - [R^{2-}]) = \frac{K_4}{K_6} [B^-] ([K^+] - 2Q + [B^-])$$

$$(2Q^2 - 3Q[R^{2-}] + [R^{2-}]^2) + (2[R^{2-}] - 3Q)[B^-] + [B^-]^2 = \frac{K_4([K^+] - 2Q)}{K_6} [B^-] + \frac{K_4}{K_6} [B^-]^2$$

$$\left(1 - \frac{K_4}{K_6}\right) [B^-]^2 + \left(2[R^{2-}] - 3Q - \frac{K_4([K^+] - 2Q)}{K_6}\right) [B^-] + (2Q^2 - 3Q[R^{2-}] + [R^{2-}]^2) = 0$$

Thus,

$$\left(1 - \frac{K_4}{K_6}\right)[B^-]^2 + G[B^-] + J = 0 \quad (3.8.5)$$

where :

$$G \equiv 2[R^{2-}] - 3Q - \frac{K_4([K^+] - 2Q)}{K_6}$$

$$J \equiv 2Q^2 - 3Q[R^{2-}] + [R^{2-}]^2$$

Therefore :

$$[B^-] = \frac{-G - \sqrt{G^2 - 4\left(1 - \frac{K_4}{K_6}\right)J}}{2\left(1 - \frac{K_4}{K_6}\right)} \quad (3.8.6)$$

Note that the other root for $[B^-]$, $\frac{-G + \sqrt{G^2 - 4\left(1 - \frac{K_4}{K_6}\right)J}}{2\left(1 - \frac{K_4}{K_6}\right)}$, turns out to be physically impossible.

Now we return to the differential equations:

$$\begin{aligned} \frac{d^2[A]}{dy^2} = & \frac{\delta^2 k_{1a}[A][R^-] \left(\frac{k_{R^-}}{k_{-1a}}[R^-] + \frac{k_w}{k_{-1a}}[W] \right) \left(1 - \frac{K_6[R^{2-}][R]}{K_{\text{Carb}}[A][R^-]^2} \right)}{D_A \left(1 + \frac{k_{R^-}}{k_{-1a}}[R^-] + \frac{k_w}{k_{-1a}}[W] \right)} \\ & + \frac{\delta^2 k_2[A]}{D_A} - \frac{\delta^2 k_2 K_6 [B^-][R]}{D_A K_2 [R^-]} + \frac{\delta^2 k_3 K_5 [A][R^-]}{D_A K_6 [R]} - \frac{\delta^2 k_3 [B^-]}{D_A K_3} \end{aligned} \quad (3.5.3)$$

$$\frac{d^2[R^{2-}]}{dy^2} = - \frac{\delta^2 k_{1a}[A][R^-] \left(\frac{k_{R^-}}{k_{-1a}}[R^-] + \frac{k_w}{k_{-1a}}[W] \right) \left(1 - \frac{K_6[R^{2-}][R]}{K_{\text{Carb}}[A][R^-]^2} \right)}{D^* \left(1 + \frac{k_{R^-}}{k_{-1a}}[R^-] + \frac{k_w}{k_{-1a}}[W] \right)} \quad (3.7.1)$$

Both $\frac{d^2[A]}{dy^2}$ and $\frac{d^2[R^{2-}]}{dy^2}$ are now expressible as functions of $[A]$, $[R^{2-}]$, y and Φ_1 .

We next make the following definitions:

$$x_1 = [A] \quad (3.8.7)$$

$$x_2 = \frac{d[A]}{dy} \quad (3.8.8)$$

$$x_3 = [R^{2-}] \quad (3.8.9)$$

$$x_4 = \frac{d[R^{2-}]}{dy} \quad (3.8.10)$$

It follows that:

$$\frac{dx_1}{dy} = x_2$$

$$\begin{aligned} \frac{dx_2}{dy} = & \frac{\delta^2 k_{1a} x_1 [R^-] \left(\frac{k_{R^-}}{k_{-1a}} [R^-] + \frac{k_w}{k_{-1a}} [W] \right) \left(1 - \frac{K_6 x_3 [R]}{K_{\text{Carb}} x_1 [R^-]^2} \right)}{D_A \left(1 + \frac{k_{R^-}}{k_{-1a}} [R^-] + \frac{k_w}{k_{-1a}} [W] \right)} \\ & + \frac{\delta^2 k_2 x_1}{D_A} - \frac{\delta^2 k_2 K_6 [B^-] [R]}{D_A K_2 [R^-]} + \frac{\delta^2 k_3 K_5 x_1 [R^-]}{D_A K_6 [R]} - \frac{\delta^2 k_3 [B^-]}{D_A K_3} \end{aligned}$$

$$\frac{dx_3}{dy} = x_4$$

$$\frac{dx_4}{dy} = - \frac{\delta^2 k_{1a} x_1 [R^-] \left(\frac{k_{R^-}}{k_{-1a}} [R^-] + \frac{k_w}{k_{-1a}} [W] \right) \left(1 - \frac{K_6 x_3 [R]}{K_{\text{Carb}} x_1 [R^-]^2} \right)}{D^* \left(1 + \frac{k_{R^-}}{k_{-1a}} [R^-] + \frac{k_w}{k_{-1a}} [W] \right)}$$

The boundary conditions are:

$$\begin{aligned}x_1(0) &= \alpha p_{A,G} & x_1(\delta) &= [A]_{x=\delta} \\x_2(0) &= -\frac{\Phi_1 \delta}{D_A} & x_3(\delta) &= [R^{2-}]_{x=\delta} \\x_4(0) &= 0\end{aligned}$$

Since the boundary conditions refer to two different positions, a solution must be obtained by trial and error:

Guess Φ_1 and $[R^{2-}]_{x=0}$, then solve the above four first-order ODE's numerically. If $x_1(\delta) = [A]_{x=\delta}$ and $x_3(\delta) = [R^{2-}]_{x=\delta}$, the initial guess is correct. Once $[R^{2-}]_{x=0}$ and Φ_1 are known, all the rest concentrations can be calculated.

4. RESULTS FOR CO₂-PG SYSTEM

4.1 Physiochemical parameters

The temperature and concentration dependence of solubility, density, and diffusivity, as well as equilibrium and kinetic parameters is discussed below.

4.1.1 Solubility data

Since the partial pressures of CO₂ considered are generally less than 1 atm, the vapor phase may be considered ideal and vapor-liquid equilibrium may be described by the Henry law:

$$p_{A,G} \equiv \frac{1}{\alpha} [A] = H_{CO_2} [A] \quad (4.1.1.1)$$

where $p_{A,G}$ is CO₂ partial pressure and H_{CO_2} is the Henry's law coefficient of CO₂ in the PG solution given by Portugal, Versteeg and van Swaaij [77, 78]:

$$\log \left(\frac{H_{CO_2}}{H_{CO_2,w}} \right) = K[PG]_0 \quad (4.1.1.2)$$

$$\text{where } K = \frac{62.183098}{T} - 0.111175 \text{ (mol}^{-1} \text{ dm}^3\text{)}$$

$$H_{CO_2,w} = \frac{\exp(-2044/T)}{3.54 \times 10^{-7}} \text{ (Pa mol}^{-1} \text{ m}^3\text{)}$$

4.1.2 Density data

Densities of PG solutions were measured by Portugal et al. [77], and are tabulated in Table 4.1.2.

Table 4.1.2 Densities of PG solutions [77]

T (K)	ρ (kg m ⁻³)			
	293	298	303	313
[PG] (M)				
0	998.29	997.13	995.71	992.25
0.102	1004.37	1003.06	1001.59	997.28
0.296	1015.97	1014.56	1013.00	1001.77
0.594	1033.28	1031.69	1030.02	1025.15
1.003	1056.57	1054.81	1052.98	1047.94
1.992	1112.29	1110.13	1108.01	1102.34
2.984	1163.85	1161.44	1159.07	1150.37

4.1.3 Diffusivity data

Diffusivities of CO₂ and ionic species in aqueous PG solutions are as follows

[77-79]:

$$D_A = [2.35 \times 10^{-6} \exp(-2119/T)](1 + 0.2109[\text{PG}] + 0.05124[\text{PG}]^2)^{-0.48} (\text{m}^2 \text{ s}^{-1}) \quad (4.1.3.1)$$

$$D^* = D_{\text{PG}} = (-2.412 - 9.403 \times 10^{-3}T + 7.110 \times 10^{-5}T^2 - 0.2177[\text{PG}] - 5.447 \times 10^{-2}[\text{PG}]^2 + 1.296 \times 10^{-3}T[\text{PG}]) \times 10^{-9} (\text{m}^2 \text{ s}^{-1}) \quad (4.1.3.2)$$

4.1.4 Reaction equilibrium data

Benamor and Aroua [80] provide the following data:

For CO₂ hydration, equation 3.1.2:

$$K_2 = \left(\frac{[B^-][H^+]}{[A]} \right)_{\text{eq}} = \exp\left(-\frac{12092.1}{T} - 36.7816 \ln T + 235.482\right) \quad (\text{mol dm}^{-3}) \quad (4.1.4.1)$$

For carbonate formation, equation 3.1.4:

$$K_4 = \frac{[C^{2-}][H^+]}{[B^-]} = \exp\left(-\frac{12431.7}{T} - 35.4819 \ln T + 220.067\right) \quad (\text{mol dm}^{-3}) \quad (4.1.4.2)$$

For water's ionization, equation 3.1.5, the constant $K_5 \left(= \frac{K_2}{K_3} \right)$ varies as follows:

$$K_5 = [OH^-][H^+] = \exp\left(-\frac{13445.9}{T} - 22.4773 \ln T + 140.932\right) \quad (\text{mol dm}^{-3})^2 \quad (4.1.4.3)$$

For carbamate zwitterion deprotonation, equation 3.1.6, K_6 is given by Perrin [81]:

$$K_6 = \frac{[R^-][H^+]}{[R]} = \exp(-0.000237956T^2 + 0.202203T - 61.6499) \quad (\text{mol dm}^{-3}) \quad (4.1.4.4)$$

The di-carbamate hydrolysis reaction is not treated as an independent reaction in the analysis here, since it is a linear combination of the di-carbamate and carbonate formation reactions. Its equilibrium constant is given by Jensen [82] as:

$$K_{\text{carbohydrolysis}} = \left(\frac{[R^-][B^-]}{[R^{2-}]} \right)_{\text{eq}} = \exp\left(\frac{-2767.18}{T} + 6.10312\right) \quad (\text{mol dm}^{-3}) \quad (4.1.4.5)$$

It follows from the definition, $K_{\text{Acid}_j} \equiv \left(\frac{[B_j H^+]}{[B_j][H^+]} \right)_{\text{eq}}$, that:

$$K_{\text{Acid}, R^-} = \frac{1}{K_6} \quad (4.1.4.6)$$

and:

$$K_{\text{Acid}, W} = \frac{1}{[W]} \quad (4.1.4.7)$$

From the definition, $K_{\text{Carb}} \equiv \left(\frac{[\text{R}^{2-}][\text{H}^+]}{[\text{A}][\text{R}^-]} \right)^{\text{eq}}$, it follows that:

$$K_{\text{Carb}} = \frac{K_2}{K_{\text{carbohydrolysis}}} \quad (4.1.4.8)$$

4.1.5 Kinetic data

The zwitterion mechanism rate constants are given as follows by Portugal et al.

[77], but do not account for ionic strength:

$$k_{\text{Ia}} = 2.81 \times 10^{10} \exp(-5800/T) \quad (\text{m}^3 \text{ mol}^{-1} \text{ s}^{-1}) \quad (4.1.5.1)$$

$$k_{\text{W}} / k_{\text{-Ia}} = 1.05 \times 10^{-4} \exp(-1265/T) \quad (\text{m}^3 \text{ mol}^{-1}) \quad (4.1.5.2)$$

$$k_{\text{R}^-} / k_{\text{-Ia}} = 4.89 \times 10^3 \exp(-5307/T) \quad (\text{m}^3 \text{ mol}^{-1}) \quad (4.1.5.3)$$

The forward rate constant of carbonic acid formation (at zero ionic strength) is given by Danckwerts and Sharma [20] as:

$$\log k_2 = 329.850 - 110.541 \log T - 17265.4/T \quad (k_2 [=] \text{s}^{-1}) \quad (4.1.5.4)$$

The forward rate constant of bicarbonate formation (at zero ionic strength), is given by Danckwerts and Sharma [20] as:

$$\log k_3 = 13.635 - 2895/T \quad (k_3 [=] \text{dm}^3 \text{ mol}^{-1} \text{ s}^{-1}) \quad (4.1.5.5)$$

4.2 Results and discussion

4.2.1 Speciation profiles

Figures 4.2.1.1-4 depict equilibrium speciation profiles in 1 M and 3 M PG at 293 K and 313 K, respectively, calculated from equations 3.1.4, 3.1.6, 3.4.1, 3.5.1, 3.5.5 and 3.6.4 at equilibrium.

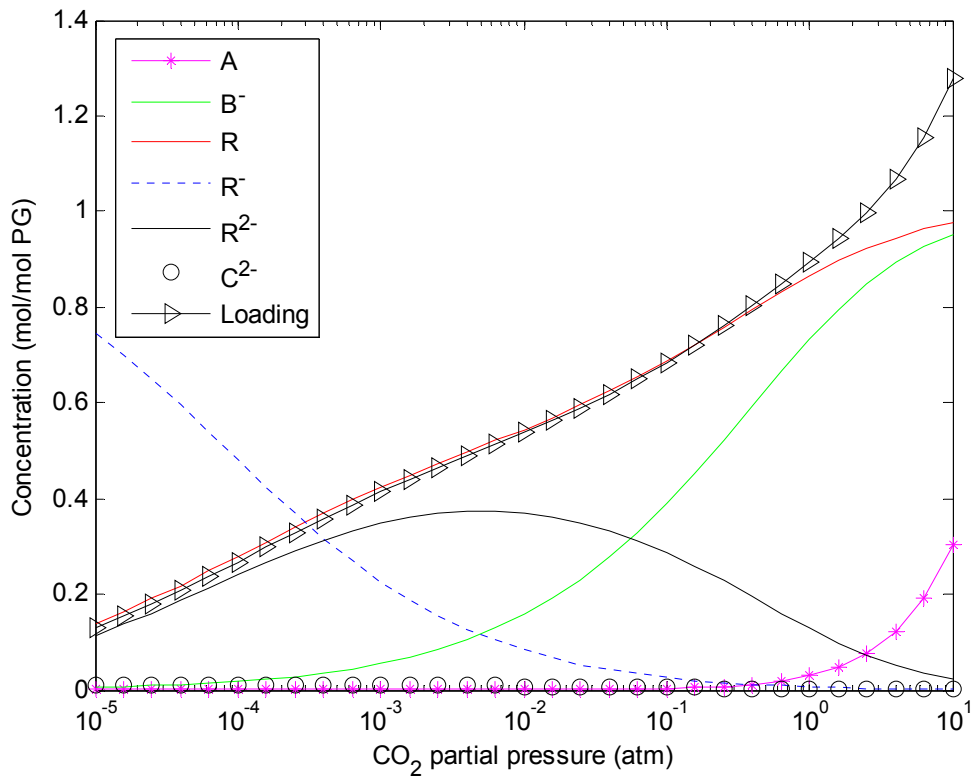


Figure 4.2.1.1 Speciation and CO₂ loading vs. CO₂ partial pressure (1 M PG, 293 K).

At CO₂ partial pressures less than 10⁻⁴ atm, dissolved CO₂ combines primarily with R⁻ to form equal proportions of R and R²⁻. At higher partial pressures, CO₂ forms mainly bicarbonate ions via CO₂ hydration reactions. Until it is exhausted, R⁻ buffers hydrogen ions co-produced with bicarbonate ions, forming the carbamate zwitterions R. Therefore,

at higher CO₂ partial pressures, the dominant overall reaction is:



After the free glycinate anion is depleted, the hydrogen ions combine with di-carbamate ions in reverse reaction 3.1.1, which explains the peak in R²⁻. The overall reaction becomes:

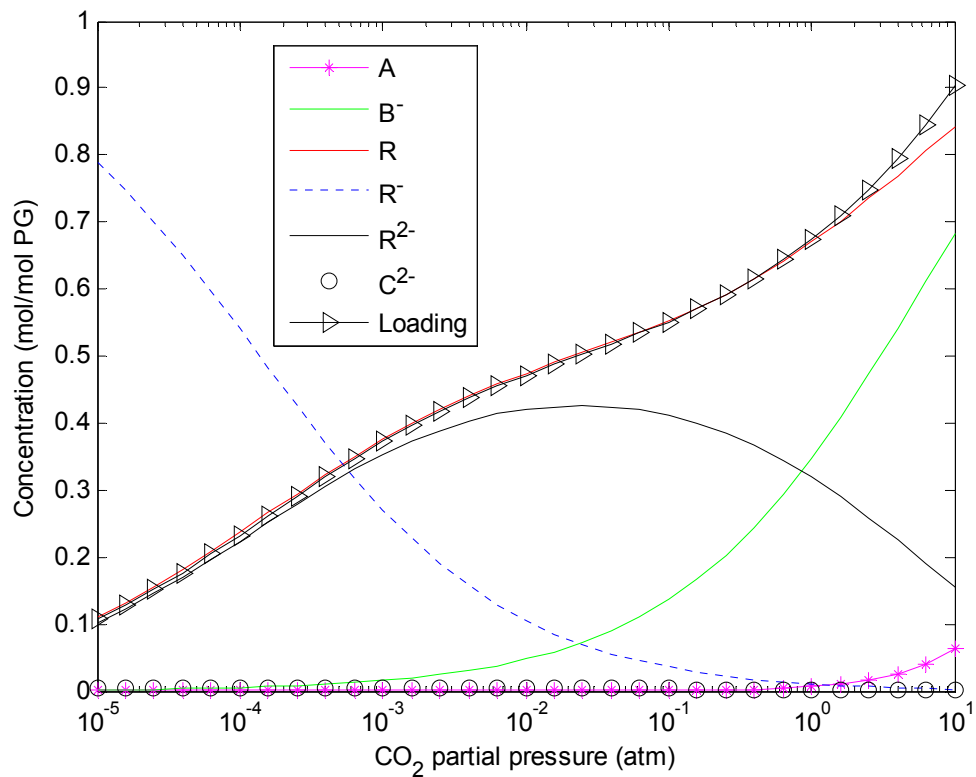
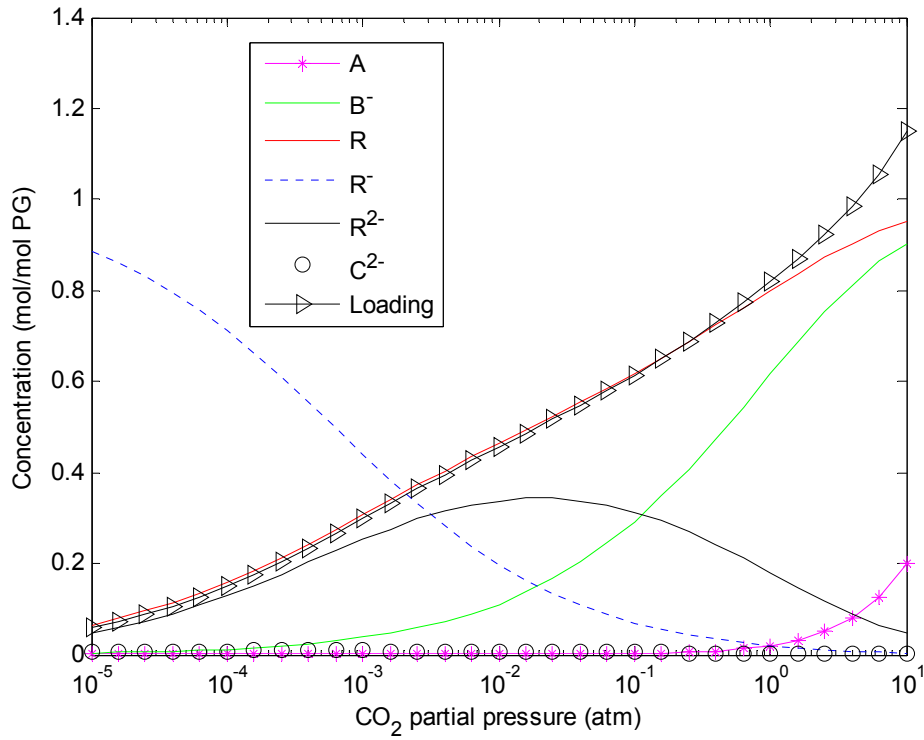
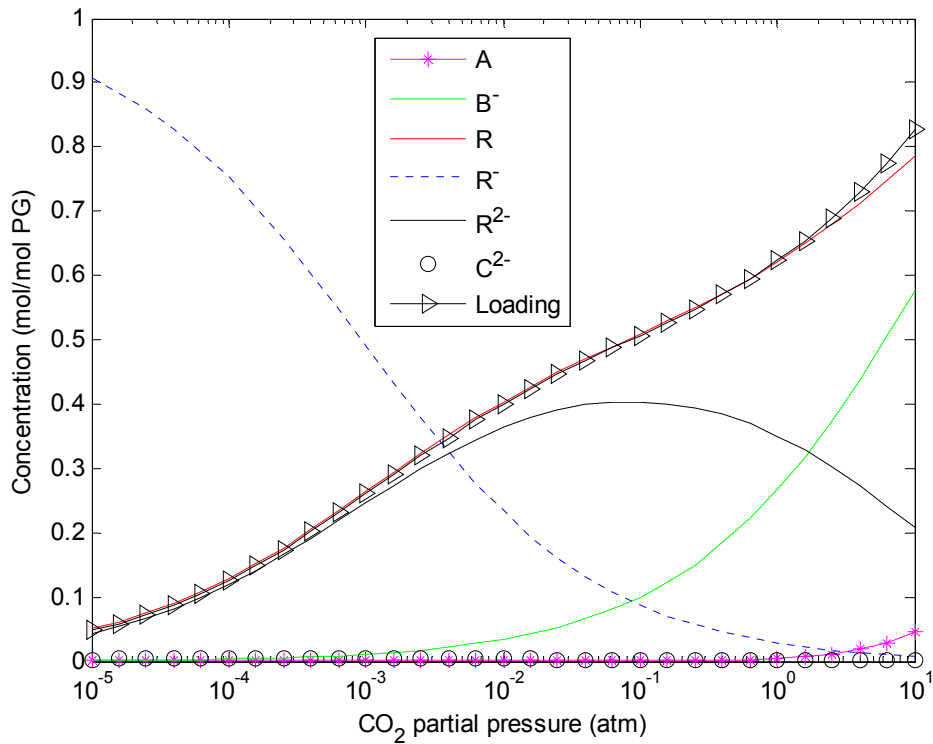


Figure 4.2.1.2 Speciation and CO₂ loading vs. CO₂ partial pressure (3 M PG, 293 K).

Figure 4.2.1.3 Speciation and CO₂ loading vs. CO₂ partial pressure (1 M PG, 313 K).Figure 4.2.1.4 Speciation and CO₂ loading vs. CO₂ partial pressure (3 M PG, 313 K).

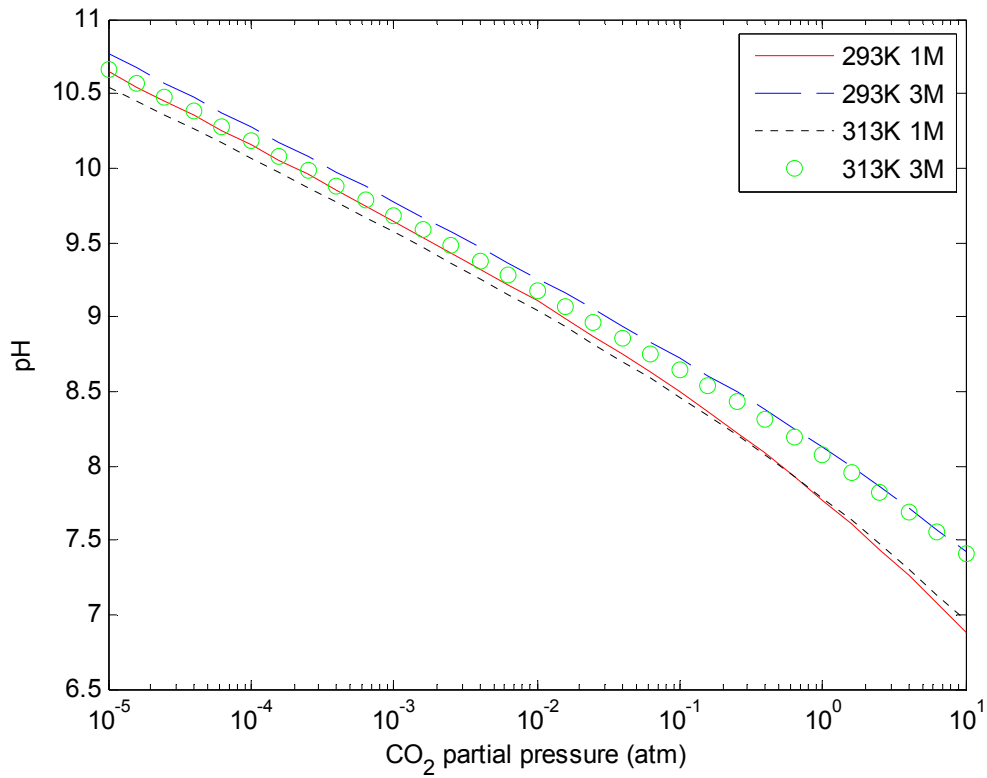


Figure 4.2.1.5 pH vs. CO₂ partial pressure.

Figure 4.2.1.5 depicts the dependence of pH on CO₂ partial pressure. The pH decreases steadily from around 10.6 at 10⁻⁵ atm to under 7.5 at 10 atm, with more sensitivity to PG concentration than to temperature. At CO₂ partial pressures typical of coal-fired power plant flue gases (0.10-0.15 atm), the solution is slightly basic (pH ≈ 8.5).

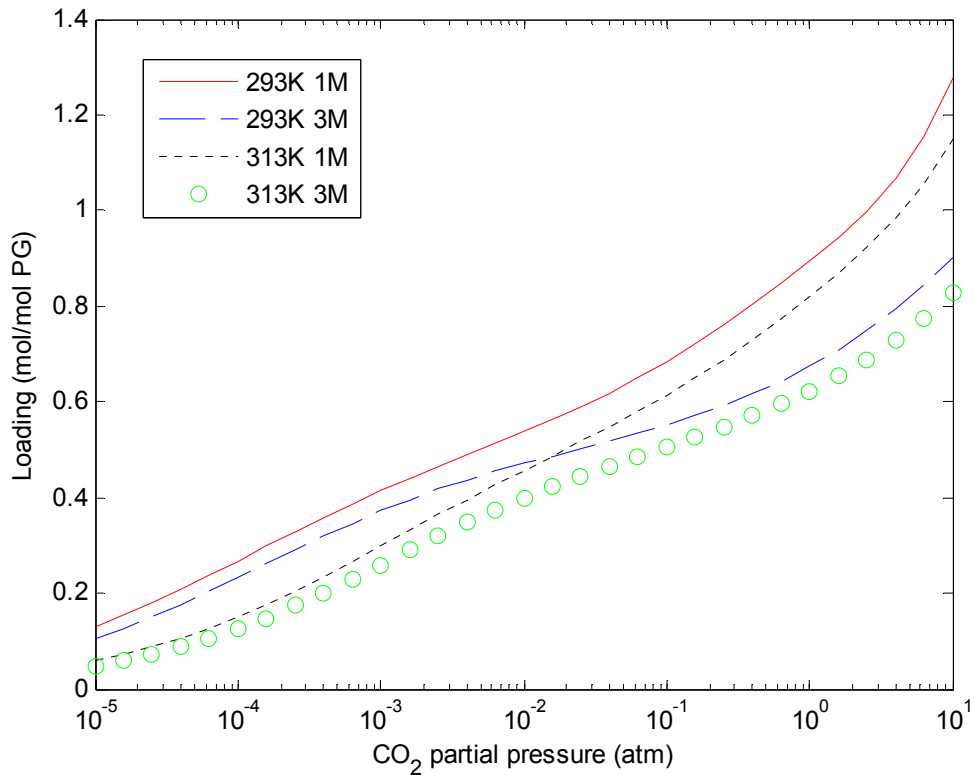


Figure 4.2.1.6 Loading vs. CO₂ partial pressure.

Figure 4.2.1.6 shows the CO₂ partial pressure dependence of the liquid “loading”, L , i.e. total moles of reversibly absorbed CO₂ per mole of amine or amino acid (which is included in figures 4.2.1.1-4). At CO₂ partial pressures typical of post-combustion flue gas, absorber bottoms loadings are unlikely to exceed 50%. Note that loading increases as temperature decreases; it varies slightly with amine concentration at lower CO₂ partial pressures and more markedly at higher partial pressures. Lower PG concentrations promote loading mainly because the CO₂ solubility increases with decreasing ionic strength.

Because physically dissolved gas is generally negligible compared with the total

CO₂ absorbed (see figures 4.2.1.1-4), equation 3.6.3 may be rewritten as:

$$E^{\text{eq}} \cong \beta \left(\frac{[\text{PG}]_{\text{total}}}{\alpha} \right) \frac{\Delta L}{\Delta p_{\text{A,G}}} \quad (4.2.1.3)$$

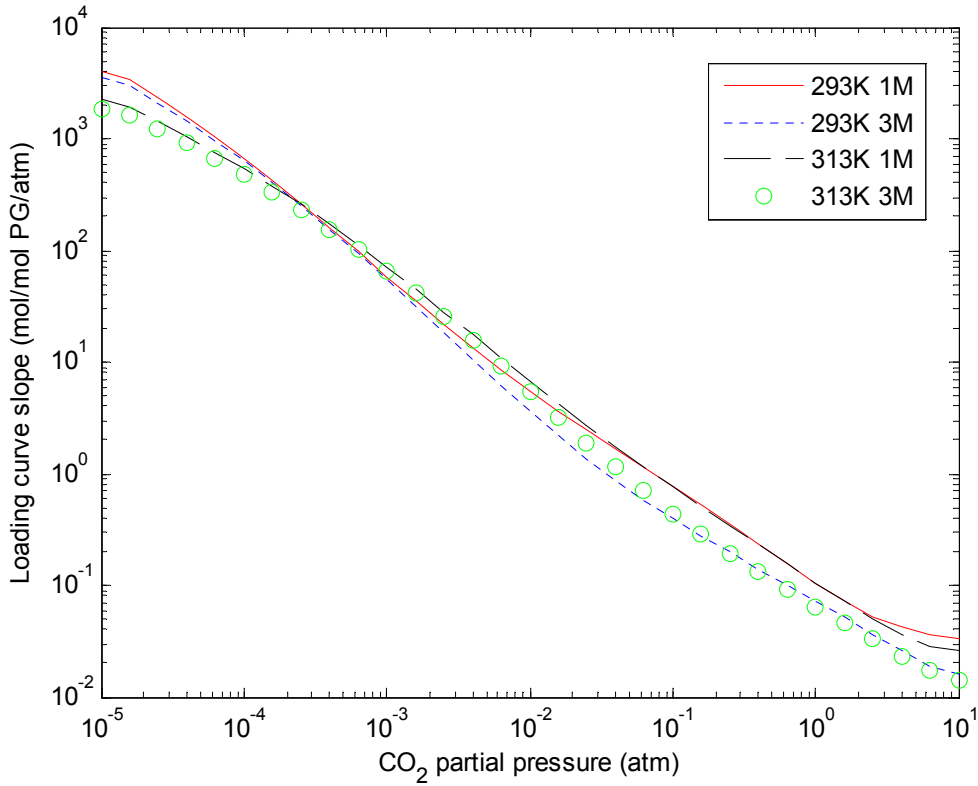


Figure 4.2.1.7 Loading curve slope vs. CO₂ partial pressure.

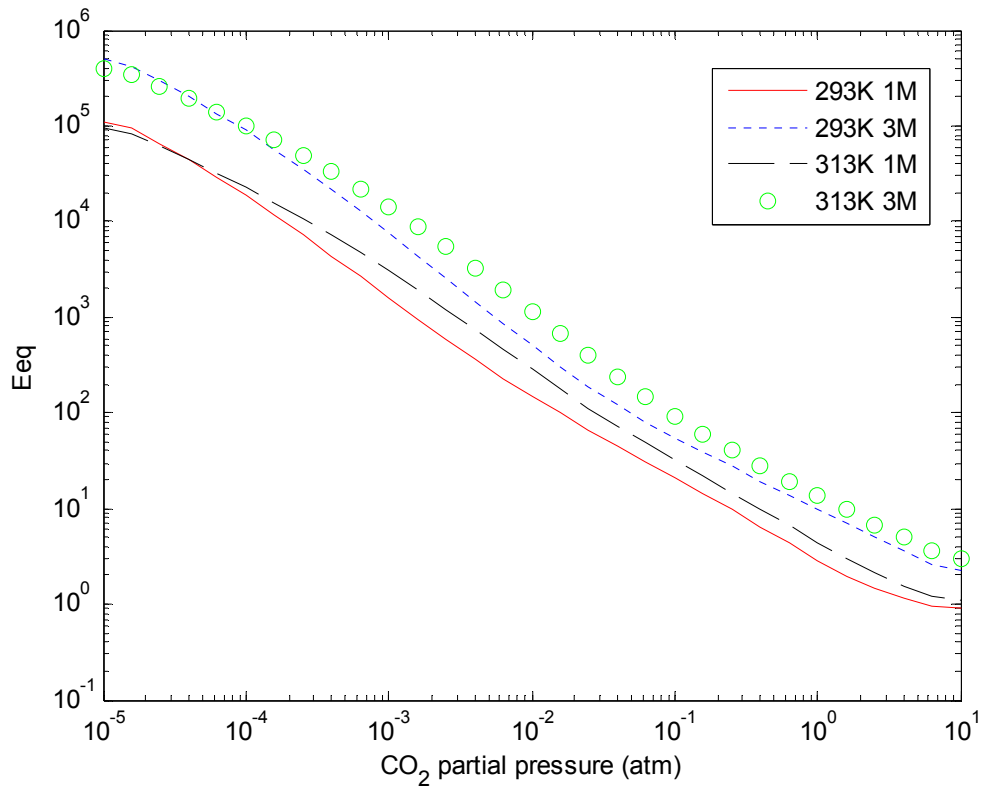


Figure 4.2.1.8 Equilibrium enhancement factor vs. CO₂ partial pressure.

Figure 4.2.1.7 depicts the CO₂ partial pressure dependence of $dL/dp_{A,G}$ (i.e., $\lim_{\Delta p_{A,G} \rightarrow 0} \Delta L / \Delta p_{A,G}$). Equilibrium enhancement factors, E^{eq} , calculated using equation 4.2.1.3, are shown in figure 4.2.1.8. Extremely high E^{eq} values are possible at very low partial pressures, suggesting PG solutions as candidate sorbents for capturing the CO₂ in ambient air. For operating conditions more typical of flue gas capture, E^{eq} values are in the 10 to 10² range. Larger E^{eq} values are attained at higher amino acid concentrations, because of both the increased contributions of carbamate and bicarbonate to CO₂ transport, and the lower CO₂ solubility at higher ionic strengths.

4.2.2 Validation of perturbation analysis

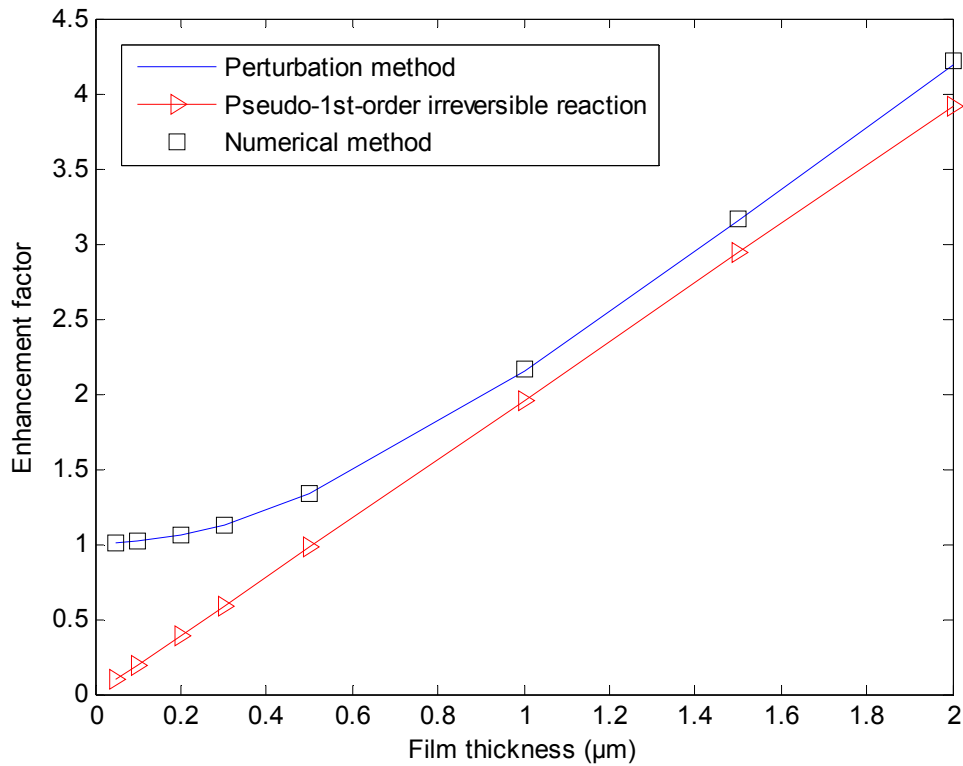


Figure 4.2.2 Absorption enhancement factor vs. liquid film thickness. (3 M PG, 293 K, bulk gas CO₂ partial pressure = 0.15 atm, bulk liquid loading = 40%)

Figure 4.2.2 compares enhancement factors (with liquid film thickness ranging from zero to 2 μm) calculated on the basis of Thick Film theory, with those obtained via: the numerical method; and from the closed-form expression for pseudo-first-order-irreversible reaction, E^+ .

The pseudo-first-order-irreversible assumption retains only the forward reactions and assigns all concentrations other than that of dissolved CO₂ to their bulk liquid values (see equation 4.2.2.1).

$$D_A \frac{d^2[A]}{dx^2} = \left(\frac{k_{1a}[R^-](k_{R^-}[R^-] + k_w[W])}{k_{-1a} + k_{R^-}[R^-] + k_w[W]} + k_2 + \frac{k_3 K_2 [R^-]}{K_3 K_6 [R]} \right) [A] \equiv k_+ [A] \quad (4.2.2.1)$$

$$\text{where } k_+ = \left(\frac{k_{1a}[R^-](k_{R^-}[R^-] + k_w[W])}{k_{-1a} + k_{R^-}[R^-] + k_w[W]} + k_2 + \frac{k_3 K_2 [R^-]}{K_3 K_6 [R]} \right)_{x=\delta}$$

$$\text{Equation 4.2.2.1 can be rearranged as } \frac{d^2[A]}{dx^2} = \frac{k_+}{D_A} [A] \quad (4.2.2.1')$$

A general solution to equation 4.2.2.1' should be: $[A] = ae^{\sqrt{\frac{k_+}{D_A}}x} + be^{-\sqrt{\frac{k_+}{D_A}}x}$

However, as film thickness $x \rightarrow +\infty$, the term, $e^{\sqrt{\frac{k_+}{D_A}}x}$, gives a positive infinite value,

which violates the physical model that $[A]$ must be finite.

$$\text{Therefore, a reasonable solution should be: } [A] = [A]_{x=0} e^{-\sqrt{\frac{k_+}{D_A}}x} \quad (4.2.2.2)$$

where $[A]_{x=0}$ (= b) is the concentration of CO₂ at the gas-liquid interphase.

$$\text{Thus, reaction-enhanced flux} = -D_A \left(\frac{d[A]}{dx} \right)_{x=0} = D_A [A]_{x=0} \sqrt{\frac{k_+}{D_A}} \quad (4.2.2.3)$$

$$\text{while purely mass transfer flux} = D_A \frac{[A]_{x=0}}{\delta} \quad (4.2.2.4)$$

$$\text{Therefore, } E^+ = \frac{\text{reaction-enhanced flux}}{\text{purely mass transfer flux}} = \frac{D_A [A]_{x=0} \sqrt{\frac{k_+}{D_A}}}{D_A \frac{[A]_{x=0}}{\delta}} = \delta \sqrt{\frac{k_+}{D_A}} = \text{Ha} \quad (4.2.2.5)$$

(Ha is the Hatta number).

Figure 4.2.2 shows that thick film theory enhancement factors match those based on numerical analysis and are in close agreement with E^+ values calculated from equation 4.2.2.5.

With liquid film thickness in the range from 0.1 to 0.3 μm , E values are not much

greater than 1 and di-carbamate formation is effectively a “slow” reaction. The pseudo-first-order irreversible reaction assumption is roughly valid from thickness 1 μm to 10 μm , although by 10 μm the combined negative effects of glycinate anion depletion and reaction reversibility are apparent in Figure 4.2.3.1’s 1 M PG curve and Figure 4.2.3.2’s 50% loading curve.

4.2.3 Results of perturbation analysis

4.2.3.1 Effect of concentration

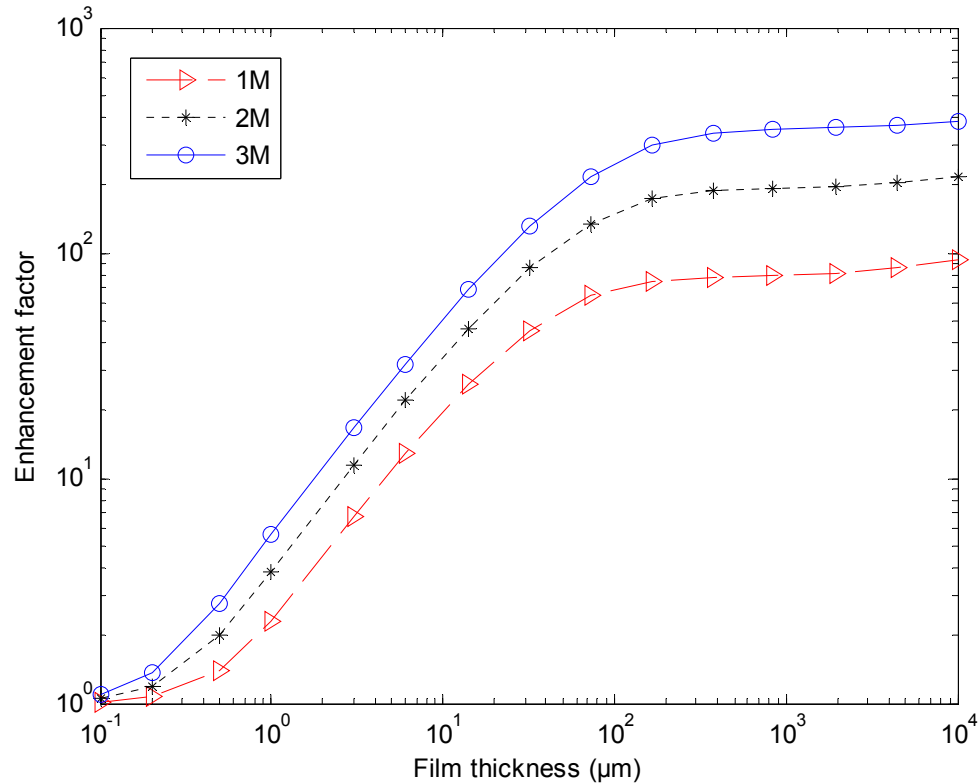


Figure 4.2.3.1 Absorption enhancement factor vs. liquid film thickness. (Bulk liquid loading = 10%, 293 K, bulk gas CO₂ partial pressure = 0.15 atm)

Figure 4.2.3.1 reflects the fact that at higher salt concentration there is less glycinate anion depletion at the interphase. At large film thicknesses, absorption of CO₂ becomes limited by the diffusion of the glycinate anions to the reaction zone, especially at low PG concentrations, and E values plateau, and then approaches a maximum value reflecting the effect of the CO₂ hydration reaction.

4.2.3.2 Effect of bulk liquid loading

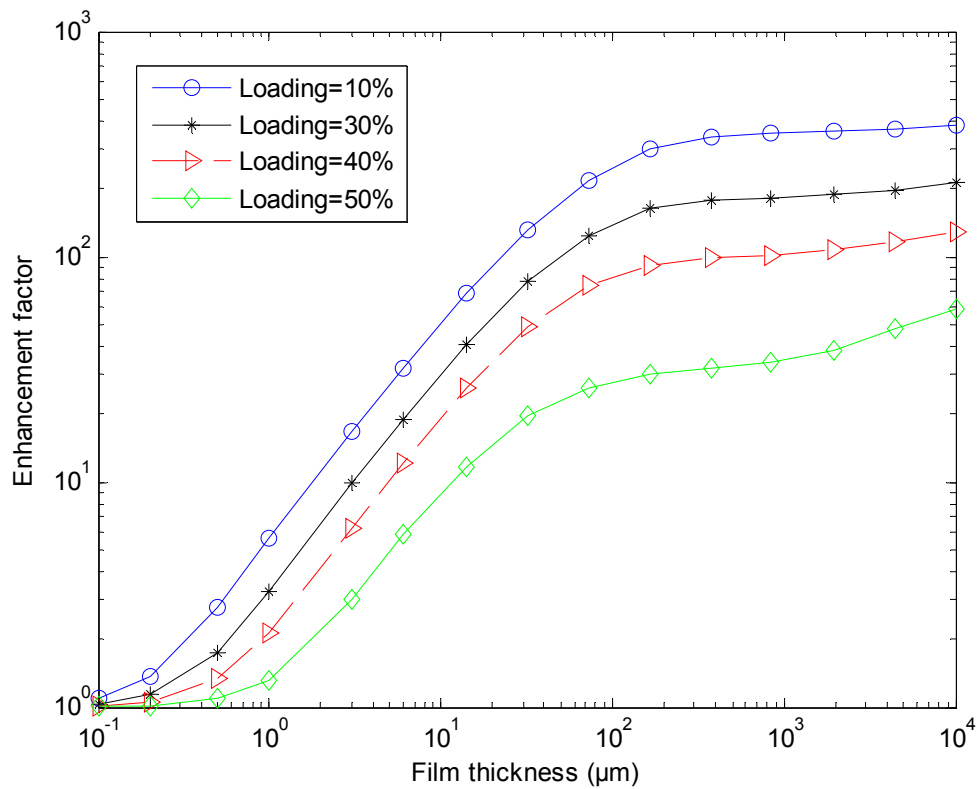


Figure 4.2.3.2 Absorption enhancement factor vs. liquid film thickness. (3 M PG, 293 K, bulk gas CO₂ partial pressure = 0.15 atm)

Figure 4.2.3.2 shows that lower bulk liquid loading promote consumption of absorbed CO₂ and therefore yield higher absorption rates (E values). At still higher film thicknesses, curves appear to plateau, which suggests local equilibrium of reaction 3.1.1. However, the plateaus are only temporary, because as thickness increases from 100 μm to 1000 μm, CO₂ diffusion becomes so slow that even intrinsically slow reactions, such as CO₂ hydration, proceeds significantly within the liquid film. The E values ultimately approach the E^{eq} values in Figure 4.2.1.8 as bicarbonate ions finally facilitate CO₂ transport from the gas-liquid interface to bulk liquid. This phenomenon is more obvious

in Figure 4.2.3.2's 50% loading curve. Although CO₂ reacts much faster with hydroxyl ion, its effects are suppressed by hydroxyl ion depletion at the interface.

4.2.3.3 Concentration profiles within a film

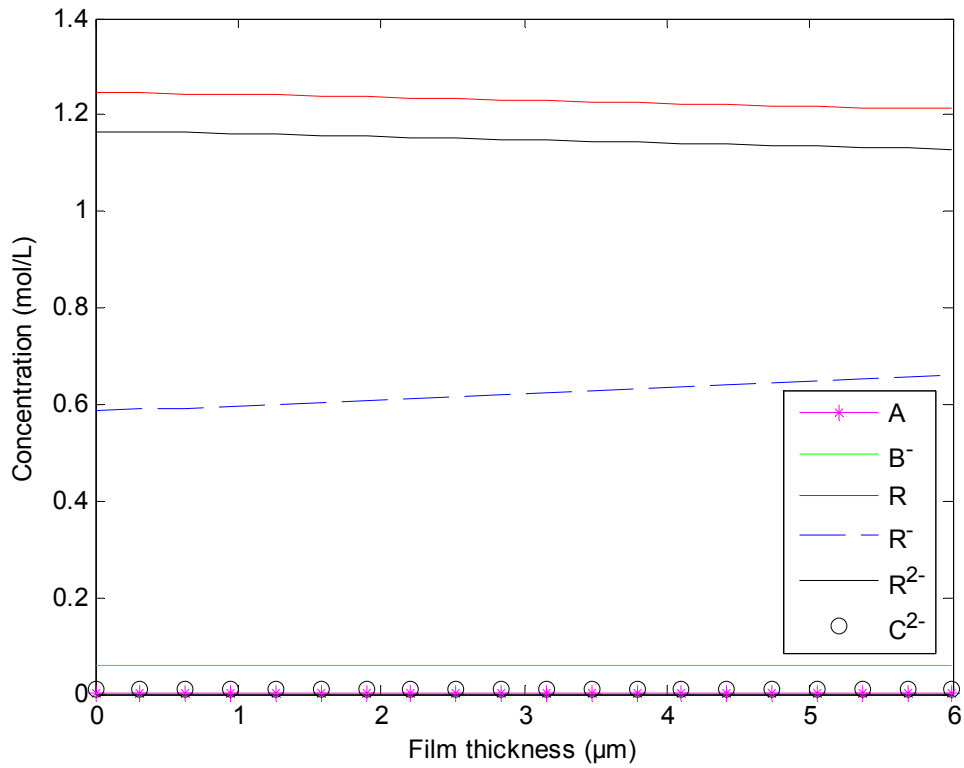


Figure 4.2.3.3.1 Concentration profiles within a 6 μm liquid film. (3 M PG, 293 K, bulk liquid loading = 40%, bulk gas CO₂ partial pressure = 0.15 atm)

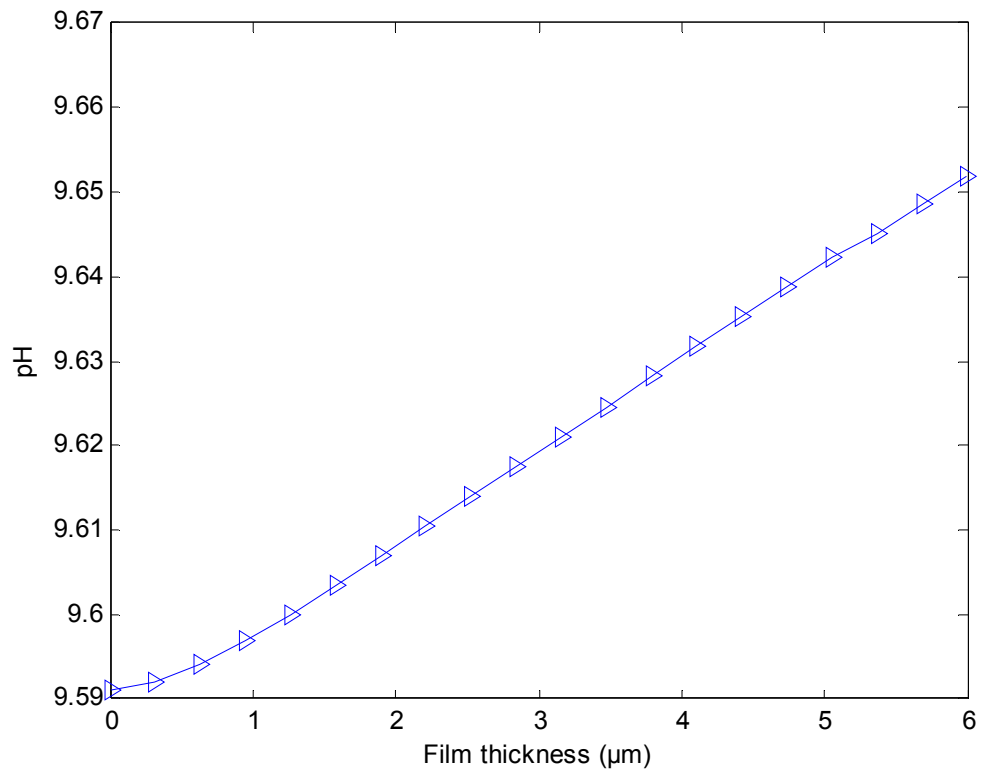


Figure 4.2.3.3.2 pH change within a 6 μm liquid film. (3 M PG, 293 K, bulk liquid loading = 40%, bulk gas CO₂ partial pressure = 0.15 atm)

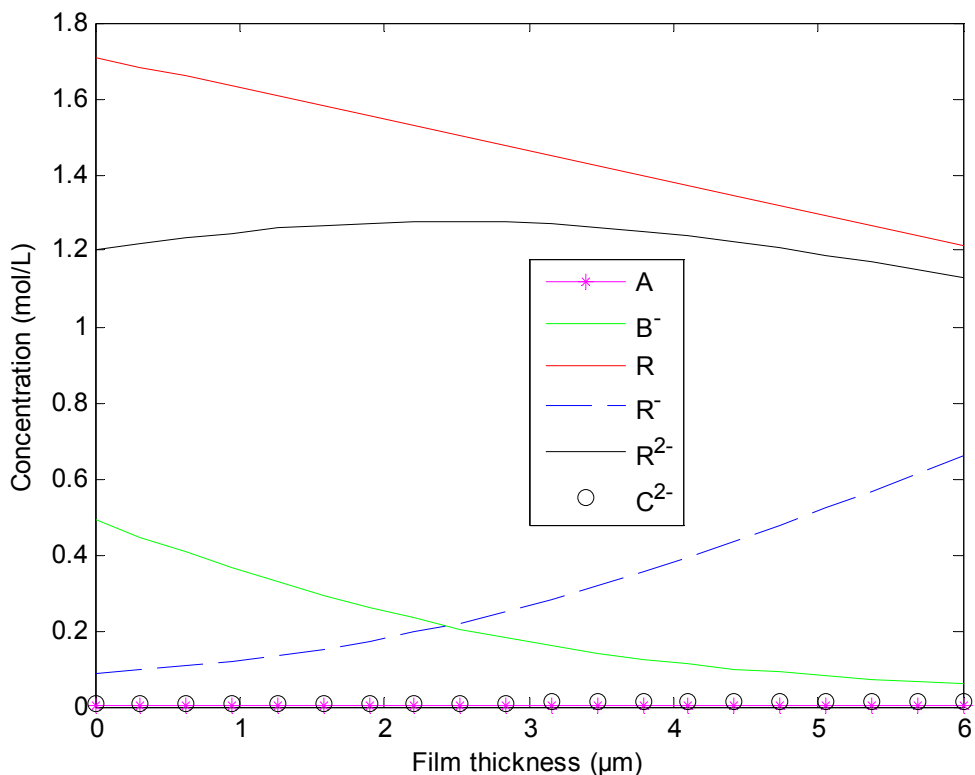


Figure 4.2.3.3.3 Equilibrium concentration profiles within a 6 μm liquid film. (3 M PG, 293 K, bulk liquid loading = 40%, bulk gas CO₂ partial pressure = 0.15 atm)

Figure 4.2.3.3.1 and 4.2.3.3.2 show concentration and pH profiles within a 6 μm liquid film. The film is relatively acidic near the gas-liquid interface; still, the pH is close to 10 throughout the film. As compared with the equilibrium concentration profiles (Figure 4.2.3.3.3), reaction effects are small because chemical reaction is effectively “slow” within a 6 μm liquid film (through which diffusion is rapid).

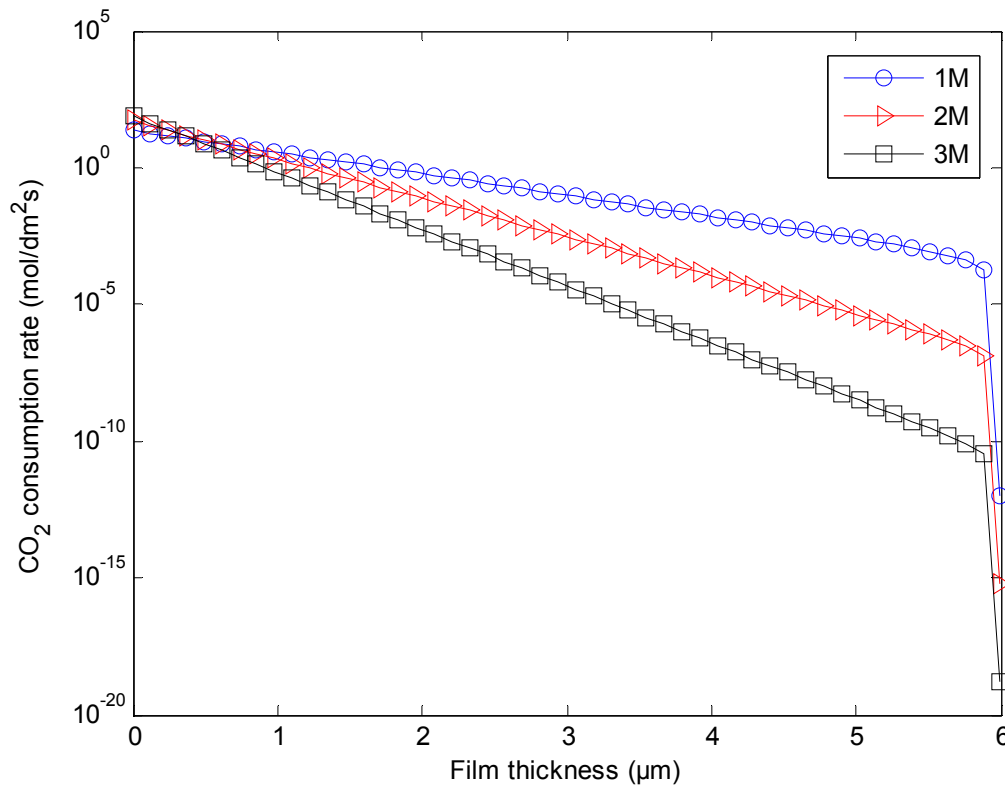
4.2.3.4 CO₂ consumption rate within a film

Figure 4.2.3.4.1 CO₂ consumption rate within a 6 μm liquid film. (Bulk liquid loading = 10%, 293 K, bulk gas CO₂ partial pressure = 0.15 atm)

Figure 4.2.3.4.1 compares CO₂ consumption rates within a 6 μm liquid film at three PG concentrations. The CO₂ reaction rate is fastest at the gas-liquid interface in the most concentrated film, and drops precipitously further into the film, because of the equilibrium condition in bulk liquid.

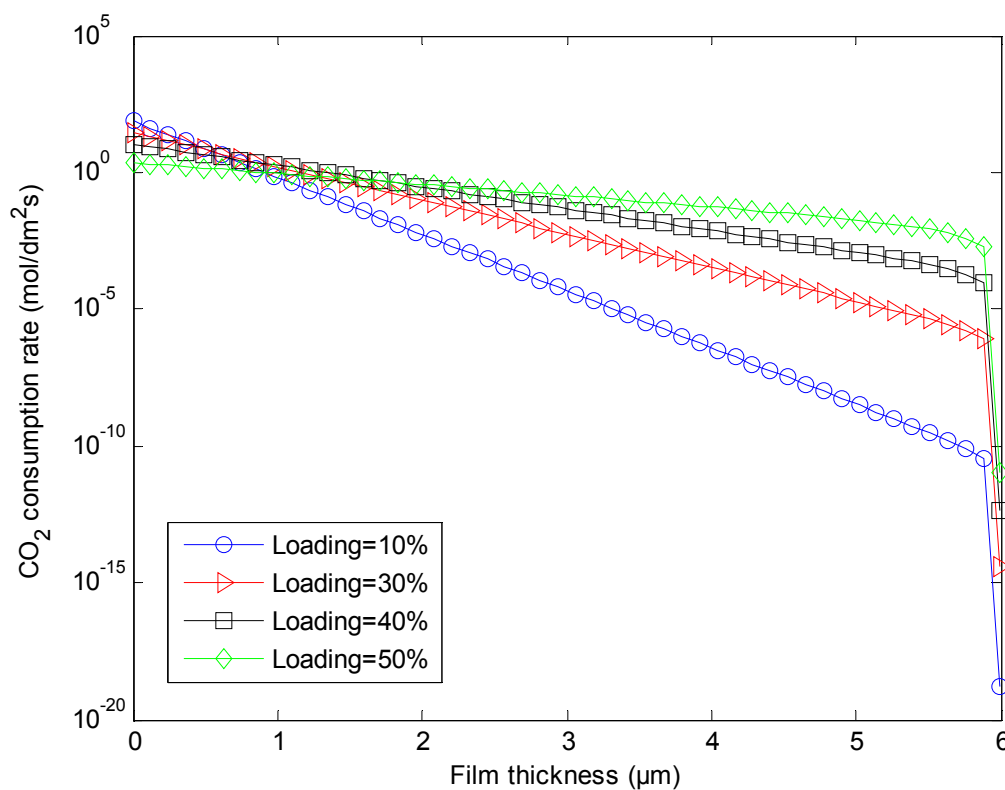


Figure 4.2.3.4.2 CO₂ consumption rate within a 6 µm liquid film. (3 M PG, 293 K, bulk gas CO₂ partial pressure = 0.15 atm)

Figure 4.2.3.4.2 shows CO₂ consumption rates within a 6 µm liquid film at 3 M PG concentration, with different bulk liquid loadings. CO₂ is consumed most rapidly at the lowest bulk liquid loading because the correspondingly suppressed reverse reaction.

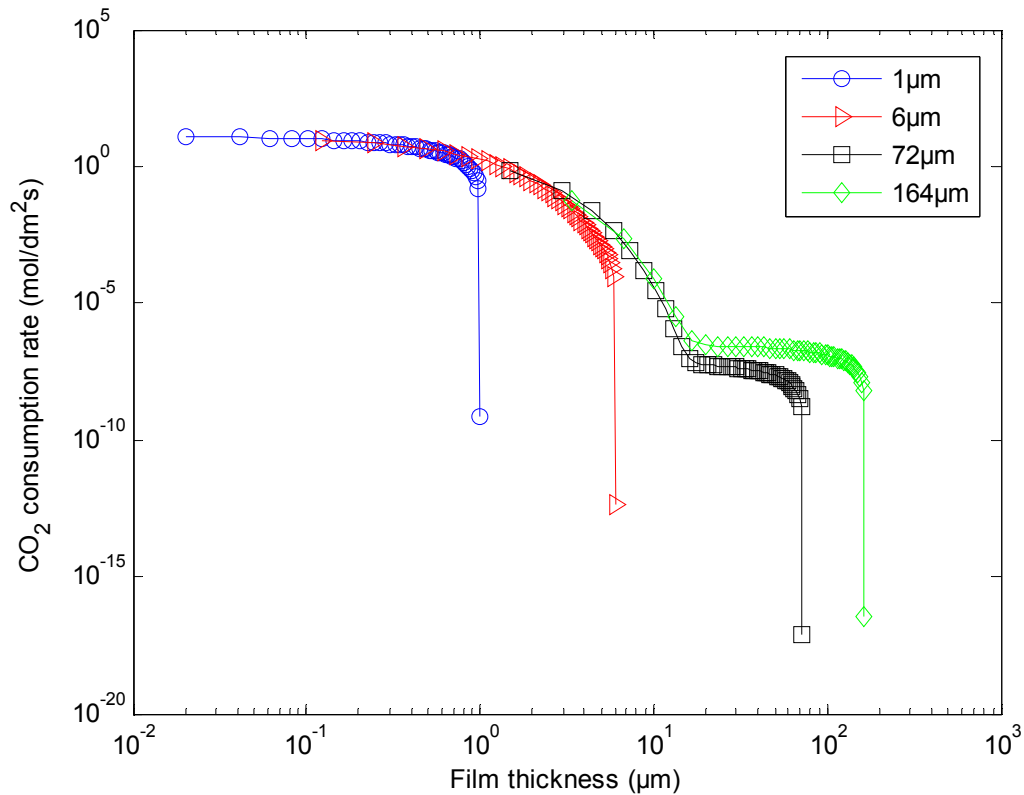


Figure 4.2.3.4.3 CO₂ consumption rate within liquid film. (3 M PG, 293 K, Bulk liquid loading = 40%, bulk gas CO₂ partial pressure = 0.15 atm)

Figure 4.2.3.4.3 shows CO₂ consumption rate in films of different thicknesses. In the two thinner films (1 and 6 μm), diffusion is fast relative to reaction, so CO₂ is consumed throughout the liquid film until its reaction equilibrium prevails at the entrance to the bulk liquid zone. In thicker films (72 and 164 μm), CO₂ consumption is relatively fast compared to diffusion, and local equilibrium prevails before the CO₂ reaches the bulk liquid. Given the reaction kinetics and operating conditions considered here, films thicker than 10 μm may be regarded as “thick”.

4.2.3.5 Fluxes of CO₂-containing species

In this section, it is shown that the fluxes of CO₂ into and out of the liquid film are equal, consistent with the steady-state assumption.

The calculation is in two steps – a first calculation is conducted at $x = 0$ to solve all the constants.

We start with the following equations, which are calculated by fixing the underbarred constants at $x = 0$, as is the case for standard perturbation method:

$$\frac{d\Delta[R^{2-}]}{dx} = \sum_{j=1}^4 \varepsilon_j m_j b_j e^{m_j x} \quad (3.7.22d)$$

$$\frac{d\Delta[A]}{dx} = \sum_{j=1}^4 m_j b_j e^{m_j x} \quad (3.7.23d)$$

$$\Delta[B^-] = \omega_1 \Delta[R^{2-}] + \omega_2 \Delta[A] \quad (3.7.16)$$

$$\Delta[R] = \omega_3 \Delta[R^{2-}] + \omega_4 \Delta[A] \quad (3.7.17)$$

$$\Delta[R^-] = \omega_5 \Delta[R^{2-}] + \omega_6 \Delta[A] \quad (3.7.18)$$

(ω 's are fixed at $x = 0$)

$$\Delta[C^{2-}] = \left(\frac{K_4[B^-]}{K_6[R]} \right)_{x=0} \Delta[R^-] + \left(\frac{K_4[R^-]}{K_6[R]} \right)_{x=0} \Delta[B^-] - \left(\frac{K_4[R^-][B^-]}{K_6[R]^2} \right)_{x=0} \Delta[R] \quad (3.7.15)$$

Once the other constants are solved (i.e., b 's, E 's, m 's etc.), we then continue on to

obtain $\frac{d\Delta[R^{2-}]}{dx}$ and $\frac{d\Delta[A]}{dx}$ values at $x = \delta$:

$$\left(\frac{d\Delta[R^{2-}]}{dx} \right)_{x=\delta} = \sum_{j=1}^4 \varepsilon_j m_j b_j e^{m_j \delta} \quad (4.2.3.5.1)$$

$$\left(\frac{d\Delta[A]}{dx} \right)_{x=\delta} = \sum_{j=1}^4 m_j b_j e^{m_j \delta} \quad (4.2.3.5.2)$$

And the other delta derivatives are obtained at $x = \delta$:

$$\left(\frac{d\Delta[B^-]}{dx}\right)_{x=\delta} = \omega_1 \left(\frac{d\Delta[R^{2-}]}{dx}\right)_{x=\delta} + \omega_2 \left(\frac{d\Delta[A]}{dx}\right)_{x=\delta} \quad (4.2.3.5.3)$$

$$\left(\frac{d\Delta[R]}{dx}\right)_{x=\delta} = \omega_3 \left(\frac{d\Delta[R^{2-}]}{dx}\right)_{x=\delta} + \omega_4 \left(\frac{d\Delta[A]}{dx}\right)_{x=\delta} \quad (4.2.3.5.4)$$

$$\left(\frac{d\Delta[R^-]}{dx}\right)_{x=\delta} = \omega_5 \left(\frac{d\Delta[R^{2-}]}{dx}\right)_{x=\delta} + \omega_6 \left(\frac{d\Delta[A]}{dx}\right)_{x=\delta} \quad (4.2.3.5.5)$$

(ω 's are again constants at $x = 0$)

$$\begin{aligned} \left(\frac{d\Delta[C^{2-}]}{dx}\right)_{x=\delta} &= \left(\frac{K_4[B^-]}{K_6[R]}\right)_{x=0} \left(\frac{d\Delta[R^-]}{dx}\right)_{x=\delta} + \left(\frac{K_4[R^-]}{K_6[R]}\right)_{x=0} \left(\frac{d\Delta[B^-]}{dx}\right)_{x=\delta} \\ &\quad - \left(\frac{K_4[R^-][B^-]}{K_6[R]^2}\right)_{x=0} \left(\frac{d\Delta[R]}{dx}\right)_{x=\delta} \end{aligned} \quad (4.2.3.5.6)$$

Note that we fix the θ 's at $x = 0$ for solving the constants; but for calculating the underbarred derivatives at $x = \delta$, we re-calculate the θ 's at $x = \delta$ (while Φ_1 remains to be its value at $x = 0$) following equations 3.7.47 and 3.7.48:

$$\left(\frac{d[A]}{dx}\right)_{x=\delta} = - \left(\frac{\theta_1}{\theta_1 D_A + D^*(\theta_2 + \theta_4 + \theta_5)}\right)_{x=\delta} (\Phi_1)_{x=0} \quad (4.2.3.5.7)$$

$$\left(\frac{d[R^{2-}]}{dx}\right)_{x=\delta} = - \left(\frac{\theta_4}{\theta_1 D_A + D^*(\theta_2 + \theta_4 + \theta_5)}\right)_{x=\delta} (\Phi_1)_{x=0} \quad (4.2.3.5.8)$$

$$\left(\frac{d[B^-]}{dx}\right)_{x=\delta} = - \left(\frac{\theta_2}{\theta_1 D_A + D^*(\theta_2 + \theta_4 + \theta_5)}\right)_{x=\delta} (\Phi_1)_{x=0} \quad (4.2.3.5.9)$$

$$\left(\frac{d[C^{2-}]}{dx}\right)_{x=\delta} = - \left(\frac{\theta_5}{\theta_1 D_A + D^*(\theta_2 + \theta_4 + \theta_5)}\right)_{x=\delta} (\Phi_1)_{x=0} \quad (4.2.3.5.10)$$

Finally, the fluxes of CO₂-containing species (i.e. physically dissolved CO₂,

di-carbamate, carbonate, and bicarbonate) are calculated as follows:

$$-D_A \left(\frac{d[A]}{dx} \right)_{x=\delta} = -D_A \left[\left(\frac{d[A]}{dx} \right)_{x=\delta} + \left(\frac{d\Delta[A]}{dx} \right)_{x=\delta} \right] \quad (4.2.3.5.11)$$

$$-D^* \left(\frac{d[R^{2-}]}{dx} \right)_{x=\delta} = -D^* \left[\left(\frac{d[R^{2-}]}{dx} \right)_{x=\delta} + \left(\frac{d\Delta[R^{2-}]}{dx} \right)_{x=\delta} \right] \quad (4.2.3.5.12)$$

$$-D^* \left(\frac{d[B^-]}{dx} \right)_{x=\delta} = -D^* \left[\left(\frac{d[B^-]}{dx} \right)_{x=\delta} + \left(\frac{d\Delta[B^-]}{dx} \right)_{x=\delta} \right] \quad (4.2.3.5.13)$$

$$-D^* \left(\frac{d[C^{2-}]}{dx} \right)_{x=\delta} = -D^* \left[\left(\frac{d[C^{2-}]}{dx} \right)_{x=\delta} + \left(\frac{d\Delta[C^{2-}]}{dx} \right)_{x=\delta} \right] \quad (4.2.3.5.14)$$

Table 4.2.3.5 shows that the sum of the fluxes of CO₂-containing species (i.e. physically dissolved CO₂, di-carbamate, carbonate, and bicarbonate) into bulk liquid (i.e. at $x = \delta$) equals the CO₂ absorption flux at the gas-liquid interface (i.e. $x = 0$). This is consistent with the steady-state assumption, which requires that there be no accumulation or depletion of CO₂ within the film.

Under actual operating conditions, the slow CO₂ hydration reaction is likely to be unequilibrated in bulk liquid, to an extent that varies within a given contactor – whereas that reaction, like all others, is simply assumed in the foregoing analysis to be at equilibrium. Whether carbonate and bicarbonate will diffuse to (attain a positive value) or from (attain a negative value) the bulk liquid interface will depend on operating conditions.

In general, the dicarbamate fluxes dominates CO₂ transport away from the

interface, although their magnitude approaches zero as the film thickness approaches zero or bulk liquid approaches equilibrium with the feed gas.

Table 4.2.3.5 Summary of fluxes of CO₂-containing species at boundaries

Conditions: T = 293 K, [PG] = 3 M, CO ₂ partial pressure = 0.15 atm			
Flux [=] mol dm ⁻² s ⁻¹	CO ₂ bulk loading = 10%		
	1 μm	6 μm	164 μm
Total fluxes of CO ₂ -containing species at x = 0	1.7293×10 ⁻⁴	1.6298×10 ⁻⁴	5.5402×10 ⁻⁵
Total fluxes of CO ₂ -containing species at x = δ	1.7293×10 ⁻⁴	1.6298×10 ⁻⁴	5.5402×10 ⁻⁵
Flux percentage of A at x = δ	1.2659 %	1.7288×10 ⁻⁴ %	0.0088 %
Flux percentage of R ²⁻ at x = δ	98.7764 %	100.2533 %	127.6770 %
Flux percentage of B ⁻ at x = δ	1.1625 %	0.9660 %	-23.7942 %
Flux percentage of C ²⁻ at x = δ	-1.2047 %	-1.2195 %	-3.8916 %
	CO ₂ bulk loading = 30%		
	1 μm	6 μm	164 μm
Total fluxes of CO ₂ -containing species at x = 0	1.0305×10 ⁻⁴	9.6250×10 ⁻⁵	2.9918×10 ⁻⁵
Total fluxes of CO ₂ -containing species at x = δ	1.0305×10 ⁻⁴	9.6250×10 ⁻⁵	2.9918×10 ⁻⁵
Flux percentage of A at x = δ	9.8560 %	0.0039 %	0.1313 %
Flux percentage of R ²⁻ at x = δ	90.3781 %	101.4173 %	161.8368 %
Flux percentage of B ⁻ at x = δ	1.7038 %	0.8744 %	-55.3794 %
Flux percentage of C ²⁻ at x = δ	-1.9379 %	-2.2956 %	-6.5887 %

Table 4.2.3.5 Summary of fluxes of CO₂-containing species at boundaries (continued)

	CO ₂ bulk loading = 40%		
	1 μm	6 μm	164 μm
Total fluxes of CO ₂ -containing species at x = 0	6.8064×10 ⁻⁵	6.1141×10 ⁻⁵	1.6663×10 ⁻⁵
Total fluxes of CO ₂ -containing species at x = δ	6.8064×10 ⁻⁵	6.1141×10 ⁻⁵	1.6663×10 ⁻⁵
Flux percentage of A at x = δ	28.3461 %	0.0381 %	0.4833 %
Flux percentage of R ²⁻ at x = δ	72.3880 %	104.3154 %	174.3202 %
Flux percentage of B ⁻ at x = δ	1.8752 %	-0.4386 %	-66.3413 %
Flux percentage of C ²⁻ at x = δ	-2.6093 %	-3.9149 %	-8.4622 %
	CO ₂ bulk loading = 50%		
	1 μm	6 μm	164 μm
Total fluxes of CO ₂ -containing species at x = 0	3.6999×10 ⁻⁵	2.5190×10 ⁻⁵	4.6833×10 ⁻⁶
Total fluxes of CO ₂ -containing species at x = δ	3.6999×10 ⁻⁵	2.5190×10 ⁻⁵	4.6833×10 ⁻⁶
Flux percentage of A at x = δ	67.3343 %	2.0324 %	2.7094 %
Flux percentage of R ²⁻ at x = δ	34.3089 %	104.7317 %	128.6508 %
Flux percentage of B ⁻ at x = δ	1.4385 %	2.8951 %	-19.4862 %
Flux percentage of C ²⁻ at x = δ	-3.0817 %	-9.6592 %	-11.8740 %

5. RESULTS FOR CO₂-PS SYSTEM

5.1 Physiochemical parameters

The solubility, density, diffusivity, equilibrium and kinetic parameters as functions of temperature and concentration are presented below.

5.1.1 Solubility data

CO₂ vapor-liquid equilibrium is described by the Henry law:

$$p_{A,G} \equiv \frac{1}{\alpha} [A] = H_{CO_2} [A] \quad (5.1.1)$$

where $p_{A,G}$ is the CO₂ partial pressure and H_{CO_2} is the Henry coefficient of CO₂ in the PS solution given by U. E. Aronu [53] and is derived by interpolation for the following temperatures:

Table 5.1.1 Henry coefficients of PS [48]

[PS] (M)	H/10 ⁶ (Pa dm ³ mol ⁻¹)	
	313.15 (K)	333.15 (K)
1	6.20	8.37
2	8.20	10.69
3	10.02	12.81

5.1.2 Density data

Densities of PS solutions were studied by Jacco van Holst et al. [83] as:

$$\rho = -0.4862T + 50.00[PS] + 1146 \quad (\rho [=] \text{ g/L}, C [=] \text{ mol/L}, T [=] \text{ K}) \quad (5.1.2)$$

5.1.3 Diffusivity data

The diffusivity of CO₂ in the aqueous sarcosine salt solution is estimated using the relation as follows [47, 48]:

$$D_{\text{CO}_2,\text{PS}} = D_{\text{CO}_2,\text{W}} (\mu_{\text{W}} / \mu_{\text{PS}})^{0.8} \quad (5.1.3.1)$$

where $D_{\text{CO}_2,\text{W}}$ is the diffusivity of CO₂ in water, given by J. Haubrock [84] as:

$$\log D_{\text{CO}_2,\text{W}} = -8.176 + 712.5/T - 2.591 \times 10^5 / T^2 \quad (D_{\text{CO}_2,\text{W}} [=] \text{m}^2 / \text{s}) \quad (5.1.3.2)$$

μ_{W} is the viscosity of water, given by Kampmeyer [85] as:

$$\log \mu_{\text{W}} = -5.485471 + 5.041873 \times 10^3 / T - 1.551439 \times 10^6 / T^2 + 0.1849577 \times 10^9 / T^3$$

$$(\mu_{\text{W}} [=] \text{poise} \times 10^{-3} [=] 0.1 \text{ mPa} \cdot \text{s}) \quad (5.1.3.3)$$

μ_{PS} is the viscosity of PS solution, given by Jacco van Holst [83] as:

$$\mu_{\text{PS}} = 2.950 \times 10^{-3} \exp \left[\frac{14150 \exp(0.1047[\text{PS}])}{R_{\text{g}} T} \right] \exp(-0.3124[\text{PS}]) \quad (\text{mPa} \cdot \text{s}) \quad (5.1.3.4)$$

$$(R_{\text{g}} = 8.314 \text{ J} / \text{mol} \cdot \text{K})$$

Diffusion coefficients of PS is tabulated in Table 5.1.3 [79].

Table 5.1.3 Diffusion coefficients of PS [79]

T (K)	$D_{\text{PS}} / 10^{-7} \text{ (dm}^2 \text{ s}^{-1}\text{)}$			
[PS] (M)	293.15	298.15	313.15	333.15
0.971 ± 0.001	0.945	1.04	1.50	2.27
1.99	0.893	0.993	1.45	2.18
2.91	0.865	0.982	1.39	2.08

5.1.4 Equilibrium data

Equilibrium constants - K_2 , K_4 , K_5 - in the aqueous CO₂-PS system are the same as those in CO₂-PG system.

For carbamate zwitterion deprotonation, equation 3.1.6, K_6 is given by U. E. Aronu [53] as:

$$K_6 = \frac{[R^-][H^+]}{[R]} = \exp(-5.9752 - 5185.10/T) \quad (\text{mol dm}^{-3}) \quad (5.1.4.1)$$

The di-carbamate hydrolysis reaction is regarded as a dependent reaction in the analysis. It is a linear combination of di-carbamate formation and carbonate formation reactions and its equilibrium constant is given by Aronu [53] as follows (valid in temperature range 40-120 °C):

$$K_{\text{carbohydrolysis}} = \left(\frac{[R^-][B^-]}{[R^{2-}]} \right)_{\text{eq}} = \exp(-7.4569 + 0.9753/T) \quad (\text{mol dm}^{-3}) \quad (5.1.4.2)$$

From the definition of $K_{\text{Acid},j} \equiv \left(\frac{[B_j H^+]}{[B_j][H^+]} \right)^{\text{eq}}$, it follows that:

$$K_{\text{Acid},R^-} = \frac{1}{K_6} \quad (5.1.4.3)$$

and that:

$$K_{\text{Acid},W} = \frac{1}{[W]} \quad (5.1.4.4)$$

From the definition of $K_{\text{Carb}} \equiv \left(\frac{[R^{2-}][H^+]}{[A][R^-]} \right)^{\text{eq}}$, it is easily shown that:

$$K_{\text{Carb}} = \frac{K_2}{K_{\text{carbohydrolysis}}} \quad (5.1.4.5)$$

5.1.5 Kinetic data

The zwitterion mechanism constants are given by Aronu [48], and the nonidealities of the salt solution are lumped into a salt effect expressed by:

$$k_{\text{ion}} = \exp(bI) \quad (5.1.5.1)$$

where the ionic strength is defined by:

$$I = \frac{1}{2} \sum c_i z_i^2$$

and b is a constant that depends on the nature of the salt. In this thesis, an average value of $b = 0.45$ is adopted from a rigorous model used to interpret the effect of the ionic strength that was developed by Cullinane and Rochelle [86]. From equation 5.1.5.1, it is clear that for the limiting case where ionic strength is zero, k_{ion} is one, which then reduces to the nonionic case, as with an alkanolamine. As a first approximation, the ionic strength used in the calculations is that in bulk liquid, i.e.:

$$I = \frac{1}{2} \left([K^+]_{x=\delta} + [B^-]_{x=\delta} + [R^-]_{x=\delta} + 4[R^{2-}]_{x=\delta} + 4[C^{2-}]_{x=\delta} \right) \quad (5.1.5.2)$$

Note that the ionic strength contribution by the zwitterion, R , is considered to be zero [87].

In addition:

$$k_{1a} = 2.6198 \times 10^9 \exp(-915.8/T) \quad (\text{dm}^3 \text{ mol}^{-1} \text{ s}^{-1}) \quad (5.1.5.3)$$

$$k_w / k_{-1a} = \frac{3.9805 \times 10^8 \exp(-3924.4/T)}{k_{1a}} \quad (\text{dm}^3 \text{ mol}^{-1}) \quad (5.1.5.4)$$

$$k_{R^-} / k_{-1a} = \frac{6.3494 \times 10^6 \exp(-1589.6/T)}{k_{1a}} \quad (\text{dm}^3 \text{ mol}^{-1}) \quad (5.1.5.5)$$

The large value of k_{1a} indicates that deprotonation of the zwitterion is the

rate-determining step. The deprotonation rate constants for water and sarcosinate anion are such that water has a more significant effect in deprotonation of the zwitterion of AAS compared to alkanolamines.

The forward reaction rate of carbonic acid formation (k_2), and the forward reaction rate of bicarbonate formation (k_3) are assumed to be the same as those in the CO₂-PG system, which themselves do not account for ionic strength effects consideration [20].

Incorporation of ionic strength effects modifies the key differential mass balance (3.7.1) to become:

$$D^* \frac{d^2[R^{2-}]}{dx^2} = - \frac{k_{\text{ion}} k_{1a} [A][R^-] (k_{R^-} [R^-] + k_w [W]) \left(1 - \frac{K_6 [R^{2-}][R]}{K_{\text{Carb}} [A][R^-]^2} \right)}{k_{-1a} + k_{R^-} [R^-] + k_w [W]} \quad (3.7.1')$$

In the same fashion, terms in all other differential mass balances in the linearized analysis, i.e. η_1 , η_2 , λ_1 , and λ_2 , are multiplied by k_{ion} .

5.2 Results and discussion

5.2.1 Speciation profiles

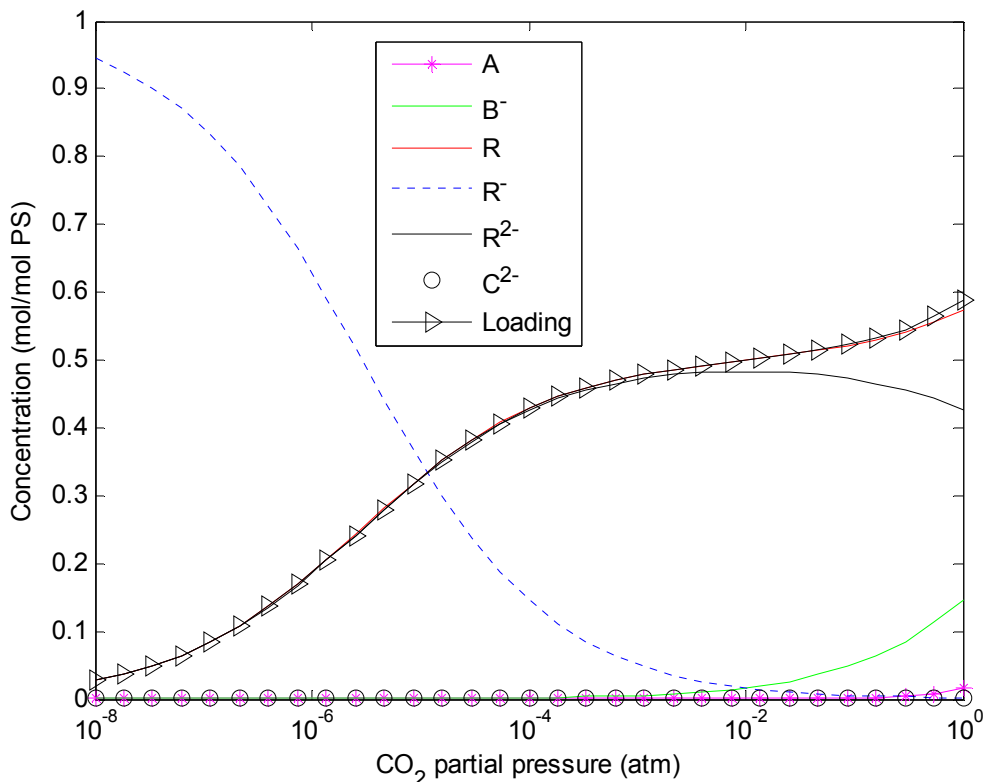
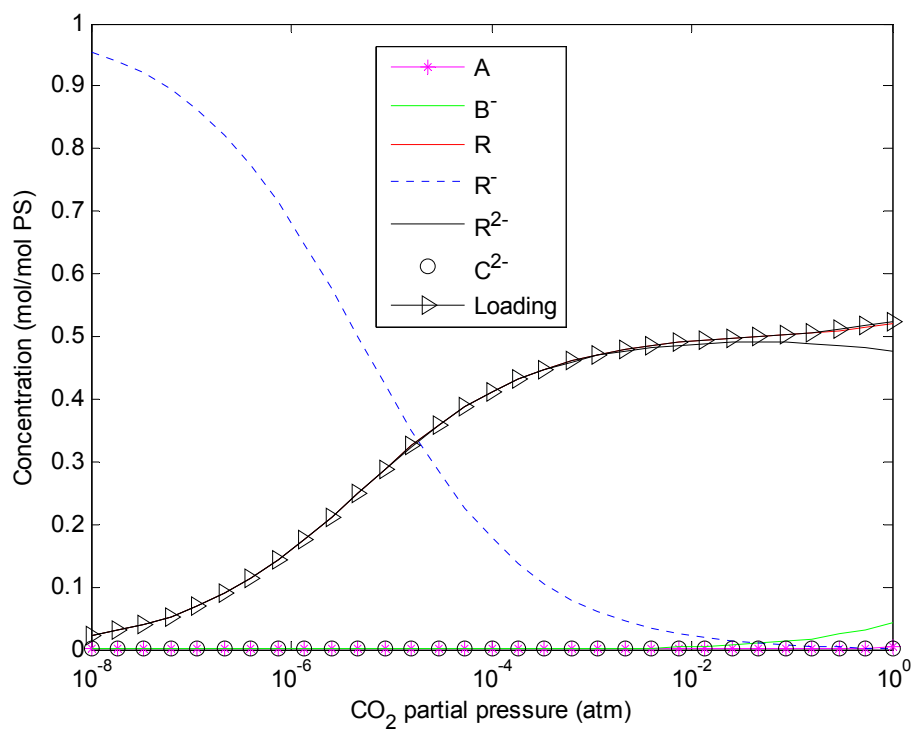
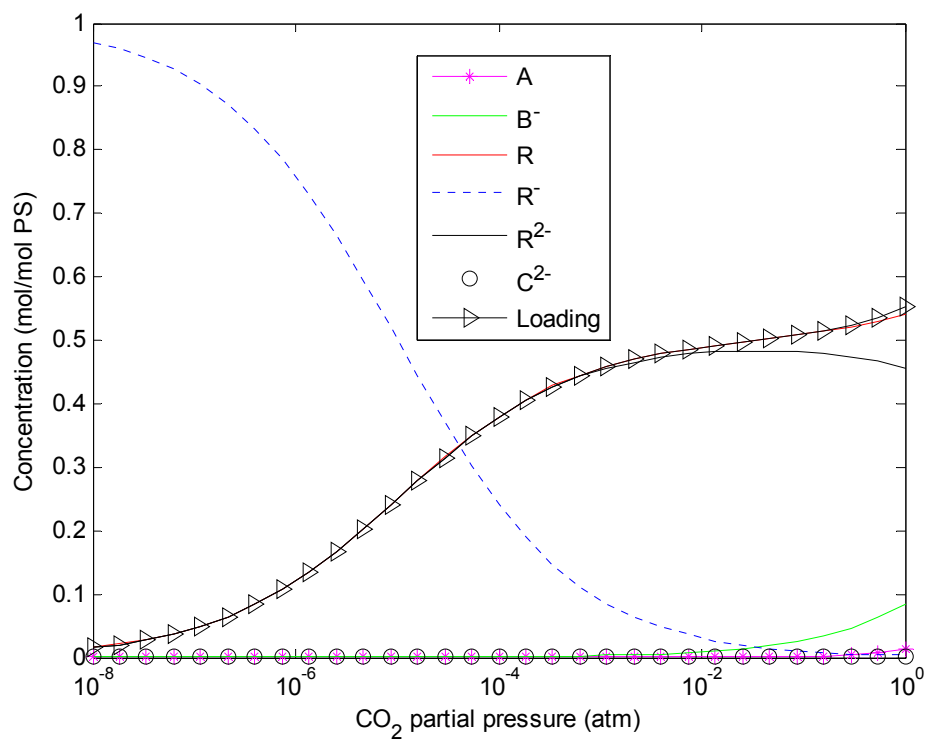


Figure 5.2.1.1 Speciation and CO₂ loading vs. CO₂ partial pressure (1 M PS, 313 K).

Figure 5.2.1.1-4 depict the speciation profiles for 1 M and 3 M PS solutions at 313 K and 333 K, respectively. It is striking that the di-carbamate forms in the presence of even trace amounts of CO₂, i.e. at CO₂ partial pressures as low as 10⁻⁸ atm. At partial pressure up to 10⁻² atm, dissolved CO₂ combines primarily with R⁻ to form equal proportions of R and R²⁻. In PS solutions, bicarbonate formation is vanishingly low under the operating conditions typical of post-combustion CO₂ capture at power plants.

Figure 5.2.1.2 Speciation and CO₂ loading vs. CO₂ partial pressure (3 M PS, 313 K).Figure 5.2.1.3 Speciation and CO₂ loading vs. CO₂ partial pressure (1 M PS, 333 K).

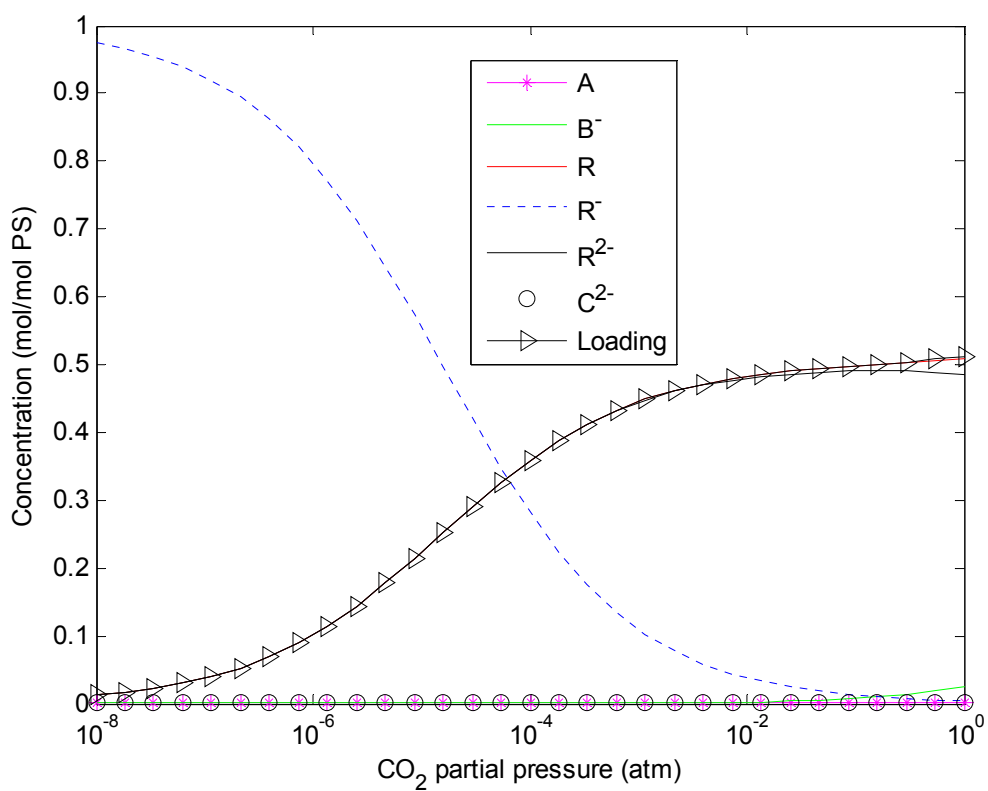


Figure 5.2.1.4 Speciation and CO₂ loading vs. CO₂ partial pressure (3 M PS, 333 K).

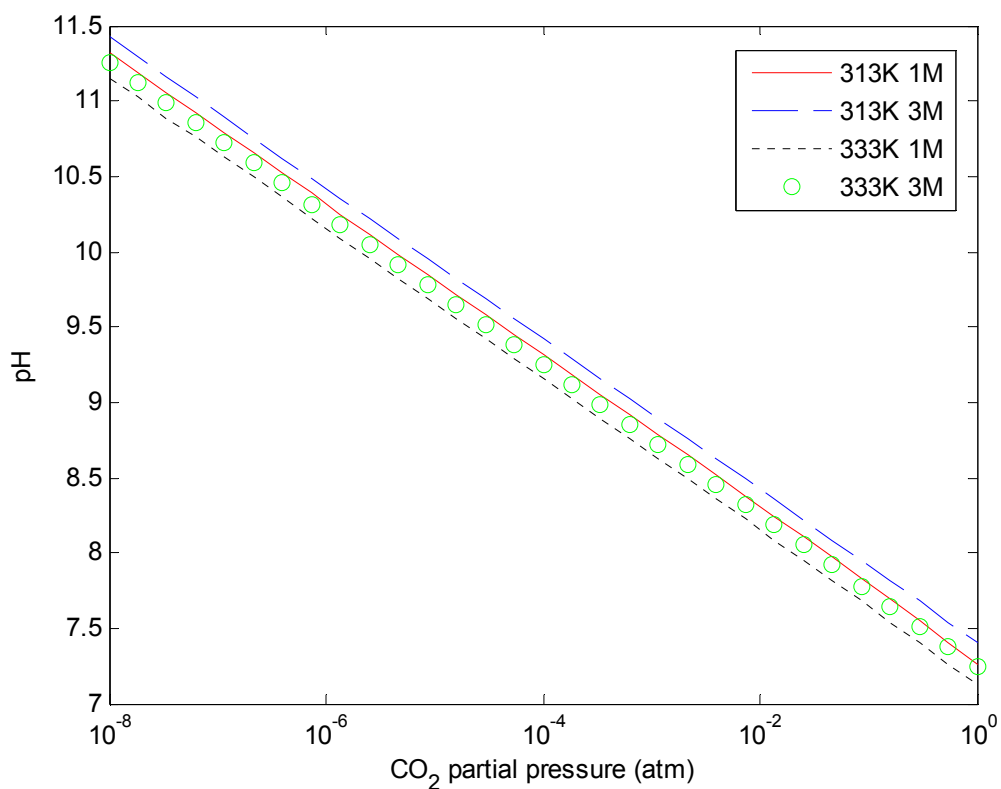


Figure 5.2.1.5 pH vs. CO₂ partial pressure.

Figure 5.2.1.5 shows that the pH in the bulk liquid at overall equilibrium decreases from *ca.* 11.5 at 10⁻⁸ atm CO₂ partial pressure to around 7.25 at 1 atm; the same trend is followed at all four assumed temperatures.

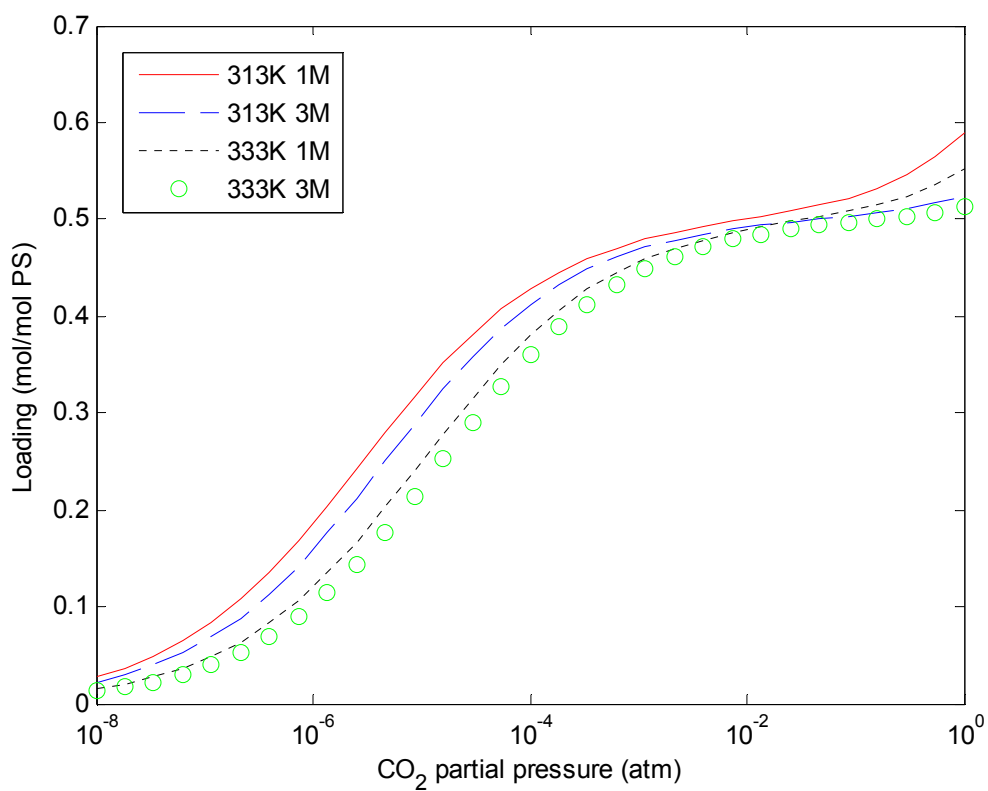
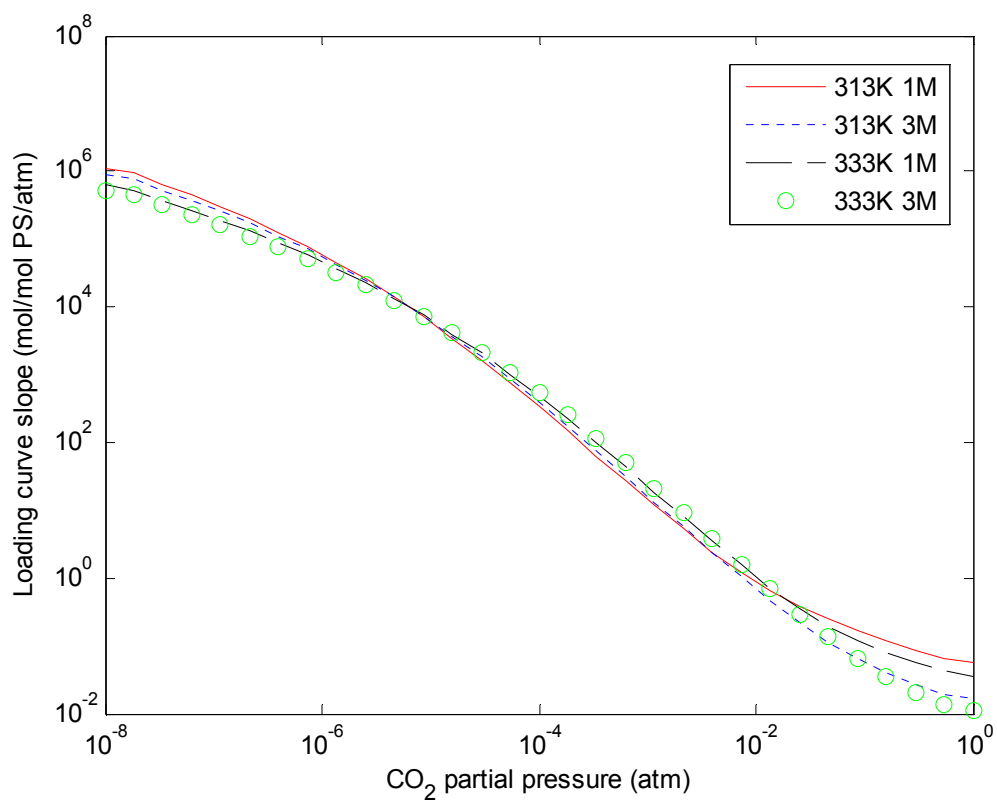


Figure 5.2.1.6 Loading vs. CO₂ partial pressure.

Figure 5.2.1.6 shows CO₂ loadings at the same temperatures. Lower temperatures favor CO₂ loading, which varies only slightly with amine concentration.

Figure 5.2.1.7 Loading curve slope vs. CO₂ partial pressure.

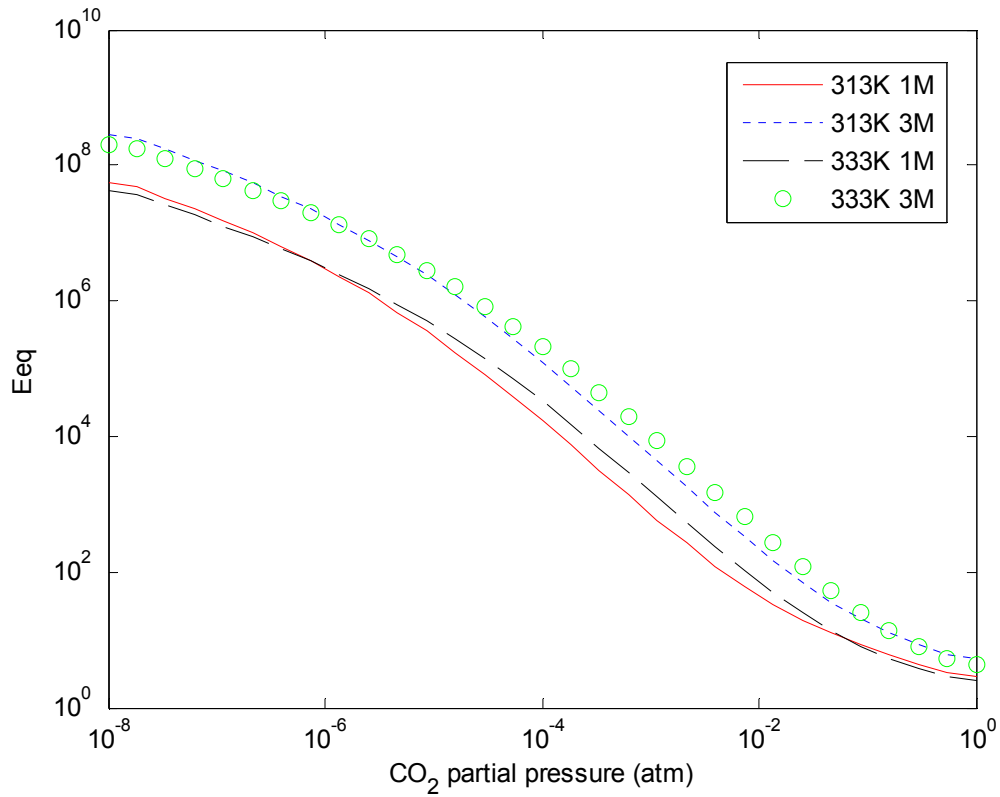


Figure 5.2.1.8 Equilibrium enhancement factor vs. CO₂ partial pressure

Figure 5.2.1.8 again indicates very large enhancement factors at very low CO₂ partial pressure, which even exceed those calculated for PG solutions. Furthermore, E is apparently more sensitive to PS concentration than temperature change.

5.2.2 Validation of perturbation analysis

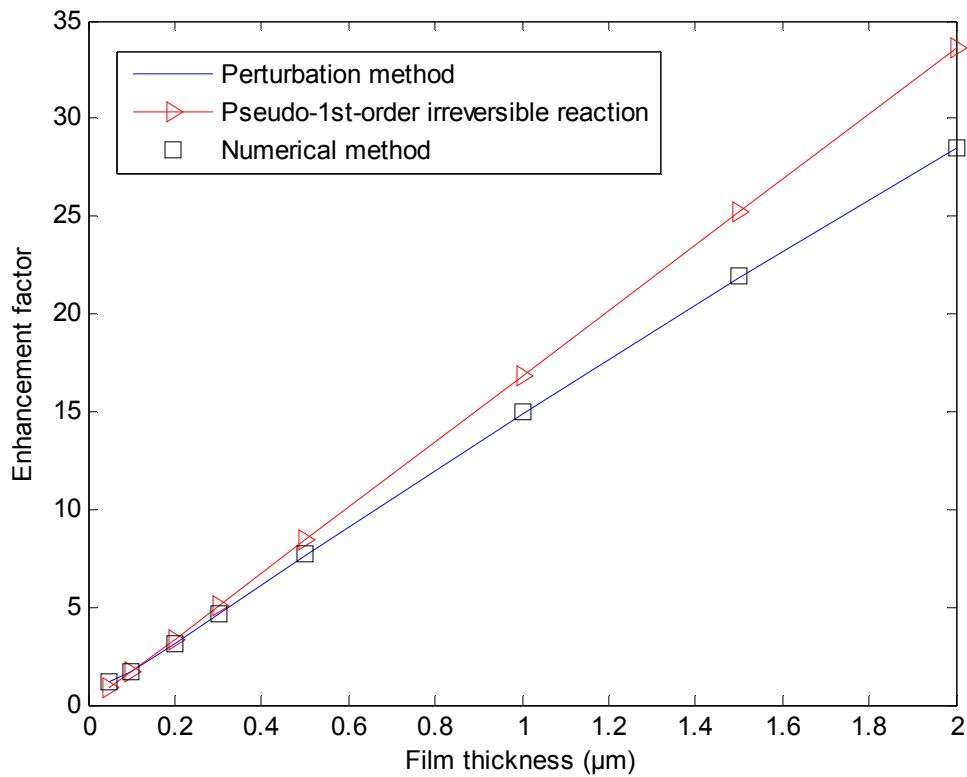


Figure 5.2.2 Absorption enhancement factor vs. liquid film thickness. (3 M PS, 313 K, bulk gas CO₂ partial pressure = 0.15 atm, bulk liquid loading = 40%)

Calculated enhancement factors are again in line with those obtained from numerical analysis. The rapid increase of E with film thickness reflects the very rapid reaction of PS with CO₂.

5.2.3 Results from perturbation analysis

5.2.3.1 Effect of concentration

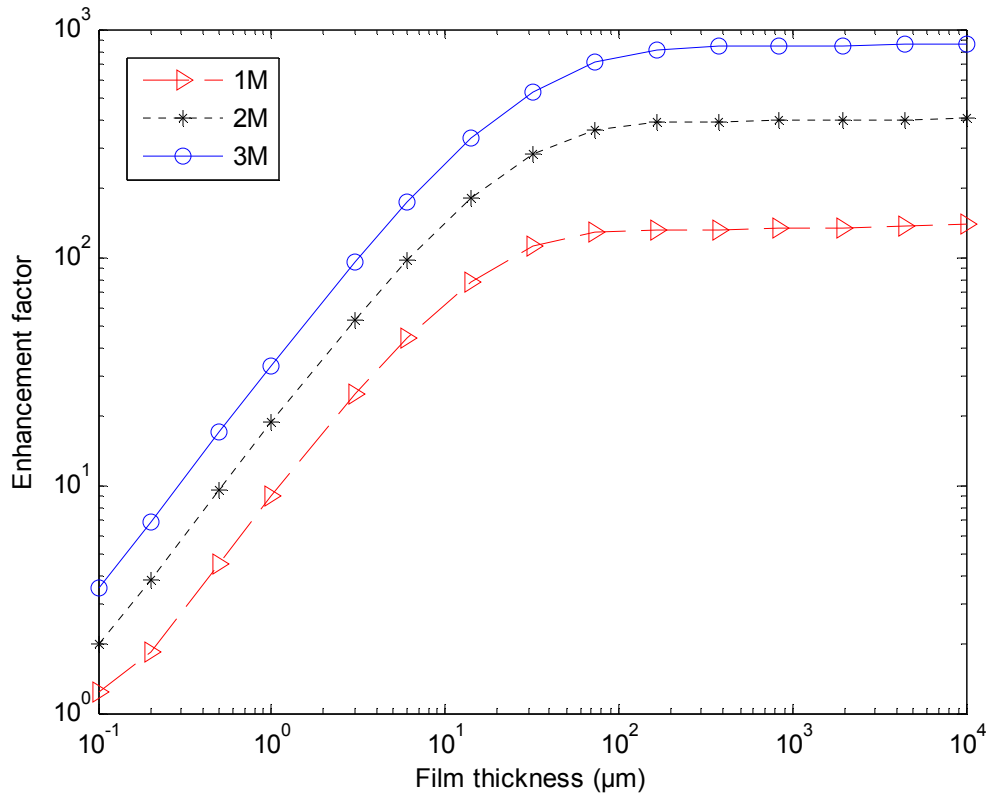


Figure 5.2.3.1 Absorption enhancement factor vs. liquid film thickness. (Bulk liquid loading = 10%, 313 K, bulk gas CO₂ partial pressure = 0.15 atm)

Figure 5.2.3.1 compares E values at three concentrations - 1, 2 and 3 M PS. The results are qualitatively similar to those obtained for PG solutions, although the E values are roughly double those for the latter; notably, E exceeds 1 even in very thin films.

5.2.3.2 Effect of bulk liquid loading

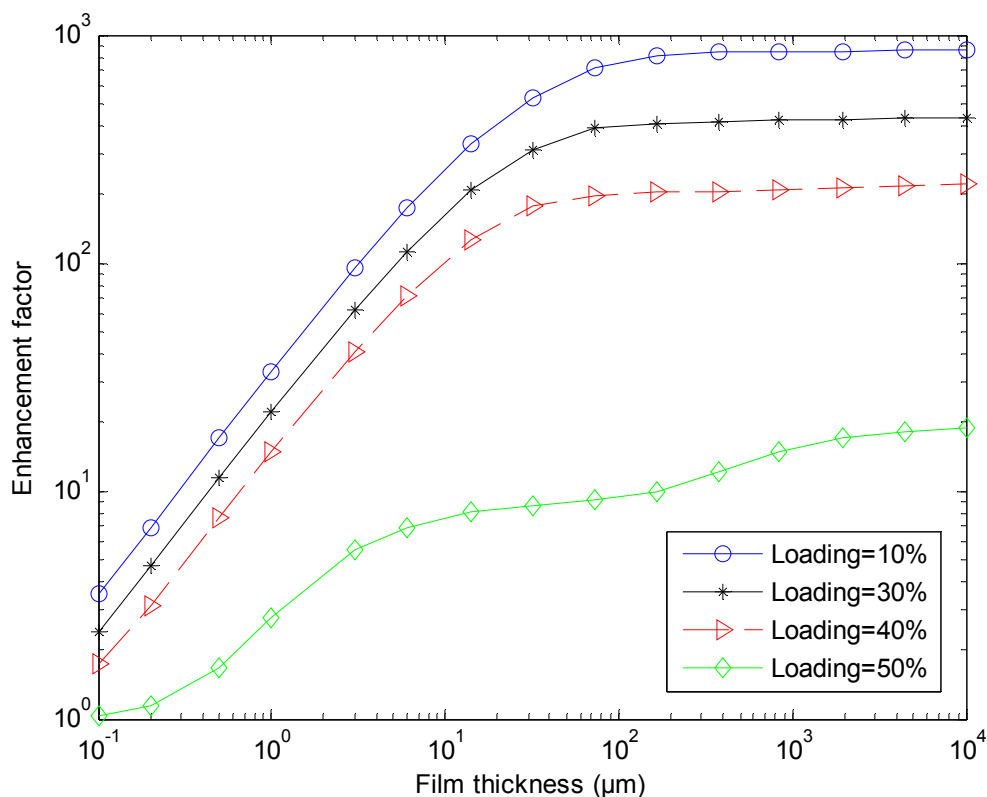


Figure 5.2.3.2 Absorption enhancement factor vs. liquid film thickness. (3 M PS, 313 K, bulk gas CO₂ partial pressure = 0.15 atm)

Calculated enhancement factors with various bulk liquid CO₂ loadings are shown in Figure 5.2.3.2, and the behavior is again similar to that observed for PG solutions. Notably, the results for 50% loading reflect the fact that the gas-phase CO₂ partial pressure only slightly exceeds that in equilibrium with bulk liquid (which is 0.05 atm). Thus, the reverse reaction markedly inhibits the absorption of CO₂.

6. CONCLUSION

The perturbation method efficiently models CO₂ absorption in PG and PS solutions. This bodes well for its general applicability to diffusion with multiple reversible reactions, including CO₂ absorption in solutions of mixed amines and amino acids; and its potential utility as an adjunct to scoping calculations for the design of coupled absorber-stripper systems.

NOMENCLATURE

A = CO₂

AAS = amino acid salt

b = salt-dependent constant

b₁ – b₄ = constants defined by equations 3.7.26, 3.7.30 and 3.7.31

B_j = base species

B⁻ = HCO₃⁻

c₁, c₂ = constants defined by equations 3.7.28-29

C_i, [i] = concentration for species i

C²⁻ = CO₃²⁻

D_A = diffusivity of CO₂ into aqueous amino acid salt solution, m² s⁻¹

D* = diffusivity of amino acid salt into aqueous solution, m² s⁻¹

E = enhancement factor defined by equation 3.5.11

G = defined by equation 3.8.5

H = 1/α, Henry constant, atm m³ mol⁻¹

Ha = Hatta number

i = component i

[i] = pseudo local equilibrium concentration for component i

Δ[i] = concentration departure from pseudo local equilibrium

I = ionic strength

J = defined by 3.8.5

k_1, k_{-1} = forward and reverse reaction rate constants of equation 2.2.4

k_{1a}, k_{-1a} = forward and reverse reaction rate constants of equation 3.1.1a

k_2, k_{-2} = forward and reverse reaction rate constants of equation 3.1.2

k_3, k_{-3} = forward and reverse reaction rate constants of equation 3.1.3

k_4, k_{-4} = forward and reverse reaction rate constants of equation 3.1.4

k_5, k_{-5} = forward and reverse reaction rate constants of equation 3.1.5

k_6, k_{-6} = forward and reverse reaction rate constants of equation 3.1.6

k_{Bj} = rate constant of protonated di-carbamate deprotonation by base

($B_j = W, OH^-, R^-$)

k_{ion} = ionic strength correction factor

k_L^o = CO_2 liquid phase mass transfer coefficient

K_{1a} = equilibrium constant for reaction 3.1.1a

K_{1j} = equilibrium constant for reaction 3.1.1

$K_{Acid,j}$ = reciprocal of acid dissociation constants

K_{Carb} = di-carbamate formation constant

K_2 = equilibrium constant for reaction 3.1.2

K_3 = equilibrium constant for reaction 3.1.3

K_4 = equilibrium constant for reaction 3.1.4

K_5 = equilibrium constant for reaction 3.1.5

K_6 = equilibrium constant for reaction 3.1.6

$K_{carbohydrolysis}$ = equilibrium constant defined by equations 4.1.4.5 and 5.1.4.2

L = CO₂ bulk liquid loading

$m_1 - m_4$ = constants defined by equations 3.7.22-23

MEA = monoethanolamine

ODE = ordinary differential equation

$P_{A,G}$ = CO₂ partial pressure in gas phase

PG = potassium glycinate

PS = potassium sarcosinate

Q = defined by 3.8.1

r = reaction rate, mol dm⁻³ s⁻¹

R = shorthand notation for protonated amino acid anion (a zwitterion)

R_g = gas constant, 8.314 J mol⁻¹ K⁻¹

R^- = shorthand notation for amino acid anion

R^{2-} = shorthand notation for di-carbamate anion

R^z = shorthand notation for protonated di-carbamate anion

R' , R'' = alkyl group

T = temperature, K

u_1, u_2 = constants defined by 3.7.22-23

W = water

x = position, defined in Figure 3.2, μm

$x_1 - x_4$ = defined by equations 3.8.7-10

y = dimensionless position

z = ion charge

GREEK LETTERS

β = square root of D^* over D_A defined by equation 3.5.11

δ = liquid film thickness, μm

Φ_1 = CO_2 absorption rate expressed as a flux defined by equation 3.5.7

Φ_2 = constant defined by equation 3.5.9

$\varepsilon_1, \varepsilon_2, E_1, E_2$ = constants defined by equations 3.7.22-23

$\eta_1 - \eta_4$ = constants defined by equations 3.7.19 and 3.7.21

$\theta_1 - \theta_5$ = constants defined by equations 3.7.42-46

λ_1, λ_2 = constants defined by equation 3.7.20

μ = viscosity, $\text{kg m}^{-1} \text{s}^{-1}$

ξ = defined by equation 3.8.1

ρ = density, kg m^{-3}

ρ_i = consumption rate for component i , $\text{mol dm}^{-3} \text{s}^{-1}$

$\omega_1 - \omega_6$ = constants defined by equations 3.7.16-18

SUPERSCRIPTS

eq = equilibrium

+ = pseudo-first-order irreversible reaction

SUBSCRIPTS

j = index

o = initial condition

∞ = infinite

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MAIN MATLAB CODE

The computer program shown below indicates potassium glycinate. For potassium sarcosinate, change the physiochemical parameters accordingly.

```

%%-----
Speciation Calculation:
%%-----
function F=speciation(x,A)
T=313;
C=3;% [PG] M
K=62.183098/T-0.111175;
Hco2w=exp(-2044/T)/(3.54*10^-7);
H=Hco2w*10^(C*K)*(1000/101325); % atm mol-1 dm3
K2=exp(-12092.1/T-36.7816*log(T)+235.482);
K4=exp(-12431.7/T-35.4819*log(T)+220.067);
K6=exp(-0.000237956*(T^2)+0.202203*T-61.6499);
Kcarb=K2/exp(-2767.18/T+6.10312);
% B=x(1)
% R=x(2)
% Rn=x(3)
% Rnn=x(4)
% C=x(5)
F=[x(2)+x(3)+x(4)-C;% [PG]
    x(1)+x(3)+2*x(4)+2*x(5)-C;% [PG]
    x(4).*x(2)-(Kcarb/K6).*A.*(x(3)^2);
    x(1).*x(2)-(K2/K6).*A.*x(3);
    x(5).*x(2)-(K4/K6).*x(3).*x(1)];

% Command window:
T=313;
C=3; % M [PG]
K=62.183098/T-0.111175;
Hco2w=exp(-2044/T)/(3.54*10^-7);
H=Hco2w*10^(C*K)*(1000/101325); %atm

x0=[0.02*C;0.4*C;0.3*C;0.3*C;0.04*C];
A=(1/H)*logspace(-5,1,31);
for i=1:length(A)
    AA=A(i);
    x=fsolve(@(x)speciation(x,AA),x0);

```

```

    x1(i)=x(1);
    x2(i)=x(2);
    x3(i)=x(3);
    x4(i)=x(4);
    x5(i)=x(5);
end

atm=logspace(-5,1,31);
Ad=A./C;
x1d=x1./C;
x2d=x2./C;
x3d=x3./C;
x4d=x4./C;
x5d=x5./C;
Loading=Ad+x1d+x4d+x5d;
semilogx(atm,Ad,'-m',atm,x1d,'-g',atm,x2d,'-r',atm,x3d,':b',atm,x4d,'-
k',atm,x5d,'ok',atm,Loading,'->k');
xlabel('CO_{2} partial pressure (atm)');
ylabel('Concentration (mol/mol PG)');
legend('A','B^{-}','R','R^{-}','R^{2-}','C^{2-}','Loading');

%%-----
Perturbation method:
%%-----
function F=speciationtry(x,A)
T=293;
C=1;% [PG] M
K2=exp(-12092.1/T-36.7816*log(T)+235.482);
K4=exp(-12431.7/T-35.4819*log(T)+220.067);
K6=exp(-0.000237956*(T^2)+0.202203*T-61.6499);
Kcarb=K2/exp(-2767.18/T+6.10312);
% B=x(1)
% R=x(2)
% Rn=x(3)
% Rnn=x(4)
% C=x(5)
F=[x(2)+x(3)+x(4)-C;% [PG]
    x(1)+x(3)+2*x(4)+2*x(5)-C;% [PG]
    x(4).*x(2)-(Kcarb/K6).*A.*(x(3)^2);
    x(1).*x(2)-(K2/K6).*A.*x(3);
    x(5).*x(2)-(K4/K6).*x(3).*x(1)];

```

```

function f=check(z)
% parameters
global delta
T=293;
C=1;
pA=0.15;
density=1056.57;
K=62.183098/T-0.111175;
Hco2w=exp(-2044/T)/(3.54*10^-7);
H=Hco2w*10^(C*K)*(1000/101325);
K2=exp(-12092.1/T-36.7816*log(T)+235.482);
K4=exp(-12431.7/T-35.4819*log(T)+220.067);
K6=exp(-0.000237956*(T^2)+0.202203*T-61.6499);
Kcarb=K2/exp(-2767.18/T+6.10312);

DA=2.35*100*(10^-6)*exp(-2119/T)*(1+0.2109*C+0.05124*C^2)^-0.48;
Dionic=(-2.412-9.403*(10^-3)*T+7.11*(10^-5)*(T^2)-0.2177*C-5.447*(10^-2)
)*(C^2)+1.296*(10^-3)*T*C*(10^-9)*100;
MW=113;
W=(density-C*MW)/18;
K5=exp(-13445.9/T-22.4773*log(T)+140.932);
K3=K2/K5;
k1a=2.81*(10^10)*exp(-5800/T)*1000;
kw=1.05*(10^-4)*exp(-1265/T)*1000;
kRn=4.89*(10^3)*exp(-5307/T)*1000;
k2=10^(329.850-110.541*log10(T)-17265.4/T);
k3=10^(13.635-2895/T);
AL=1.5480*10^-7;
BL=0.0038;
% RL=0.1096;
% RnL=0.8038;
RnnL=0.0866;
CL=0.0096;

global Bobar Robar Rnobar Rnnobar Cobar
% Bobar=x(1);
% Robar=x(2);
% Rnobar=x(3);
% Rnnobar=x(4);
% Cobar=x(5);

```

```

% one expression for phi_1
phiex1=(DA*(z-AL)+Dionic*(Bobar-BL+Rnnobar-RnnL+Cobar-CL))/delta;
% solve for constants
thetalmatrix=zeros(3,3);
thetalmatrix(1,1)=1+2*K4*Rnobar/(K6*Robar);
thetalmatrix(1,2)=1+2*K4*Bobar/(K6*Robar)+2*K4*Rnobar*Bobar/(K6*Robar^2
);
thetalmatrix(1,3)=2+2*K4*Rnobar*Bobar/(K6*Robar^2);
thetalmatrix(2,2)=Rnnobar+2*Kcarb*z*Rnobar/K6;
thetalmatrix(2,3)=Rnnobar-Robar;
thetalmatrix(3,1)=-Robar;
thetalmatrix(3,2)=Bobar+K2*z/K6;
thetalmatrix(3,3)=Bobar;
theta1=det(thetalmatrix);

theta2matrix=zeros(3,3);
theta2matrix(1,2)=1+2*K4*Bobar/(K6*Robar)+2*K4*Rnobar*Bobar/(K6*Robar^2
);
theta2matrix(1,3)=2+2*K4*Rnobar*Bobar/(K6*Robar^2);
theta2matrix(2,1)=-Kcarb*Rnobar^2/K6;
theta2matrix(2,2)=Rnnobar+2*Kcarb*z*Rnobar/K6;
theta2matrix(2,3)=Rnnobar-Robar;
theta2matrix(3,1)=-K2*Rnobar/K6;
theta2matrix(3,2)=Bobar+K2*z/K6;
theta2matrix(3,3)=Bobar;
theta2=det(theta2matrix);

theta3matrix=zeros(3,3);
theta3matrix(1,1)=1+2*K4*Rnobar/(K6*Robar);
theta3matrix(1,3)=2+2*K4*Rnobar*Bobar/(K6*Robar^2);
theta3matrix(2,2)=-Kcarb*Rnobar^2/K6;
theta3matrix(2,3)=Rnnobar-Robar;
theta3matrix(3,1)=-Robar;
theta3matrix(3,2)=-K2*Rnobar/K6;
theta3matrix(3,3)=Bobar;
theta3=det(theta3matrix);

theta4matrix=zeros(3,3);
theta4matrix(1,1)=1+2*K4*Rnobar/(K6*Robar);
theta4matrix(1,2)=1+2*K4*Bobar/(K6*Robar)+2*K4*Rnobar*Bobar/(K6*Robar^2

```

```

);
theta4matrix(2,2)=Rnnobar+2*Kcarb*z*Rnobar/K6;
theta4matrix(2,3)=-Kcarb*Rnobar^2/K6;
theta4matrix(3,1)=-Robar;
theta4matrix(3,2)=Bobar+K2*z/K6;
theta4matrix(3,3)=-K2*Rnobar/K6;
theta4=det(theta4matrix);

theta5=K4*Bobar*theta3/(K6*Robar)+K4*Rnobar*theta2/(K6*Robar)+K4*Rnobar
*Bobar*(theta3+theta4)/(K6*Robar^2);

MTA=zeros(3,3); % matrix template for deltas
MTA(1,2)=1;
MTA(1,3)=1;
MTA(2,1)=1+2*K4*Rnobar/(K6*Robar);
MTA(2,2)=-2*K4*Rnobar*Bobar/(K6*Robar^2);
MTA(2,3)=1+2*K4*Bobar/(K6*Robar);
MTA(3,1)=1+K4*Rnobar/(K6*Robar);
MTA(3,2)=-K4*Rnobar*Bobar/(K6*Robar^2);
MTA(3,3)=K4*Bobar/(K6*Robar);

MTB=zeros(3,2); % matrix template for deltas
MTB(1,1)=-1;
MTB(2,1)=-2;
MTB(3,1)=-1;
MTB(3,2)=-DA/Dionic;

DeltaMatrix=MTA\MTB;

omega1=DeltaMatrix(1,1);
omega2=DeltaMatrix(1,2);
omega3=DeltaMatrix(2,1);
omega4=DeltaMatrix(2,2);
omega5=DeltaMatrix(3,1);
omega6=DeltaMatrix(3,2);

lenda1=k2*omega5*z/Rnobar-k2*omega3*z/Robar+k3*K5*omega5*z/(K6*Robar)-k
3*omega3*Bobar/(K3*Robar)-k3*omega1/K3-k2*omega1*z/Bobar;
lenda2=k2-k3*omega2/K3-k2*omega2*z/Bobar-k2*omega4*z/Robar+k2*omega6*z/
Rnobar+k3*K5*Rnobar/(K6*Robar)+k3*K5*omega6*z/(K6*Robar)-k3*omega4*Boba
r/(K3*Robar);

```



```

eta1=(kla/Dionic)*((kRn*z*Rnobar^2+kw*W*z*Rnobar)/(1+kRn*Rnobar+kw*W))*
(1/Rnobar+omega3/Robar-2*omega5/Rnobar);
eta2=(kla/Dionic)*((kRn*z*Rnobar^2+kw*W*z*Rnobar)/(1+kRn*Rnobar+kw*W))*
(omega4/Robar-1/z-2*omega6/Rnobar);
eta3=(lenda1/DA)-(Dionic*eta1/DA);
eta4=(lenda2/DA)-(Dionic*eta2/DA);

u1=(eta1+eta4+sqrt((eta1+eta4)^2-4*(eta1*eta4-eta2*eta3)))/2;
u2=(eta1+eta4-sqrt((eta1+eta4)^2-4*(eta1*eta4-eta2*eta3)))/2;

m1=sqrt(u1);
% m2=-m1;
m3=sqrt(u2);
% m4=-m3;

E1=eta2/(u1-eta1);
E2=eta2/(u2-eta1);

c1=pA/H-z;

diffAobar=-thetal*phiex1/(thetal*DA+Dionic*(theta2+theta4+theta5));
c2=-theta4*phiex1/(thetal*DA+Dionic*(theta2+theta4+theta5));
%=diffRnobar
diffdeltaAo=-(m1*(E2*m3*c1-c2*tanh(m3*delta))+m3*(c2*tanh(m1*delta)-E1*
m1*c1))/(E2*m3*tanh(m1*delta)-E1*m1*tanh(m3*delta));

% second expression of phi_1
% phiex2=-DA*(diffAobar+diffdeltaAo);

% check if phiex1=phiex2
f=phiex1+(diffAobar+diffdeltaAo)*DA;

% Command window:
clear all
clc
global delta
delta=10000*10^-5; % dm
C=1;x0=[0.02*C;0.4*C;0.3*C;0.3*C;0.04*C];
A=linspace(0.0005122,0.0005136,100);
for i=1:length(A)

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```

AA=A(i);
x=fsolve(@(x)speciationtry(x,AA),x0);
x1(i)=x(1);
x2(i)=x(2);
x3(i)=x(3);
x4(i)=x(4);
x5(i)=x(5);
end

for k=1:length(A)
global Bobar Robar Rnobar Rnnobar Cobar
Bobar=x1(k);
Robar=x2(k);
Rnobar=x3(k);
Rnnobar=x4(k);
Cobar=x5(k);
z(k)=feval(@check,A(k));
end

ID=37; % find z(ID)==0, and change the index accordingly
z=A(ID);
Bobar=x1(ID);
Robar=x2(ID);
Rnobar=x3(ID);
Rnnobar=x4(ID);
Cobar=x5(ID);

% Follow the equations shown below to calculate the enhancement factor
b2=(E2*m3*c1*(1+exp(-2*m3*delta))-(1-exp(-2*m3*delta))*c2)/(E2*m3*(1+exp(-2*m3*delta))*(1-exp(-2*m1*delta))-E1*m1*(1+exp(-2*m1*delta))*(1-exp(-2*m3*delta)));
b4=((1-exp(-2*m1*delta))*c2-E1*m1*c1*(1+exp(-2*m1*delta)))/(E2*m3*(1+exp(-2*m3*delta))*(1-exp(-2*m1*delta))-E1*m1*(1+exp(-2*m1*delta))*(1-exp(-2*m3*delta)));

deltaA=((E2*m3*c1-c2*tanh(m3*delta))*tanh(m1*delta)+(c2*tanh(m1*delta)-E1*m1*c1)*tanh(m3*delta))/(E2*m3*tanh(m1*delta)-E1*m1*tanh(m3*delta));
deltaRnn=E1*b2*(1-exp(-2*m1*delta))+E2*b4*(1-exp(-2*m3*delta));

deltaB=omega1*deltaRnn+omega2*deltaA;
deltaR=omega3*deltaRnn+omega4*deltaA;

```

```

deltaRn=omega5*deltaRnn+omega6*deltaA;
deltaC=K4*Bobar*deltaRn/(K6*Robar)+K4*Rnobar*deltaB/(K6*Robar)-K4*Rnobar*Bobar*deltaR/(K6*Robar^2);

FA=z+deltaA;
FB=Bobar+deltaB;
FR=Robar+deltaR;
FRn=Rnobar+deltaRn;
FRnn=Rnnobar+deltaRnn;
FC=Cobar+deltaC;

EnhanceFactor=1+sqrt(Dionic/DA)*(FB-BL+FRnn-RnnL+FC-CL)/(pA/H-AL)

% Follow the equations shown below to calculate the concentration profiles
within the liquid film
function F=bsearch(x,y)
T=293;
C=3;% [PG] M
delta=6*10^-5;
phil=6.1141*10^-5; % 293K 3M L40 delta=6um
K2=exp(-12092.1/T-36.7816*log(T)+235.482);
K4=exp(-12431.7/T-35.4819*log(T)+220.067);
K6=exp(-0.000237956*(T^2)+0.202203*T-61.6499);
Kcarb=K2/exp(-2767.18/T+6.10312);
DA=2.35*100*(10^-6)*exp(-2119/T)*(1+0.2109*C+0.05124*C^2)^-0.48;
Dionic=(-2.412-9.403*(10^-3)*T+7.11*(10^-5)*(T^2)-0.2177*C-5.447*(10^-2)*C^2)+1.296*(10^-3)*T*C*(10^-9)*100;
AL=3.3037*10^-5;
BL=0.0605;
RnnL=1.1283;
CL=0.0113;
phi2=DA*AL+Dionic*(BL+RnnL+CL)+phil*delta;
% Bbar=x(1)
% Rbar=x(2)
% Rnbar=x(3)
% Rnnbar=x(4)
% Cbar=x(5)
% Abar=x(6)
F=[x(2)+x(3)+x(4)-C;% [PG]
x(1)+x(3)+2*x(4)+2*x(5)-C;% [PG]
x(4).*x(2)-(Kcarb/K6).*x(6).*(x(3)^2);

```

```

x(1).*x(2)-(K2/K6).*x(6)*x(3);
x(5).*x(2)-(K4/K6).*x(3).*x(1)
DA*x(6)+Dionic*(x(1)+x(4)+x(5))+phi1*y-phi2];

% Command Window:
x0=[0.0694;1.2503;0.5913;1.1585;0.0112;4.3679*10^-5];
y=linspace(0,delta,20);
for i=1:length(y)
    x=fsolve(@(x)bsearch(x,y(i)),x0);
    x1(i)=x(1);
    x2(i)=x(2);
    x3(i)=x(3);
    x4(i)=x(4);
    x5(i)=x(5);
    x6(i)=x(6);
end

for k=1:length(y)
b2=(E2*m3*c1*(1+exp(-2*m3*delta))-(1-exp(-2*m3*delta))*c2)/(E2*m3*(1+exp(-2*m3*delta))*(1-exp(-2*m1*delta))-E1*m1*(1+exp(-2*m1*delta))*(1-exp(-2*m3*delta)));
b4=((1-exp(-2*m1*delta))*c2-E1*m1*c1*(1+exp(-2*m1*delta)))/(E2*m3*(1+exp(-2*m3*delta))*(1-exp(-2*m1*delta))-E1*m1*(1+exp(-2*m1*delta))*(1-exp(-2*m3*delta)));

deltaA(k)=(E2*m3*c1-c2*tanh(m3*delta))*sinh(m1*(delta-y(k)))/(E2*m3*sinh(m1*delta)-E1*m1*tanh(m3*delta)*cosh(m1*delta)+(c2*tanh(m1*delta)-E1*m1*c1)*sinh(m3*(delta-y(k)))/(E2*m3*tanh(m1*delta)*cosh(m3*delta)-E1*m1*sinh(m3*delta)));
deltaRnn(k)=E1*b2*(exp(-m1*y(k))-exp(-2*m1*delta+m1*y(k)))+E2*b4*(exp(-m3*y(k))-exp(-2*m3*delta+m3*y(k)));

deltaB(k)=omega1*deltaRnn(k)+omega2*deltaA(k);
deltaR(k)=omega3*deltaRnn(k)+omega4*deltaA(k);
deltaRn(k)=omega5*deltaRnn(k)+omega6*deltaA(k);
deltaC(k)=K4*Bobar*deltaRn(k)/(K6*Robar)+K4*Rnobar*deltaB(k)/(K6*Robar)-K4*Rnobar*Bobar*deltaR(k)/(K6*Robar^2);
end

FA=x6+deltaA;
FB=x1+deltaB;

```

```

FR=x2+deltaR;
FRn=x3+deltaRn;
FRnn=x4+deltaRnn;
FC=x5+deltaC;
pH=-log10(K6*FR./FRn);

Y=y*10^5;
plot(Y,FA,'-m',Y,FB,'-g',Y,FR,'-r',Y,FRn,'--b',Y,FRnn,'-k',Y,FC,'ok');
xlabel('Film thickness (µm)');
ylabel('Concentration (mol/L)');
legend('A','B{-}','R','R{-}','R^{2-}','C^{2-}');

plot(Y,pH,'->b');
xlabel('Film thickness (µm)');
ylabel('pH');

%%-----
Pseudo-1st-order irreversible reaction:
%%-----
kforward=k1a*RnL*(kRn*RnL+kw*W)/(1+kRn*RnL+kw*W)+k2+k3*K2*RnL/(K3*K6*RL
);
Enforward=(delta*10^-5)*sqrt(kforward/DA);

%%-----
Numerical method:
%%-----
function dx=PGodefull(y,x,phi)
% parameters
delta=2*10^-5; % um
T=293;
C=3;
pA=0.15;
density=1163.85;
K=62.183098/T-0.111175;
Hco2w=exp(-2044/T)/(3.54*10^-7);
H=Hco2w*10^(C*K)*(1000/101325);
K2=exp(-12092.1/T-36.7816*log(T)+235.482);
K4=exp(-12431.7/T-35.4819*log(T)+220.067);
K6=exp(-0.000237956*(T^2)+0.202203*T-61.6499);
Kcarb=K2/exp(-2767.18/T+6.10312);

```

```

DA=2.35*100*(10^-6)*exp(-2119/T)*(1+0.2109*C+0.05124*C^2)^-0.48;
Dionic=(-2.412-9.403*(10^-3)*T+7.11*(10^-5)*(T^2)-0.2177*C-5.447*(10^-2
)*(C^2)+1.296*(10^-3)*T*C)*(10^-9)*100;
MW=113;
W=(density-C*MW)/18;
K5=exp(-13445.9/T-22.4773*log(T)+140.932);
K3=K2/K5;
k1a=2.81*(10^10)*exp(-5800/T)*1000;
kw=1.05*(10^-4)*exp(-1265/T)*1000;
kRn=4.89*(10^3)*exp(-5307/T)*1000;
k2=10^(329.850-110.541*log10(T)-17265.4/T);
k3=10^(13.635-2895/T);
AL=3.3037*10^-5;
BL=0.0605;
RL=1.2113;
RnL=0.6604;
RnnL=1.1283;
CL=0.0113;
kesi=DA*AL+Dionic*(BL+RnnL+CL);

Q=(phi.*delta.*(1-y)+kesi-DA.*x(1))./Dionic;
G=2.*x(3)-3.*Q-K4.*(C-2.*Q)./K6;
J=2.*Q.^2-3.*Q.*x(3)+(x(3).^2);

Bn=(-G-sqrt(G.^2-4*(1-K4/K6).*J))/(2.*(1-K4/K6));
Cnn=Q-x(3)-Bn;
Rn=C-2*Q+Bn;
R=2*Q-Bn-x(3);

% numerical method ODEs
dx=[x(2)

    delta.^2.*k1a.*x(1).*Rn.*(kRn*Rn+kw*W).*(1-K6*x(3)).*R./(Kcarb.*x(1).
    *Rn.^2))./(DA*(1+kRn*Rn+kw*W))+delta.^2*k2.*x(1)./DA-delta.^2*k2*K6.
    *Bn.*R./(DA*K2.*Rn)+delta.^2*k3*K5.*x(1).*Rn./(DA*K6.*R)-delta^2*k3.
    *Bn./(DA*K3)

    x(4)

    -delta.^2.*k1a.*x(1).*Rn.*(kRn*Rn+kw*W).*(1-K6*x(3)).*R./(Kcarb.*x(1)
    .*Rn.^2))./(Dionic*(1+kRn*Rn+kw*W))] ;

```

```

function res=PGbc(xa,xb,phi)
% parameters
delta=2e-5;
T=293;
C=3;
pA=0.15;
K=62.183098/T-0.111175;
Hco2w=exp(-2044/T)/(3.54*10^-7);
H=Hco2w*10^(C*K)*(1000/101325);
AL=3.3037*10^-5;
RnnL=1.1283;
DA=2.35*100*(10^-6)*exp(-2119/T)*(1+0.2109*C+0.05124*C^2)^-0.48;

% numerical method BCs
res=[xa(1)-pA/H;
     xa(2)+phi*delta/DA;
     xa(4);
     xb(1)-AL;
     xb(3)-RnnL];

% Command window:
phi=6.4498e-5; % initial guess of phi1
solinit=bvpinit(linspace(0,1,10000),[pA/H;-phi*delta/DA;1.1392;0],phi);
sol=bvp5c(@PGodefull,@PGbc,solinit);
y=linspace(0,1,10000);
x=deval(sol,y);

phires=-x(2,1)*DA/delta;

% calculate conc profiles
Q=(phires*delta*(1-y)+kesi-DA*x(1))/Dionic;
G=2*x(3)-3*Q-K4*(C-2*Q)/K6;
J=2*Q.^2-3*Q.*x(3)+(x(3).^2);

Bn=(-G-sqrt(G.^2-4*(1-K4/K6)*J))/(2*(1-K4/K6));
Cnn=Q-x(3)-Bn;
Rn=C-2*Q+Bn;
R=2*Q-Bn-x(3);
EnNum=1+sqrt(Dionic/DA)*(Bn(1)-BL+x(3,1)-RnnL+Cnn(1)-CL)/(pA/H-AL);

```

```
plot(delta,EnThickFilm,'-b',delta,Enforward,'-r>',delta,EnNum,'sk');  
xlabel('Film thickness ( $\mu\text{m}$ )');  
ylabel('Enhancement factor');  
legend('Perturbation method','Pseudo-1st-order irreversible  
reaction','Numerical method');
```